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6 RESULTS AND DISCUSSION

6.1 Text

6.1.1 Biochemical parameters

The long range goal of studies such as ~~this~~ one is to find biochemical markers for specific FLC types, e. g. macrophages, granulocytes and lymphocytes and/or subpopulations of a single FLC type, e. g. bacterio~~logica~~lly active macrophages. As classical biochemical methods generally require homogenization of tissue, the previous identification and separation of specific cell population is a prerequisite to the assignment of any marker to a given cell type.

Flow cytometric methods may one day replace classical biochemical methods by analyzing single cells in suspension individually or at least by sorting out specific subpopulations for further biochemical analysis. ~~With these thoughts in mind,~~ 3 classical biochemical methods were ~~also~~ included as parameters in this study. These methods are still under development and are especially limited by their determination in homogenates of the crude pools containing various cell types. As described in the following chapters on microscopic and flow cytometric methods. The short-term goal here is to find the detection limits and test for their general applicability in FLC studies.

6.1.1.1 FLC protein pattern in SDS-PAGE

The 1st problem in developing a small scaled electrophoresis method lies in the determination of small amounts of protein (.LT.1 microgram) in solutions containing substances which ~~can~~ cause artifacts or high blanks, e. g. SDS. This problem has now

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and groups

Large differences in the morphology of FLC in the various pools could be seen microscopically from cytocentrifuge preparations (see BC Figures 1 to 9). These effects appeared to be dependent on the lavage method and/or the type of smoke exposure. In the following, various methods have been tested for their usefulness as an objective and quantitative measure of these changes.

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been resolved by the adaption of the method of where protein is precipitated (Schaffner and Weissman, 1973), washed and determined photometrically by dye binding (see ... METHOD). With this method, protein can be determined from almost any solution in nanogram amounts.

The protein determinations indicate that 1E6 macrophages/milliliter yield approx. 0.1 milligram protein/milliliter (see BC TABLES ... and ...). Except for the increase due to granulocytes in the 1-GR, no pool dependent variations were found. A comparison of 1-GR and 2-GR to control, however, revealed an approx. 2-fold increase in protein for both groups relative to the 0-GR (see BC TABLE ..). At least part of the increase in the 1-GR can be explained by the presence of the large number of granulocytes, however, the increase for the 2-GR appears to be due neither to granulocyte or macrophage number nor an increase in macrophage size (see microscopic and flow cytometric parameters ...).

Due to the low concentration of FLC protein in samples after washing only a maximum of approx. 5 micrograms protein per slot could be applied to the gel. This amount of protein was too small to allow quantification of the smaller components of the protein pattern when stained with Coomassie Brilliant Blue (see BC TABLE .. and BC FIGURE ..). Using this method, a decrease in a protein of approx. 45000 daltons was found for MSS and SSS-exposed groups relative to control, while another protein of approx. 15000 daltons demonstrated an increase (see BC TABLE .. and BC FIGURE ..).

As the concentration of small samples is tedious and accompanied by large losses, the recently described silver staining method of Merril et al. (1981) was tested on these samples. This protein stain is 10 to 50-fold more sensitive than Coomassie Brilliant Blue (see BC FIGURE ..) and offered the possibility of quantitating some of the smaller FLC protein components. With this method the decrease in a protein at or about 45000 daltons was confirmed but the increase at or about 15000 daltons was not reproduced.

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The largest change seen after silver staining of FLC proteins was an increase in a protein of approx. 12000 daltons in treatment groups. This increase was 4-fold for 1-GR and 7-fold for 2-GR, and appears to correlate with a change in FLC protein pattern seen in a previous study (a). A small decrease in the amount of an approx. 16000-dalton component was also found for the 1-GR and 2-GR relative to the 0-GR, however changes in this molecular weight range could also be due to hemoglobin from contaminating erythrocytes (see flow cytometer differential counting).

From the above it appears that there are major changes in the protein pattern of FLC after smoke exposure, and silver staining increases the sensitivity of this method so that fractions of FLC subpopulations, i. e. 1E4 FLC, which could be obtained by sorting in a flow cytometer, might be studied.

6.1.1.2 FLC phospholipids and fatty acids

The evaluation of

Due to high blanks and low sensitivity of phospholipid determination based on an anorganic phosphorus assay, the amount of phospholipid in FLC could only be determined for 5 pools (see BC TABLE 9). This parameter also reflects the problem of reference point in crude FLC fractions. The number of macrophages as a reference do not allow for the contribution of granulocytes, while the smaller size of granulocytes is neglected when the number of FLC is taken as a reference. Thus protein appears to be the best reference point and should be determined routinely for all fractions in future studies. Despite the small number of data and large variations, an extreme elevation of phospholipid in FLC, as reported for chlorphentermine by phospholipidosis (Reasor, 1983) after smoke exposure would not appear probable. This ~~suggested finding~~

(a) see REFERENCES: INBIFO study A 0500/3056 (FLC substudy)

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is also in agreement with another smoke exposure study reported (De Lucia, 1982). With a detection limit of 5E6 macrophages, the determination of phospholipid using the methods described does not appear promising for future studies.

As a capillary gas chromatographic method for the analysis of fatty acid methyl esters (FAME) was available ~~in the analytical chemistry department~~ and this method is extremely sensitive (a), an effort was made to determine the fatty acid content and pattern in the remaining samples.

As a measure of phospholipid, the total integrated area of FAME per 1E6 FLC in each sample was determined. The mean of all weighted pools for this parameter in the 0-GR was 4.2 ± 0.6 (N = 6) units FAME total area per 1E6 FLC (see BC TABLES 10 and 11). The mean value was decreased in the 1-GR to 0.6-fold relative to the 0-GR, while the mean value for the 2-GR was similar to control, 1.1-fold. This decrease in the total amount of FAME found per FLC in the 1-GR, however, is probably mostly due to the smaller contribution of granulocytes per cell due to their smaller size.

In addition to total FAME the amounts of palmitic acid methyl ester (PAME) and stearic acid methyl ester (SAME) were determined. The mean value for PAME of weighted pools for 0-GR was 29.4 (N = 6) micrograms per 1E6 FLC (see BC TABLES 12 and 13). The same parameter for the 1-GR yielded a value of 23.0 or 0.8-fold of control (N = 3). The 1-GR also demonstrated an increasing trend with increasing number of cycles. This effect could be explained by the decrease in proportion of granulocytes, ~~while a similar trend seen for pools 5, 6 and 7 for the single determinations of 2-GR could not be explained by the same~~. The small number of data especially for 2-GR, however, do not allow a definitive interpretation of these effects.

(a) detection limit for total fatty acid methyl ester (FAME) pattern from .LT.1E3 macrophages

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The mean value for SAME of weighted pools for the 0-GR was 10.7 ± 0.4 (N = 5) micrograms per 1E6 FLC (see BC TABLES 14 and 15), or approx. 0.3-fold of PAME for control FLC. This parameter yielded values of 7.0 ± 0.4. (N = 4) and 10.9 ± ?. (N = 2) for the 1-GR and 2-GR respectively. ~~Thus there was no large difference seen between PAME and SAME for the various groups for the mean values.~~

For the ratio of PAME (16 : 0) to SAME (18 : 0) in the individual pools ~~, however,~~ there was a difference between early and late cycles (see BC TABLE 16). The mean ratio (16 : 0/18 : 0) for all groups for pools 1, 3 and 5 was 1.6-fold of that found for pools 2, 4, 6 and 7 (see BC TABLE 17). As the ratio (16 : 0/18 : 0) content of surfactant is very high in comparison to cellular membrane lipids (Spalding, 1983), this difference in FAME probably is related to the presence of surfactant in FLC either as non-specifically bound material on the cell surface, or as phagocytized material (Eckert, 1983). Thus despite the incomplete data set due to lack of material, FAME analyses of FLC appear to be of interest in future studies. (and BC Figure)

6.1.1.3 Acid phosphatase activity (ACP)

ACP activity of rat pulmonary macrophages has been reported to be increased by cigarette smoke exposure (Martin, 1973). A fluorometric method has been developed at INBIFO for this parameter (a), but no comparison of control and smoke-exposed FLC was made in this previous study; ~~Thus~~ therefore, this parameter was included in the present study.

Although this method is very sensitive, with a detection limit of approx. 2E4 FLC, the day to day variations were as much as 4-fold for the control (see BC TABLE 18). The mean ACP for all pools in the 3 experiments for 0-GR was 257 ± 41 units per 1E6

(a) see REFERENCES: INBIFO study A 0500/3052

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FLC. The same values for the 1-GR and 2-GR were 313 ± 42 and 547 ± 61 respectively (see BC TABLES 18 to 24). The mean ratio of treatment groups versus control group for each pool and experiment were also evaluated in an effort to compensate for the day to day variations (see BC TABLE 22). The mean ratio for 1-GR for all pools in the 3 experiments was 1.9 ± 0.8 relative to control, indicating at least some induction of ACP after exposure to MSS. The mean ratio for 2-GR for all pools in experiment 3 was 1.3 ± 0.2. In consideration of the 4-fold lower dose for SSβ these data could indicate that no extreme difference between SSβ and MSS should be expected in the induction of ACP. The confirmation of such an interpretation, however, will require more data at various doses.

6.1.2 FLC distribution and number

As hemocytometer counts from lavage medium have been shown in a previous study (a) to be inaccurate due to the low concentration of FLC, only cytocentrifuge preparations from lavage medium were made and evaluated for comparison to preparations from resuspension medium in this study.

Although the lack of absolute macrophage number did not allow the calculation of weighted means for the various lavage media, some information can be gained by the comparison of individual pools.

In cytocentrifuge preparation from lavage medium and resuspension medium, 0-GR FLC consisted of approx. 98 percent macrophages, 1 percent granulocytes and 1 percent lymphocytes (see BC TABLES 23 to 34 and BC FIGURES ..). A slight increase in the number of granulocytes in FLC of 0-GR was seen when PBS plus BSA with or

(a) see REFERENCES: INBIFO study P 0500/3097

2. without w
with
1% Rinderserum
albumin

Hier heißt sich "normal" und "INBIFO" Engl. gleich

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without PBS plus calcium and magnesium were used as lavage medium instead of PBS alone. This effect of lavage medium on the relative number of granulocytes in FLC was confirmed by the data for 2-GR. In this group, the relative number of granulocytes increased from approx. 13 percent in pool 1 to 31 percent in pool 3 and 22 percent in pool 5 for lavage medium (see BC TABLE 28 and BC FIGURE ..).

The respective values for relative number of granulocytes for resuspension medium were 7 percent, 16 percent and 10 percent (see BC TABLES 30 and BC FIGURE ..). A similar decrease in the relative number of granulocytes for 1-GR in pool 1 from 72 percent in lavage medium to 60 percent in resuspension medium indicate a specific loss of granulocytes during harvest when PBS alone was the lavage medium. Due to their lack of adherence, granulocytes were concentrated in the 1st 3 cycles, however, this effect was not increased as much as expected by the addition of calcium and magnesium to PBS (see BC TABLES 28 and 30 and BC FIGURES ..).

The relative number of lymphocytes was independent of group, pool and medium (see BC TABLES 31 to 34 and BC FIGURES ..).

The means of weighted pools for absolute number of macrophages in the 0-GR were 1.4 ± 0.3E6, 2.1 ± 0.3E6 and 3.3 ± 0.1E6 per rat for PBS, PBS plus BSA and PBS plus BSA prelavaged with PBS plus calcium and magnesium respectively. The respective values for the 1-GR were 1.0 ± 0.1E6, 2.1 ± 0.1E6 and 2.8 ± 0.5E6 (see BC TABLES 35 and 36 and BC FIGURE ..). Thus both sham and MSB-exposed rats demonstrated significant differences in the number of macrophages lavaged depending on the lavage medium used but independent of their treatment (see BC TABLES 35 and 36 and BC FIGURES ..). The pools 1 and 5 also demonstrated a significant decrease in the absolute number of lavaged macrophages in the 1-GR relative to 0-GR which could be interpreted as an increase in macrophage

There was however, no lavage medium dependent increase in the number of granulocytes for the 1-GR relative

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adherence due to MSS exposure. Similar effects on absolute macrophage number for 0-GR and 1-GR were also seen in a previous study (a). The decrease in absolute macrophage number also seen in the same study (a) for 2-GR, was reproduced in this study for PBS alone and PBS plus BSA prelavaged with PBS plus calcium and magnesium. The weighted means were 0.4E6 and 2.3E6 respectively. For PBS plus BSA without prelavage, this parameter for the 2-GR was elevated relative to 0-GR and 1-GR, i. e. weighted mean of 2.6E6 macrophages/rat.

For this as well as all other parameters, however, one must consider that these are single determinations in this study for SSS exposure, and can only be used as points of interest in future studies.

Another interesting observation on absolute macrophage number was that despite the unexpectedly large number of macrophages removed by prelavage with PBS plus calcium and magnesium, the number of macrophages recovered in the following 10 cycles with PBS plus BSA was increased for 0-GR and 1-GR relative to the same number of cycles with PBS plus BSA without prelavage (see BC FIGURE ..).

The determination of absolute granulocyte number in a hemocytometer with unstained viable cells has been shown in a previous study (a) to be extremly inaccurate. Thus this determination was not performed in this study. Changes in the relative number of cell populations, however, are interdependent and can be misleading or at best difficult to interpret. Therefore, the absolute number of granulocytes was calculated from relative values for macrophages and granulocytes in cytocentrifuge preparations and the absolute number of macrophages in resuspension medium (see BC TABLES 37 and 38 and BC FIGURE ,.). Both the larger relative

(a) see REFERENCES: INBIFO study P 0500/3097

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number of granulocytes in early cycles and the decrease in relative number of granulocytes recovered with PBS were confirmed by calculated absolute numbers. Further, more granulocytes were recovered by 3 cycles of PBS plus calcium and magnesium as in 10 cycles with PBS alone.

6.1.3 Viability of macrophages

The viability of FLC determined immediately after harvest by dye exclusion appeared to be good, 94 percent or more viable macrophages (see BC TABLES 39 and 40 and BC FIGURES ..). Closer observation, however, revealed that pool 1 of experiment 1 for 0-GR and 1-GR were the lowest values found (see BC TABLE 39). The means of pools (see BC TABLE 40) also revealed a decrease in viability for

(1) early versus late cycles,

(2) smoke-exposed versus sham control groups

and

(3) PBS versus other lavage media.

These differences, however, were too small to be convincing despite the small variation seen (see BC FIGURE ..). The calculation of the absolute number of nonviable macrophages per rat, however, lended support to the significance of these observations (see BC TABLE 41 and BC FIGURE ..). The only exception to this statement was the smaller number of nonviable macrophages found for 2-GR lavaged with PBS. This discrepancy is probably related to the extremely low number of macrophages recovered in this group. The problems associated with this "acute" parameter have been discussed in detail in a previous report (a).

(a) see REFERENCES; INBIFO study P 0500/3097

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6.1.4 Number of multinucleated macrophages (microscopic method)

The number of multinucleated macrophages has been reported to be increased by smoke exposure in a previous study (a), therefore this parameter was recorded simultaneously during differential counting. Although the apparent variation in this parameter was small, the microscopic data should be considered preliminary as only a maximum of 500 macrophages were examined and the relative number of multinucleated cells was very small.

This parameter demonstrated no pool or lavage medium dependent variations. The mean of weighted pools for relative number of multinucleated macrophages in all 3 lavage media for 0-GR was 1.1 percent ± 0.1 (N = 8). This value for the 1-GR, however, was increased to 5.3 percent ± 0.9 (N = 8), while the 2-GR was intermediate, 2.8 percent ± 0.3 (N = 3) (see BC TABLES 43 and 44 and BC FIGURES ..).

The mean absolute number of multinucleated macrophages also confirmed this ranking. The values were 19E3 ± 5E3, 103E3 ± 11E3 and 48E3 ± 20E3 for the 0-GR, 1-GR and 2-GR respectively (see BC TABLES 45 and 46, BC FIGURES ....). The variation, however, was larger for the absolute values. This is largely due to the large difference in macrophage number between the various lavage media.

6.1.5 Morphometry of macrophages

As changes in macrophage size and vacuolization have been reported to increase after smoke exposure (~~Cenix~~, ...), the area of macrophages, nuclei and vacuoles was determined planimetrically from

(a) see REFERENCES: INBIFO study A 0500/3016

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microphotographic slides of stained cytocentrifuge preparations in this study.

The changes in macrophage morphology are easily observed by microscopic examination of cytocentrifuge preparations (see BC FIGURES 1. to 8...). The quantification of these parameters, however, is difficult, and very much dependent on the quality of the cell preparation. In agreement with viability data (see also flow cytometer method) cell preparations from lavage with PBS alone were of such poor quality that no morphometry could be made (see BC FIGURES 1a.02).

The mean macrophage area of weighted pools from lavage with PBS plus BSA without and with prelavage were 210 square micrometers (N = 240) and 224 square micrometers (N = 469) respectively. The respective values for 1-GR were 365 square micrometers (N = 397), and for 2-GR, 259 square micrometers (N = 190) and 275 square micrometers (N = 230) (see BC TABLE 47). The difference for macrophage size between lavage media for 1-GR and the increase with cycle number for 2-GR correlated with an increase in nuclear area (see BC TABLES 47 and 49) and is considered to be due to unspecific swelling during handeling and/or cytocentrifuge procedure. The increase found in smoke-exposed groups was too large to be explained by these effects and has also been confirmed by flow cytometry.

The main change in macrophage morphology after smoke exposure was the increase in vacuolization. The mean vacuole area of weighted pools from lavage with PBS plus BSA without and with prelavage were 0.6 square micrometers (N = 493) and 0.8 square micrometers (N = 1433) respectively (see BC TABLE 51 and BC FIGURE ...). The respective values for 1-GR were 2,2 square micrometers (N = 1038) and 2.3 square micrometers (N = 2840). This is equivalent to a 3- to 4-fold increase in mean vacuole area relative to the control. The increase in mean vacuole area for the 2-GR was

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much smaller than for 1-GR with a maximum of 1.7-fold relative to the control (see BC TABLE 51), however, the vacuole area per macrophage was increased considerably, 2 to 3-fold relative to the control (see BC TABLE 52, BC FIGURE .....). Thus SSS exposure increased the number but not so much the size of vacuoles. The increase in percent of macrophage area covered by vacuoles in the 1-GR was even larger, 5 to 6-fold of control, than the increase in size of vacuoles (see BC TABLE 53). Thus MSS exposure increased the number and the size of vacuoles.

6.1.6 Flow cytometry parameters

6.1.6.1 Evaluation Schemes

6.1.6.1.1 Viability and esterase assay

FLC from smoke-exposed rats mainly consist of macrophages and granulocytes, which were found to be resolved to a limited degree by FWD (signal area) and AXL (signal height) (see BC FIGURE ..). Enhanced resolution was, in some instances, achieved when the 2 parameters were correlated in a cytogram of FWD versus AXL (see (2), (3), and (6), BC FIGURE .. and BC FIGURE ..).

The 4 parameters involved in the viability and esterase assay, FWD, AXL, red and green fluorescence, may be evaluated in different ways. The simplest method of differentiating between cell types using FWD and AXL (see (1), (2) and (5), BC FIGURE .. and BC FIGURE ..) followed by separation of viable and nonviable cells in the FWD versus AXL region (see (3), (4) and (6), BC FIGURE ..) was found to be inappropriate as nonviable cells were not exclusively in the same region as viable cells. Therefore, all signals were used to create a green versus red fluorescence cytogram (see (6), BC FIGURE .. and BC FIGURE ..). A FWD versus AXL cytogram (7) was then created only from cells in a specified region of the green versus red fluorescence (6). This cytogram

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therefore displayed viable, intermediate (acutely damaged), or nonviable cells only (see BC FIGURE ..). Nonviable cells were defined by negligible green fluorescence and red fluorescence corresponding to at least the DNA content of granulocytes. As the precise location of granulocyte DNA was difficult in (6), the red fluorescence signals in region ... of (6) were also gated (a) in a histogram (8) (see BC FIGURE ..). For the evaluation of intermediate cells, the FWD versus AXL cytogram (7) was created for signals which were not in region .... and .... of (6) (see BC FIGURE ..). The number of nonviable or intermediate cells of the selected subpopulation was counted by setting appropriate regions in (7) and related to the total number of the respective cell type as counted in (5). ~~Using~~ The use of the FWD versus AXL cytogram (5) for the determination of total cells may result in a slight underestimation of this value for the reason given above, but the error was considered to be negligible, at least for macrophages. Nonviable granulocytes were found to be in a completely different location in (7) as compared to the majority of granulocytes seen in (5) (see BC FIGURE ..). Due to difficulties in finding their correct position, especially results obtained for the 0-GR are considered to be unreliable. Therefore, results on viability will be presented for macrophages only.

An additional ~~way~~ scheme of looking at viability was employed for some samples by creating a FWD or AXL histogram of total macrophages from (5), selecting viable, intermediate or nonviable cells in (6) and (7) and creating FWD or AXL histograms of selected macrophages ((9) or (10)). The histograms of selected and total macrophages were then divided ~~by each other~~ channel by channel, (12). This procedure results in a histogram displaying the relative number of ~~e.g.~~ viable cells for example as a function of FWD or AXL, which is equivalent to the distribution of viability in the corresponding

(a) i. e. selected by setting a region

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FWD or AXL-histogram of total macrophages (see BC FIGURE ..). In this example it is seen that the relative number of viable macrophages is highest in the center of the AXL distribution and levels off at the ascending and descending parts of the peak. This is in accordance with a broader distribution seen for non-viable as compared to viable macrophages.

The esterase activity of FLC was evaluated by gating on viable macrophages or granulocytes only ((5) and (6)), and calculating the mean green fluorescence of the selected cells (a). The green fluorescence signals were also divided by the corresponding AXL signals on a cell by cell basis (13) in order to obtain some information about the contribution of cell size to esterase activity, which in (11) is only expressed as mean total fluorescein content per cell.

For the viability and esterase determinations in experiment 1 an attempt was made to measure the kinetics of esterase activity continuously for 20 minutes. Due to the limited amount of sample determinations could only be made for a maximum of 16 minutes in most samples. Data for viability were therefore evaluated for the period of 10 to 15 minutes. For the determinations in experiment 3, the method was simplified by preincubation of FLC for 15 minutes and subsequent determination for the period of 15 to 20 minutes. This modification showed no influence on viability data, i. e. the values for viability were constant after approx. 5 minutes of incubation (data not shown). Furthermore, as experiment 1 was performed after only cursory experience with the application of the viability and esterase assay to FLC, readjustment of the FCM was necessary due to the occurrence of atypical samples. Therefore, no valid data are available on esterase activity (b) and light

(a) only macrophages
(b) no suitable fluorescence standard available for calibration

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scatter for this experiment. FCM readjustment does not affect the viability data and differential counts. No FCM viability and esterase assay was made in experiment 2 due to laser failure.

For experiment 3, FWD and AXL of macrophages ~~was~~ were evaluated with respect to mean value and CV in histograms created by gating on viable macrophages only (see BC FIGURE .. and .....).

Differential counts were made by setting regions in (5) and, in addition, using the FWD and AXL histograms of viable macrophages (see (7) and (8), BC FIGURE .. and BC FIGURE ..).

6.1.6.1.2 Phagocytosis assay

The determination of phagocytic activity after incubation of FLC with fluorescent latex beads basically involves 2 steps: (1) differentiation between FLC subpopulations and (2) quantification of cells with respect to the number of phagocytized beads.

Green fluorescence of free and phagocytized latex beads which, by ~~itself~~, does not provide useful information (see BC FIGURE ..), could thus be gated for the desired subpopulations by setting appropriate regions. Good resolution was obtained in this way for nonphagocytizing and phagocytizing granulocytes, but not for macrophages (see BC FIGURE ..). The reason for this is readily seen as a tilting of the macrophage distribution in a cytogram of green fluorescence versus AXL, revealing high autofluorescence of macrophages superimposed on the fluorescence of phagocytized latex beads (see BC FIGURE ..). This tilt was compensated by software using the approximately linear relation between AXL and green fluorescence in order to optimize the resolution for phagocytizing macrophages (see BC FIGURE .. and (5) BC FIGURE ..). Compensated fluorescence

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histograms were used for the evaluation of macrophages while compensation was not necessary for the evaluation of granulocytes. Residual autofluorescence was subtracted for the calculation of the mean number of beads per cell.

Cellular autofluorescence may be due to intrinsic cell components such as lipofuscin (.....), but may also be induced by the fixative glutaraldehyde (a). Autofluorescence has been reported for unfixed FLC from rabbits (Shellito, ...) and was also used for the differentiation of FLC subpopulations in rats (Steinkamp, ...). As this parameter may be interesting for future studies, data obtained from nonphagocytizing macrophages at time = 0 of incubation will be reported (see (8) and (12), BC FIGURE ..).

For samples taken immediately after addition of latex to FLC suspension, FWD, AXL and RAS were evaluated from histograms (9) and (10), BC FIGURE .., obtained by gating on macrophages with no latex beads in (6) and (7), see BC FIGURE ...

The CV of the FWD and AXL distributions will be reported only if no more than 1 percent of signals were out of range. The corresponding mean values were considered to be underestimated by less than 2 percent when up to 5 percent of signals were out of range (b). This error was tolerated. The distribution of RAS signals was extemely broad, especially in the 1-GR (see BC FIGURE ..). An appropriate analysis of FLC would require logarithmic amplifcation covering approx. 3 decades. Therefore, RAS of macrophages was evaluated by defining a threshold corresponding to 14 times the

(a) Glutaraldehyde induced fluorescence is, in a narrow sense, not autofluorescence, the term autofluorescence will, however, be used subsequently for the sake of simplicity.
(b) control made by simulation

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position of standard latex beads. RAS was then expressed as the relative number of macrophages exceeding this threshold.

Differential counts were made by setting regions in the FWD versus AXL cytogram (6) (see BC FIGURE ..).

6.1.6.1.3 DNA assay

For the present study, a 1st attempt was made at INBIFO to determine the distribution of FLC in the cell cycle phases G0/1, S and G2 (a) by FCM. An analytical technique was chosen which permits the differentiation between mononuclear cells and cells or cell aggregates containing 2 or more nuclei (b). This technique is usually applied in order to avoid interference of cell aggregates with the DNA distribution of mononuclear cells. The principle is based on the relatively high spatial resolution (c) of the FCM employed in this study which allows to draw information from the shape of the scatter or fluorescence signals produced by cells. Thus, the fluorescence signal produced by G2 cells stained for DNA is 2 times as large as for G0/1 cells (d) both with respect to height (measure of maximal local fluorescence density) and area (total cellular fluorescence) (see BC FIGURE ,.). Doublets or binuclear cells give signals with the same area as for G2 cells, as they have the same DNA content. The signal height, however, will be lower in most instances, as the DNA is shared by 2 nuclei which only accidentally pass the laser beam exactly one behind the other along the beam axis. These cells are therefore generally different in the relation between signal area and height.

(a) G0/1: G0 and/or G1, resting or potentially cycling cells
S: cells synthetizing DNA
G2: cells with 1 nucleus containing 2 x G0/1 DNA, capable of dividing into 2 G0/1 cells
(b) standard software provided by Ortho, modified
(c) laser focus approx. 5 micrometer at 1/e2 points (approx. 1/15 percent intensity)
(d) equal nuclear size assumed in this example for the sake of simplicity

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As in all other FCM assays the 1st step was the separation of macrophages and granulocytes by FWD and AXL (see BC FIGURE .. and ..). A cytogram of red fluorescence signal height versus area (6) was constructed for cells gated in (5), BC FIGURE .., (see BC FIGURE .. and ..). Regions were set in (6) in order to count the number of signals for G0/1, S etc.. In addition, histograms of signal heigt (8) and area (9) were created for cells gated in (5) and (6) for the determination of signal mean and CV (see BC FIGURE ..). In some instances, a secondary FWD versus AXL cytogram (7) was made for cells gated in (5) and (6) in order to ~~find the~~ location ~~of~~ nucleated cells or cells with a specific DNA content (see BC FIGURE ..).

No light scatter parameters were evaluated for ethanol fixed FLC as ethanol is a poor fixative with respect to cell morphology. This was also seen in the lower resolution of macrophages and granulocytes as compared to unfixed or glutaraldehyde fixed cells (see BC FIGURE ..).

Differential counts of macrophages were made fom the FWD versus AXL cytogram (5) and, in addition, by counting signals corresponding to at least G0/1 DNA content in (6) (see BC FIGURE ..).

6.1.6.2 Experimental results

6.1.6.2.1 Viability and esterase activity

In experiment 1 the relative number of viable macrophages obtained by lavage with PBS (pool 1 and 2) was extremely low, i. e. 51 and 37 percent in the 0-GR, and 51 and 57 percent in the 1-GR (see BC TABLE .. and BC FIGURE ..). These samples contained high numbers of both intermediate and nonviable macrophages and were also characterized by an appreciable increase in FWD as well as the

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appearance of a subpopulation of macrophages with low AXL (see BC FIGURE ..). As can be seen from the distribution of viability in the AXL histogram (0-GR, pool 2), the low AXL subpopulation mainly consists of intermediate macrophages, while in the high AXL subpopulation approx. 50 to 60 percent of macrophages are viable. These data indicate extensive acute cell damage in the PBS samples, which also had the lowest viability in the trypan blue assay (see BC TABLE ..), although the viability seen with this assay was generally much higher. Due to differences in all handling and time between lavage and determination (a), a direct comparison of the results from the 2 viability assays is, however, not possible. For pool 3 and 4 (PBS plus BSA) the relative number of viable macrophages was 96 and 93 percent in the 0-GR, and 83 and 88 percent in the 1-GR, respectively (see BC TABLE ..). The value for pool 5 was only 52 percent in the 1-GR, increasing to 85 percent for pool 7, while the corresponding pools in the 0-GR were 90 to 94 percent.

In experiment 3 the viability of macrophages was generally higher and there were no samples with extreme acute cell damage as in experiment 1. There seemed to be a tendency for lower viability in pool 1 and 2 for the 0-GR and the 1-GR, but not for the 2-GR, as compared to pools 3 to 7. For the 1-GR, the viability was consistently lower than the respective pools for the 0-GR. The lowest values for viability were found for the 2-GR, pool 3 to 7. In these pools there was evidence for acute cell damage (see BC TABLE .. and BC FIGURE ..).

Comparing the viability of total lavaged macrophages, i. e. the weighted mean of pools, there seems to be a tendency for lower values for PBS as compared to the other lavage media, with the exception of the 2-GR (see BC TABLE .. and BC FIGURE ..). The viability of macrophages in the 1-GR was almost invariably lower

(a) up to 9 hours for FCM in experiment 1, up to 3 hours for trypan blue assay

and might be associated with the [illegible] number of pool 1 and

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than in the 0-GR, comparing corresponding mean pools and experiments. The lowest values in experiment 3 were found for the 2-GR, pool 3 and 4 and 5, 6 and 7.

The viability data obtained by FCM indicate that there were obviously problems with cell handling and processing in the present study, which may more or less be enhanced by the different lavage media. Due to the large variability and the irregular occurrence of high number of intermediate, i. e. acutely damaged cells, these data do not lend themselves easily for an interpretation of differences between groups and/or lavage media. These results, however, demonstrate the potential of the FCM viability assay for revealing the presence of such problems thus providing a basis for the optimization and standardization of methods to be used in future FLC studies.

EXP3 ?

DRKT (R) Mue

The distribution of green fluorescence signals from macrophages, corresponding to esterase activity, generally resembled a log-normal distribution (see BC FIGURE ..). In some samples, an additional small peak was seen at the very ascending portion of the distribution. This peak was excluded from evaluation and not further examined. The esterase activity of macrophages, determined as mean green fluorescence per viable macrophage, was consistently higher in the 1-GR than in the 0-GR. The difference of 16 to 52 percent between these groups may be statistically significant. The values for the 2-GR were very close to those for the 0-GR (see BC TABLE .. and .., and BC FIGURE ..). The distribution of green fluorescence signals was broader for both 1-GR and 2-GR as compared to the 0-GR. There did not seem to be a systematic difference between the pools of each lavage group.

For cells which are identical with respect to the parameters determining the kinetics for the conversion of FDA, i. e. enzyme concentration and membrane permeability, but which differ in size,

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one can expect the amount of accumulated product to increase with size. Hence, the fluorescence per cell would also increase. The difference between the 0-GR and the 1-GR may therefore merely result as a consequence of increased size of macrophages in the 1-GR (see BC TABLE ..). Using AXL as a measure of cell size (a), the ratio of green fluorescence/AXL was calculated on a cell by cell basis in order to compensate, at least in part, for the difference in macrophage size. The resulting distributions were less broad and the mean value for the 1-GR was only slightly larger than for the 0-GR (see BC TABLE .., .., and .., and BC FIGURE ..). For the 2-GR there seemed to be a decrease compared to the 0-GR. As AXL is assumed to be less than proportional to cell volume, the actual esterase activity of macrophages in both 1-GR and 2-GR may be lower than in the 0-GR.

The granulocytes were not evaluated quantitatively for their mean esterase activity, as the fluorescence distributions were found to be bimodal (see BC FIGURE ..). The majority of granulocytes showed very low esterase activity. There was, however, a 2nd population with approx. 10-fold higher esterase activity, which was located precisely at the same position as the low-esterase granulocytes in the FWD versus AXL cytogram. These high-esterase granulocytes can be shown not to represent spill over from the adjacent macrophages. As the 2 suppopulations were not clearly separable, only a crude evaluation was made in order to estimate their relative contribution to total granulocytes (data not shown). The relative number of high-esterase granulocytes seemed to be highest in the 0-GR (approx. 20 to 45 percent) and lowest in the 1-GR (approx. 10 to 20 percent). In addition, there seemed to be an increase in this subpopulation in the later cycles of each lavage. This finding can, at present, not be interpreted in terms of known granulocyte subclasses, but demonstrates the potential of FCM for the discrimination of subpopulations by means of biochemical and functional criteria.

(a) discussed in ...

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6.1.6.2.2 Phagocytic activity

The investigation of the influence of lavage medium on pagocytic activity of FLC was hampered by the low yield in cell number for pool 8 (PBS). Thus only 2 samples (0-GR) were available for the phagocytosis assay. In addition, the sample determined in experiment 1 displayed an atypical bimodal FWD versus AXL distribution for macrophages presumably indicating extensive cell damaging. 27 percent of macrophages with approximately normal light scatter properties were found to have phagocytized at least 1 bead, while a value of 15 percent was found for the total population. 31 percent of macrophages in pool 8 were phagocytizing in experiment 2 (see BC TABLE ..). This value is within the range of those for the 0-GR, pool 9 and 10. The mean number of phagocytizing macrophages in these pools was 31 and 35 percent, respectively (see BC TABLE .. and ..). Using similar methods Shellito et al. (1981) found 80 percent of macrophages from untreated rabbits to be phagocytizing. This discrepancy may be due to a variety of methodological factors including the surface properties of the latex particles employed and/or species &

The mean number of phagocytizing macrophages increased to 54 and 58 percent in the 1-GR, pool 9 and 10, respectively. 1 sample each of pool 9 and 10 of the 2-GR was available in experiment 3 yielding values of 54 and 50 percent. This method however does not differentiate between externally adherent and internalized beads. The mean number of macrophages phagocytizing 3 or more beads was substantially lower in all groups (see BC TABLE ..). However, the relative difference between the groups was larger for these values. This finding may indicate that there were not only more phagocytizing macrophages in the 1-GR and the 2-GR, but that they were also more active. The increase in the mean number of phagocytized beads per macrophage from approx. 2 in the 0-GR to 4.3 and 4.5 in the 1-GR, pool 9 and 10, seems to confirm this interpretation (see BC TABLES .. and .., and BC FIGURE ..).

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The relative number of phagocytizing granulocytes as compared to the macrophages was lower in the 0-GR, similar in the 1-GR, and slightly higher in the 2-GR (see BC TABLE .. to ..). For 3 or more beads per granulocyte there seemed to be a slight increase in the difference between the 1-GR and the 2-GR and a decrease between the 0-GR and the 1-GR (see BC TABLE .. and BC FIGURE). The mean number of beads per granulocyte was extremely variable in the 0-GR, ranging from 0.2 to 2.7, while the values for thte 1-GR were less variable, ranging from 2.7 to 3.6. The values for the 2-GR were similar to those of the 1-GR (see BC TABLE ..). The phagocytic activity of granulocytes generally seemed to be higher in pool 10 as compared to pool 9 (see BC TABLE ..). As lavage cycles 1 to 3 (prelavage with calcium and magnesium) are not included in pool 10 this could indicate a qualitative difference between granulocytes harvested in early and late cycles. The results for the 0-GR should, however, be interpreted with caution as in this group the granulocytes made up only approx. 0.1 to 0.7 percent of total signals analyzed per FCM determination. The quantitative analysis of so few cells may be affected by background noise when cells are identified on the basis of light scatter only.

The phagocytosis data of this study are in general agreement with those obtained in the initial study (a) as far as the macrophages are concerned. A discrepancy is seen, however, in the higher mean number of beads per granulocyte in this study. As phagocytosis appears to be a sensitive parameter for the investigation of smoke inhalation effects, methods should be ~~more standardized~~ optimized for future studies.

In this context attention should be paid to the autofluorescence of FLC which, apart from being a potentially interesting parameter by itself, it limits the resolution of cells with ~~0, 1, 2~~ etc. phag and mon phag ?

(a) see REFERENCES: INBIFO study P 0500/3097

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beads. Autofluorescence is not ~~just~~ simply added to the fluorescence of phagocytized beads but also induces considerable broadening of cell distributions. Thus the CV of autofluorescence signals ranged from approx. 20 to 23 percent in the 0-GR, and from 36 to 53 percent in the 1-GR (see BC TABLE .,) as compared to less than 3 percent for the fluorescent latex beads. The mean intensity was also highest in the 1-GR reaching approximately half the intensity of the latex beads. As at least part of the autofluorescence could be induced by the fixative glutaraldehyde, other fixatives should be tested in future studies.

The difference in autofluorescence between the groups could, at least in part, be due to variation in cell size. Using AXL as an approximate measure of cell size (see .....) the ratio of autofluorescence to AXL was generated ~~on~~ by the computer on a cell ~~by~~ to cell basis. The difference between the groups, though smaller, still persisted in the mean ratio distributions (see BC TABLE ..). As AXL presumably is less than proportional to cell volume, it can not be ruled out that the macrophages in the 0-GR to 1-GR are not different on the basis of autofluorescence per cell volume. Further investigations will be necessary to clarify this point.

6.1.6.2.3 DNA distribution

DNA analysis was planned for a previous study (a) but could not be performed due to technical problems with the laser. The staining procedure was, therefore, taken from the literature (Barlogie et al., 1976, 1979) without optimization for FLC.

The unusual shape of the DNA distribution of FLC already shown in ..... (see BC FIGURE .,) adversely affected the evaluation of DNA parameters. Cells in G2 should contain exactly 2 times the DNA amount of G0/1 cells. Assuming quantitative binding of

(a) see REFERENCES: INBIFO study P 0500/3097

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the DNA stain and linearity of the FCM amplifiers, the fluorescence of G2 cells should therefore also be twice that of G0/1 cells. For the macrophages, however, the ratio was only approx. 1.8 instead of 2 (see BC TABLE ..). This discrepancy was obviously not due to a decreased fluorescence of the G2 peak, as the position of this peak was invariably more than 2 times higher than the granulocyte peak (G0). Furthermore, the CV for the G0/1 peak of macrophages was very high, i. e. 7.5 to 11.8 percent (mean value) for the 0-GR and the 1-GR, respectively. These 2 adverse properties of the DNA distribution make it unacceptable for the analysis of the relative number of cells in the different positions of the cell cycle by computer fitting methods. The G0/1 peak of macrophages was, therefore, more closely examined ~~hoping to~~ in the hope of getting some information as a basis for future optimization of DNA analysis on FLC.

For the quantitative evaluation of the G0/1 peak of macrophages it is desirable to have a reference in order to compensate for variation in instrument setting between the 3 experiments. As no suitable red fluorescent standard particles were available, the G0 peak of granulocytes was evaluated with respect to position and CV. It was found that there was very little variation in the fluorescence properties of granulocytes comparing both groups and pools. The mean position of the G0 peak for the 1-GR and the 2-GR was very close to 100 percent of the 0-GR (see BC TABLE .. and ...,). ~~Very~~ Similar values were also found by comparing the pools (see BC TABLE .. and ...,). According to these data the error in using the G0 peak of granulocytes as a reference should not exceed 3 percent, as a mean. The only systematic difference found was a small increase in the CV of the red fluorescence signal area for the 1-GR and the 2-GR as compared to the 0-GR (see BC TABLE ..). This difference is, however, not relevant for the following consideration.

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The mean excess fluorescence of the G0/1 signal area of macrophages, expressed as shift relative to G0 of granulocytes, was found to be 15, 23 and 18 percent for the 0-GR, 1-GR and the 2-GR, respectively (see BC TABLE .. and ....). The values for the shift in the signal height were negative, indicating a lower maximal DNA concentration in macrophage nuclei as compared to granulocyte nuclei (see BC TABLE ..). The increase in the red fluorescence signal area, indicating a larger amount of fluorescence, may be due to several possible reasons: (1) increased DNA content of nuclei, (2) increased binding of DNA stain, (3) presence of extranuclear DNA, (4) insufficient removal of RNA, (5) autofluorescence, i. e, presence of fluorescent cell components other than DNA or RNA, and (6) instrumental artifacts. (1) and (6) will not be discussed here, as they are not very likely to be responsible for the observed shift (a). (2) is different from (3) to (5) in that it concernes the nucleus only but not the whole macrophage.

As the laser focus is approx. 5 micrometer as compared to approx. 8 micrometer diameter of macrophage nuclei, the spatial resolution of the FCM is sufficient to make a decision between (2) and (3) to (5) on the basis of the relation between signal area and height. Considering equally sized nuclei (b), a change in nuclear fluorescence should affect both signal area and height to the same extent. This relation can be expected for (2). If, however, excess fluorescence were present in the whole macrophage, then the signal height should increase to a lesser extent than the signal area, as the signal baseline for the excess fluorescence would correspond to the macrophage instead of the nuclear diameter. As no systematic difference between the groups was found for the size of macrophage nuclei (see BC TABLE ..), the hypothesis that (2) were true can be tested by plotting signal area (4) versus signal height (x). The

(a) Item (6) will be investigated, as it cannot be ruled out completely.

(b) constant signal baseline

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slope of the regression curve was thus found to be 1.9 to 2.9 for the 3 experiments (a) (see BC FIGURE ..). This result negates hypothesis (2), i. e. the excess fluorescence was, at least in part, located outside the nuclei.

Autofluorescence, as 1 of the remaining potential reasons for the red fluorescence shift, was taken into consideration by correlating the green component autofluorescence of macrophages obtained from the phagocytosis assay to the red fluorescence shift and CV seen in the DNA assay. A positive correlation was found for both parameters (see BC FIGURE .. and ....). This finding does, of course, not confirm hypothesis (5) but it also does not negate it. The presence of residual RNA in macrophages is considered ~~to be very~~ unlikely as the samples have been treated extensively with RNAse. Further investigations will be necessary to find and, hopefully, remove the factor(s) responsible for the shift and broadening of macrophage DNA distribution. (6)?

In spite of these adverse effects on the resolution of macrophage DNA distribution the relative number of macrophages in the G0/1, S and G2 phase of the cell cycle as well as the relative number of macrophage DNA signals corresponding to 2 or more nuclei per count was evaluated using the crude method described in ..... This method tends to overestimate G0/1 and G2, and to underestimate S-phase macrophages. The resulting error is, however, considered to be negligible for G0/1 and G2 when the contribution of S to the total distribution is small. The relative number of macrophages in G0/1 was approx. 94 percent in the 0-GR, 83 to 87 percent in the 1-GR, and approx. 93 percent in the 2-GR. Correspondingly, an increase of S and G2 macrophages was seen for the 1-GR and the 2-GR (see BC TABLES .., .., .. and .., and BC FIGURE ...). The occurence of seemingly significant differences in S and G2 between pools

(a) Regression made for individual experiments as the absolute size of signal height depends on optimal focusing, which was different in the experiments.

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should be considered with caution for the reasons discussed above. The increase in the mean values of pools for the 1-GR compared to the 0-GR can be interpreted as an increase in the number of proliferating cells in the FWD versus AXL region assigned to macrophages. On the basis of the available data, however, the emergence of other proliferating cell types, e. g. interstitial macrophages or lung epithelial cells, can not be excluded. G2 cells did not appear as the largest cells in the macrophage population as would be expected for lines of cultured cells, but were in an intermediate position with respect to AXL (see BC FIGURE ..). However, pulmonary macrophages proliferation has been reported to occur under basal conditions (Adamson and Bowden, 1980) and may be stimulated e. g. by nitrous oxide (Evans et al., 1973). Further refinements in FCM analysis as well as cell sorting experiments will be necessary to identify the type of proliferating lavaged lung cells.

Sample processing and FCM analysis for the DNA assay was performed in a way considered to minimize the formation of cell aggregates in samples and the occurrence of coincident cell signals. More than 99 percent of granulocytes, expected to be all in G0, were actually found in the G0 region (see BC FIGURE .. and BC TABLE .. and ..). Up to 0.8 percent were found in the S region, presumably due to spillover from the adjacent G0 region, and less than 0.3 percent were seen in the G2 region (mean values for the 3 experiments) (see BC TABLE ..). The occurence of doublets or coincidence, and larger aggregates of granulocytes was below 0.1 and approx. 0.01 percent, respectively (see BC TABLE ..). As the cell density of macrophages and granulocytes was similar in the 1-GR samples, the frequency of coincidence of macrophages also should have been approx. 0,1 percent or less. Actually, however, approx. 2 percent of macrophages were found in the 2 x G0/1 region for the

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0-GR and the 2-GR, and approx. 5 percent for the 1-GR. Approx. 0.2 to 1.2 percent of macrophages appeared in the region corresponding to more than 2 x G0/1 DNA content per count (all groups) (see BC TABLE ..). These macrophages represent either aggregates or multinucleated cells. A comparison of the FCM data for the 2 x G0/1 region with the microscopic counts of multinucleated macrophages (a) revealed a correlation (see BC TABLE .. and BC FIGURE ..). The large region corresponding to more than 2 x G0/1 was not included in the comparison, as there were no visible clusters permitting an estimate of the contribution of background signals. In the 2 x G0/1 region, however, most of the cells seemed to be binuclear macrophages and not doublets. The determination of both signal area and height in the DNA assay may therefore provide the basis for the development of a FCM method for the quantification of multinucleated macrophages.

6.1.6.2.4 Light scatter properties of macrophages

Morphological parameters of FLC can be correlated to light scatter, as discussed in 3 INTRODUCTION and were thus included in this study.

FWD of viable macrophages was found to be almost identical for the 0-GR and the 1-GR, but increased to 105 percent in the 2-GR (see BC TABLE .. and ...). This increase was due to the presence of a macrophage subpopulation with high FWD in all 2-GR samples. In the example shown in BC FIGURE .., the macrophage subpopulation with low rising FWD demonstrated a viability of almost 100 percent, whereas the viability decreased considerably in the high FWD subpopulation. Although there were many macrophages with intermediate

(a) almost exclusively binucleated, i. e. 2 x G0, macrophages (see BC TABLE ..)

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viability in the high FWD region, the number of viable cells in this region was still large enough to increase the mean FWD in the total population of viable macrophages. Samples of the other groups with comparable mean number of nonviable and intermediate macrophages were not bimodal in FWD. Therefore the high FWD subpopulation may possibly represent a different subpopulation of macrophages which were only present to a large extent in the 2-GR. In contrast to the results of a previous study (a) with glutaraldehyde fixed FLC, a decrease of FWD in later cycles, i. e. pool 2, 4, 6 and 7, was not consistently seen for viable macrophages (see BC TABLE ..). In accordance with this previous study, however, glutaraldehyde fixed macrophages seemed to have higher FWD in the 1-GR than in the 0-GR (see BC TABLE ..). The discrepancy between fixed and unfixed cells may be due to the selection of viable unfixed macrophages. Fixation may also alter the light scatter properties of cells. However, macrophages in pool 8, 0-GR, experiment 1, had extremely high FWD (see BC TABLE .,) and displayed an atypical FWD versus AXL distribution (see BC FIGURE ..). This sample is a mixture of pool 1 and 2, 0-GR, experiment 1, which was found to contain many nonviable and intermediate macrophages, see ..... Thus the light scatter properties of extensively damaged cells are seen in glutaraldehyde fixed cells as well as viable cells. FWD may therefore not only be useful for cell type differentiation but also may yield information on the quality of samples.

AXL of viable macrophages was found to be 120 to 125 percent for the 1-GR and 104 to 105 percent for the 2-GR compared to the 0-GR (see BC TABLE .. and .., and BC FIGURE ..). Values for the 1st pool of each lavage were almost invariably higher than for subsequent pools. Very similar results for the groups were obtained from glutaraldehyde fixed macrophages in this study (see BC TABLE .. and ..), and in a previous study (a). In terms of cell size

(a) see REFERENCES: INBIFO study P 0500/3097

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(a), this means that macrophages in the 1-GR and the 2-GR were larger compared to the 0-GR. This increase in macrophage size is smaller but qualitatively in agreement with that of the determination of mean macrophage area on cytocentrifuge preparations (see BC TABLE ..). In discrepancy between the 2 methods is considered to be, in part, due to the different shape of the macrophages, i. e. approx. spherical in suspension (FCM) and more or less flat on cytocentrifuge preparations. In addition, the signal height of AXL can be assumed to be less than proportional to the projected area of spherical cells. Future studies should also include the determination of signal width by FCM as a direct measure of cell diameter. This parameter is easier to interpret and less sensitive to drastic changes in the light scatter properties of cells than AXL, but was not determined in the present study due to the lack of suitable standards required for calibration.

The analysis of RAS was included in this study although the FCM electronics could not cover the large dynamic range of signals obtained from FLC. RAS was only evaluated in a way to provide a rough estimate of smoke inhalation effects on this parameter. The relative number of macrophages exceeding 14 times the RAS of standard particles was 14 to 31 percent in the 0-GR, except for pool 8, experiment 1 (see BC TABLE .. and ..). This sample has already been shown to have atypical light scatter properties due to low viability (see .....). The values for the 1-GR were 65 to 73 percent, and 43 to 50 percent for the 2-GR. RAS as a measure of intracellular optical heterogeneity of cells was determined as it is expected to be sensitive to the presence of vacuoles in macrophages. The increase of RAS seen in the treatment groups of the present and a previous study (b) is qualitatively in agreement with the microscopic data on vacuolization (see BC TABLE ..). The relative contribution of cell size to the increase

(a) see INTRODUCTION
(b) see REFERENCES; INBIFO study P 0500/3097

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in RAS can, however, not be determined with the available data. In addition, damaged cells also appear to have very high RAS. This will have to be taken into consideration in further experiments concerned with the interpretation of RAS in the context of vacuolization. According to the results for FWD and RAS, light scatter parameters should best be investigated in combination with viability.

6.1.6.2.5 Differential counting of macrophages and granulocytes

The differential counting of cell types in FLC samples by FCM was restricted to macrophages, granulocytes and red blood cells, which were visible as more or less well defined clusters in FWD versus AXL cytograms (see BC FIGURE ..). Lymphocytes could not be identified on the basis of light scatter due to the lack of positive control samples. According to microscopic differential counting, lymphocytes constituted generally ~~mucir~~ less than 2.5 percent of FLC (excluding red blood cells) (see BC TABLE ..). Therefore the sum of macrophages and granulocytes was taken to be 100 percent for the evaluation of FCM data.

Generally a good correlation was found between the relative number of macrophages and granulocytes obtained from FWD versus AXL cytograms of unfixed (viability and esterase assay), glutaraldehyde fixed (phagocytosis assay) and ethanol fixed (DNA assay) FLC and the number obtained from microscopic counting (see BC TABLES .. to .., and BC FIGURES .. to ..). The slope of the regression line for the relative number of granulocytes was, however, 1.06 to 1.14 for the relation between FCM data (y-axis) and microscope data from resuspension medium (x-axis), indicating a tendency for relatively higher granulocyte counts by FCM. The deviation of

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the slope from the value of 1 could be explained for example by the presence of additional counts in the FWD versus AXL region for granulocytes growing in proportion to the number of granulocytes, or by a constant relative loss of granulcytes on cytocentrifuge preparations.

As the FCM data seemed to overestimate the relative number of granulocytes especially in the 0-GR, i. e. at low values, an effort was made to reduce potential contribution of non-cellular sample components, e. g. cell debris or other particles, to the number of macrophages and granulocytes counted. Thus, only viable cells (a) were counted for unfixed FLC samples, and light scatter signals from ethanol fixed FLC were only counted when they corresponded to at least normal cell DNA content (see BC TABLE .. to ..). Although this procedure substantially reduced the number of granulocytes for the 0-GR, the slope of the regression line even increased for unfixed samples, while the correlation between granulocyte counts from ethanol fixed samples and microscopic counts was essentially unchanged (see BC TABLE .. and BC FIGURES .. and .,). The increase in slope may, in part, result from variation in viability. Therefore, gating for viable cells seems to have only a limited value for cell counting. For low granulocyte counts, however, DNA gating also was effective in eliminating background signals, as is also seen from the relative difference between ungated and gated counts (see BC TABLE .. and BC FIGURE .,).

Background signals and statistical variation in multiple evaluations of the same data due to uncertainty about the precise location of cells in cytograms (see BC TABLE ..) seemed to be a limiting factor in the evaluation of FLC samples containing very few granulocytes. Gating for specific cell parameters like DNA may

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(a) cells used for the evaluation of esterase activity, see ...

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provide a basis for future method optimization. Taking into consideration that the FLC samples analyzed by FCM and microscopy were processed in completely different ways, the consistency of the results was ~~surprisingly~~ good (see BC TABLE .. and BC FIGURES .. and ..). [very?]

In many samples a sharp peak was seen in AXL histograms which corresponded to a cluster in region ... of the FWD versus AXL cytogram (see BC FIGURE ..). As no nucleated cells were found in this region, it was assumed that this peak may represent red blood cells. This interpretation is ~~strongly~~ supported by RBC counts from cytocentrifuge preparations (see BC TABLE ..).

7.1.7 Conclusions

Of the biochemical methods tested in this study, SDS-PAGE and acid phosphatase activity appear to be sensitive enough to be use for determinations in separate subpopulations of FLC. The acid phosphatase method, however, is based on detection of fluorescence product and can be easily adapted to FCM, ~~when~~ the single cell analysis should yield even more information. of this parameter

The determination of phospholipids in FLC have been shown to be too insensitive to be used extensively in FLC studies. The alternative method of FAME pattern using capillary GC, however, appears to be promising for future studies on FLC lipids in small subpopulations. sensitive enough

The cellular composition of FLC was strongly influenced by lavage procedure. Generally, the relative number of granulocytes was larger in the 1st pool of each lavage. Supplimentation of the lavage medium PBS with BSA or calcium and magnesium also increased the proportion of granulocytes.

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The large increase in relative number of granulocytes after MSβ exposure and the small increase for SSβ exposure were in agreement with a previous study (a).

The absolute number of macrophages recovered was also increased by supplimentation with BSA. Although an unexpectedly large number of macrophages were recovered in the prelavage with PBS plus calcium and magnesium, the number of macrophages obtained in the following 10 cycles with PBS plus BSA was comparable to the same number of cycles without prelavage.

The number of macrophages recovered from rats exposed to MSβ was similar to that for controls as ~~already~~ seen in a previous study (a). The decrease in macrophage number after SSβ exposure was also reproduced for PBS alone and PBS plus BSA with prelavage. In the case of PBS plus BSA without prelavage, however, the number of macrophages recovered from the SS group was increased relative to control. As this was only a single determination, no interpretation is possible.

The absolute number of granulocytes confirmed the changes seen for relative number of the same, despite the large variations in macrophage number between lavage procedures and treatment groups.

Viability, determined microscopically, appeared to be lower in PBS alone than for other media. Especially the variation in viability was larger for this medium. The 1st pool of each lavage procedure also demonstrated a small decrease in viability relative to later pools.

For the microscopic determination of macrophage morphology, PBS lavaged samples were not used as the cell preparations were not well preserved. No other changes in macrophage morphology were seen between the

(a) see REFERENCES; INBIFO study P 0500/3097

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3 lavage procedures. There was, however, a definitive increase in macrophage size after smoke exposure. The larger effect of MS$ on macrophage size relative to SS$ for PBS plus BSA without prelavage was not confirmed in PBS plus BSA with prelavage.

The determination of FLC viability by FCM was shown to be practicable in this study. In addition to the information also obtained by the trypan blue method, this method can quantitate intermediate states indicating acute damage to FLC. As this method is also independent of subjective factors, it can and should replace the microscopic method in future studies. The effect of lavage media and smoke exposure were similar to the microscopic data. ?s Text Results

seen by FCM

The determination of nonspecific esterase activity of macrophages in the same assay, was essentially unimodal and demonstrated no difference between lavage procedure or treatment groups. A bimodal distribution was found in granulocytes for this parameter. Due to the poor resolution of the 2 subpopulations, however, an exact evaluation of these data is not yet possible.

however

Phagocytosis as determined by FCM was found to be a sensitive parameter for smoke exposure effects. An increase in phagocytosis relative to control was seen for MS$ and SS$. The small number of data available in this study, especially for SS$, were in agreement with a previous ~~larger~~ study (a). The analysis of granulocyte phagocytosis is a new and interesting aspect of this parameter. Due to the small number of granulocytes in the pools for phagocytosis from controls, no exact evaluation of this parameter was possible. The use of granulocyte enriched pools, e. g. pool 5, for such studies should be incorporated into future studies.

The values for DNA content of macrophages were shifted to higher values compared to granulocytes, possibly due to autofluorescence.

(a) see REFERENCES: INBIFO study P 0500/3097

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

The determination of DNA distribution by FCM offers an opportunity to determine stages of proliferation and multinucleation of macrophages. Both proliferation and multinucleation of macrophages were significantly increased in MS$ exposed FLC. Variations and limited amount of data do not allow an interpretation of this parameter for SS exposed group.

Differential counts determined with FCM on the basis of AXL and FWD are in general agreement with microscopic data, and can be refined by the use of additional parameters such as DNA. Their inclusion in a given assay allows the separate analysis of macrophages and granulocytes in heterogenous FLC samples, and can and replace the microscopic method in future studies.

The light scatter parameters AXL, RAS and FWD were used to analyze samples for subpopulations of macrophages for differing morphology. AXL demonstrated a significant increase after MS$ exposure relative to control. A smaller increase was seen for SS$ exposure. This effect probably correlates with an increase in macrophage size seen microscopically.

This parameter for

For RAS a large increase was also seen for MS$ exposure group relative to control. The SS$ exposed group was also significantly increased relative to control. This group, however, was also significantly lower in RAS than the MS group. Due to the lack of a suitable logarithmic amplifier, evaluation of this parameter was limited, however, it appears to be of potential interest as an assay for vacuolization.

FWD from the viability assay demonstrated no difference between the MS$ exposed group and control. For the SS$ exposed group, however, a small but significant increase in this parameter was seen for viable macrophages. The FWD histogram for this group

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

demonstrated a definite bimodal distribution, indicating a high FWD subpopulation of macrophages. The appearance of such a new subpopulation might be seen in correlation with the decrease in macrophage number for SS$ exposed rats. This effect could then be explained by the compensation of macrophage loss by recruitment of monocytes.

From the results of this study, it is apparent that handling is very critical for FLC parameters. By

(1) the replacement of microscopic determination of viability and differential counts by the FCM methods and

(2) the replacement of microscopic cell counting by a coulter counter at the site of lavage

the time from lavage to analysis or fixation should be reduced significantly.

For the application of FCM to FLC parameters the most important result of this study was the development of evaluation schemes including software for

(1) viability,

(2) phagocytosis and

(3) DNA assays.

Supplimentation of PBS as a lavage medium with BSA increased macrophage yield and viability. The number of lavaged macrophages was further increased by an additional prelavage with PBS plus calcium and magnesium. Although the viability for the latter was slightly decreased relative to PBS plus BSA without prelavage, this lavage procedure including prelavage is considered to be the best compromise for future studies. This procedure not only provides sufficient FLC for the investigation of various parameters, but also yields macrophage fractions with fewer contaminating granulocytes and RBC. In addition, the prelavage yields a fraction enriched in granulocytes which appear to be of special interest in the comparison of NMS$ and SS$ as determined by FCM.

Disc

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Source: https://www.industrydocuments.ucsf.edu/tobacco/

TABLES

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

6.2 Tables and Figures

GROUP	EXPERIMENT NO.	PROTEIN (mg/l) POOL						
		1	2	3	4	5	6	7
0-GR	1	-	53.1	-	93.1	89.6	-	45.3
	2	-	92.7	-	59.6	-	61.8	61.8
	3	-	-	-	114.8	123.5	78.3	88.7
1-GR	1	-	79.2	-	72.2	-	-	92.7
	2	-	173.5	-	151.3	-	-	189.6
	3	-	-	-	187.8	-	138.7	125.3
2-GR	3	-	-	100.9	181.8	-	-	114.8

BC TABLE 1

FLC-PROTEIN

Remarks: amido black method

20290273O4

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A2 PS

GROUP	STATISTICAL PARAMETER	PROTEIN ~~(mg/l)~~ POOL						
		1	2	3	4	5	6	7
0-GR	N	-	2	-	3	2	2	3
	M (mg/l)		72.9		89.2	106.6	70.1	65.3
	SE		-		16.1	-	-	12.7
	RSD (0/0)		-		31.2	-	-	33.6
1-GR	N	-	2	-	3	-	1	3
	M (mg/l)		126.4		137.1		138.7	135.9
	SE		-		34.1		-	28.5
	RSD (0/0)		-		43.1		-	36.3
2-GR	x	-	-	100.9	181.8	-	-	114.8

BC TABLE 2

FLC-PROTEIN, STATISTICAL PARAMETER

Remarks: amido black method

20290273OS

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT	POOL	RELATIVE PEAK AREA (0/0) Mr (a) ← (DALTON 1E3) 45.5	16.0	15.0
0-GR	1	2	32.5	27.6	0
		4	19.5	26.8	27.3
		7	21.1	39.0	0
	2	2	16.7	34.5	9.9
		4	23.0	38.8	12.0
		7	32.9	33.9	0
	3	4	10.9	55.9	0
		7	29.5	48.9	0
1-GR	1	2	17.2	36.6	11.7
		4	14.6	32.2	15.9
		7	23.2	39.7	8.6
	2	2	12.2	12.2	8.2
		4	15.4	23.0	15.7
		7	15.1	24.5	15.8
	3	4	10.1	33.7	13.7
		7	13.6	49.7	0

BC TABLE 3

RELATIVE PEAK AREA FROM SDS-PAGE OF FLC PROTEINS, REDUCED EXCISION METHOD, COOMASSIE BLUE STAINING

Remarks: for Mr standards and amount of protein see BC FIGURE: values are from a single determination for each pool and day

(a) relative molecular mass

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

MD

[an nachstehende Tab. anhängen]

GROUP	EXPERIMENT	POOL	RELATIVE PEAK AREA (0/0) Mr (a) ⇐ (DALTON 1E3) 67.0	45.5	43.0	16.0	15.0
2-GR	3	4	19.3	1.8	0	16.1	43.5
		7	2.5	13.8	0	54.2	0

BC TABLE 83 (continued)

RELATIVE PEAK AREA FROM SDS-PAGE OF FLC PROTEINS, COOMASSIE BLUE STAINING

Remarks: for Mr standards and amount of protein see BC FIGURE: values are from a single determination for each pool and day

(a) relative molecular mass

2029027307

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

NO

GROUP	STATISTICAL PARAMETER	RELATIVE PEAK AREA (~~0/0~~) Mr (a) (Dalton 1E3) 45.5	16.0	15.0
0-GR	N	8	8	8
	M (%)	23.26	38.18	6.15
	SE	2.78	3.54	3.50
	RSD (0/0)	33.8	26.3	160.8
1-GR	N	8	8	8
	M (%)	15.18	31.45	11.20
	SE	1.38	4.06	1.94
	RSD (0/0)	25.6	36.5	49.0
2-GR	N	2	2	2
	M (%)	7.80	35.15	21.75
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 84

RELATIVE PEAK AREA FROM SDS-PAGE OF FLC, STATISTICAL PARAMETERS, COOMASSIE BLUE STAINING

(a) relative molecular mass

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

NO

GROUP	EXPERIMENT	POOL	RELATIVE PEAK AREA (0/0) Mr (a) ← (DALTON 1E3) 44.2	20.0 to 19.0	18 to 17.5	16.3	15.5 to 14.5	12.4
0-GR	1	2	17.4	9.7	8.6	23.6	6.2	0
		4	14.7	11.7	11.3	18.2	24.8	0.4
		5	9.8	8.1	9.0	17.0	28.5	1.0
		7	6.9	5.9	20.8	28.9	15.4	1.8
	2	2	10.3	12.5	12.7	23.0	15.7	1.8
		4	14.3	11.6	11.4	21.9	16.5	0.8
		6	17.7	8.5	9.2	23.5	15.2	1.5
		7	18.4	10.8	10.6	24.3	9.0	0.4
	3	4	5.8	14.7	14.7	41.7 (b)	3.2 (b)	0.4
		5	3.9	9.9	9.0	16.2	44.6	1.5
		6	4.5	15.6	14.7	29.5	9.3	0.53
		7	6.4	17.3	15.2	39.6 (b)	0	.LT.0.2 (b)

BC TABLE 35

RELATIVE PEAK AREA FROM SDS-PAGE OF FLC PROTEINS, SILVER STAINING

Remarks: for Mr standards and amount of protein see BC FIGURE:
values are from a single determination for each pool and day

(a) relative molecular mass
(b) not included in the calculation of means

20290273O9

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT	POOL	RELATIVE PEAK AREA (0/0)					
			Mr (a) ← (DALTON 1E3)					
			44.2	20.0 to 19.0	18.0 to 17.5	16.3	15.5 to 14.5	12.4
1-GR	1	2	8.5	11.8	11.7	22.2	23.5	2.4
		4	11.5	11.8	12.2	17.6	15.8	2.1
		7	12.4	14.6	12.2	17.6	10.2	.LT.0.1 (b)
	2	2	11.1	12.2	11.5	15.4	19.3	5.9
		4	10.3	12.9	13.2	19.1	19.1	5.7
		7	7.8/7	8.5	12.2	18.8	16.0	4.8
	3	4	6.0	11.8	12.6	23.7	19.5	4.6
		6	3.7	12.9	11.7	19.7	22.4	13.4
		7	6.6	16.5	13.2	19.9	12.8	2.1

BC TABLE 35 (continued)

RELATIVE PEAK AREA FROM SDS-PAGE OF FLC PROTEINS, SILVER STAINING

Remarks: for Mr standards and amount of protein see BC FIGURE:
values are from a single determination for each pool and day

(a) relative molecular mass
(b) not included in the calculation of means

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

no.

GROUP	EXPERIMENT	POOL	RELATIVE PEAK AREA (0/0)					
			Mr (a) ← (DALTON 1E3)					
			44.2	20.0 to 19.0	18 to 17.5	16.3	15.5 to 14.5	12.4
2-GR	3	3	6.0	8.7	8.5	14.3	26.6	17.4
		4	2.4	5.1	6.5	12.2	48.8	3.8
		7	6.8	18.9	13.4	23.5	12.4	1.0

BC TABLE 35 (continued)

RELATIVE PEAK AREA FROM SDS-PAGE OF FLC PROTEINS, SILVER STAINING

Remarks: for Mr standards and amount of protein see BC FIGURE:
values are from a single determination for each pool and day

(a) relative molecular mass
(b) not included in the calculation of means

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A6 PS

GROUP	STATISTICAL PARAMETER	RELATIVE PEAK AREA (0/0) Mr (a) ← (DALTON 1E3) 44.2	20.0 to 19.0	18.0 to 17.5	16.3	15.5 to 14.5	12.4
0-GR	N	12	12	12	10	11	11
	M (%)	10.84	11.36	12.3527	22.61	16.84	0.920
	SE	1.57	0.95	1.074	1.43	3.68	0.19 0.20
	RSD (0/0)	50.1	29.0	28.4 29.3	20.0	72.5	69.5 72.7
1-GR	N	9	9	9	9	9	8
	M (%)	8.664	12.56	12.28	19.33	17.62	5.13
	SE	0.97	0.73	0.21	0.83	1.44	1.31
	RSD (0/0)	33.56	17.4	5.1	12.9	24.6	72.1
2-GR	N	3	3	3	3	3	3
	M (%)	5.07	10.90	9.47	16.67	29.27	7.40
	SE	1.35	4.13	2.05	3.47	10.59	5.06
	RSD (0/0)	46.3	65.7	37.5	36.1	62.7	118.5

BC TABLE 16

~~MEAN~~ RELATIVE PEAK AREA FROM SDS-PAGE OF FLC, STATISTICAL PARAMETERS, SILVER STAINING

(a) relative molecular mass

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A10 PS

GROUP	EXPERIMENT	POOL	~~QUOTIENTS~~, RELATIVE PEAK AREA RATIO (a)					
			Mr (b) ← (DALTON 1E3)					
			44.2	20.0 to 19.0	18 to 17.5	16.3	15.5 to 14.5	12.4
1-GR	1	2	0.49	1.22	1.~~22~~36	0.94	3.79	-
		4	0.78	1.01	1.08	0.97	0.64	5.25
		7	1.80	2.47	0.59	0.61	0.66	.LT.0.06 (c)
	2	2	1.08	0.98	0.91	0.67	1.23	3.28
		4	0.72	1.11	1.16	0.87	1.16	7.13
		7	0.42	0.79	1.15	0.77	1.78	12.00
	3	~~2~~ 4	1.03	0.80	0.86	0.57	6.09	11.50
		~~4~~ 6	0.82	0.83	0.80	0.67	2.41	~~26.80~~ 44.67
		7	1.03	0.95	0.87	0.50	-	.GT.10.5 (c)

BC TABLE 7

RATIO OF RELATIVE PEAK AREA FROM SDS-PAGE OF FLC, SILVER STAINING

(a) treatment group versus control group
(b) relative molecular mass
(c) values not used to calculate mean quotient (see BC TABLE 8.)

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT	POOL	~~QUOTIENTS~~, RELATIVE PEAK AREA RATIO (a)					
			Mr (b) ← (DALTON 1E3)					
			44.2	20.0 to 19.0	18 to 17.5	16.3	15.5 to 14.5	12.4
2-GR	3	3	-	-	-	-	-	-
		4	0.41	0.35	0.44	0.29	15.25	9.50
		7	1.06	1.09	0.88	0.59	-	.GT. ~~5.00~~ 5.00 (c)

BC TABLE 7 (continued)

RATIO OF RELATIVE PEAK AREA FROM SDS-PAGE OF FLC, SILVER STAINING

(a) treatment group versus control group
(b) relative molecular mass
(c) values not used to calculate mean quotient (see BC TABLE 8.)

2029027314

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A12 PS

GROUP	STATISTICAL PARAMETER	RELATIVE PEAK AREA ~~(0/0)~~ ~~MEAN~~ RATIO (a) Mr (b) ← (DALTON 1E3)					
		44.2	20.0 to 19.0	18.0 to 17.5	16.3	15.5 to 14.5	12.4
1-GR	N	9	9	9	9	8	6
	M (%)	0.908	1.129	0.960 ~~76~~	0.730	2.220	~~10.993~~ 13.972
	SE	0.136	0.174	0.069 ~~77~~	0.056	0.664	~~3.457~~ 6.297
	RSD (0/0)	44.9	46.3	~~21.5~~ 23.8	22.9	84.6	~~77.0~~ 110.4
2-GR	N	2	2	2	2	1	1
	M (%)	0.735	0.720	0.660	0.440	15.25	9.50
	SE	-	-	-	-	-	-
	RSD (0/0)	-	-	-	-	-	-

BC TABLE 8

~~MEAN~~ RATIO OF RELATIVE PEAK AREA FROM SDS-PAGE OF FLC, STATISTICAL PARAMETERS

(a) treatment group versus control group
(b) relative molecular mass

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	POOL	PHOSPHOLIPID (ug/1E6 ~~microphy.~~ MACROPHAGES)	(ug/1E6 FLC)	(ug/ug FLC protein)
0-GR	2	7	103.8	100.1	1.62
	3	7	369.3	363.0	4.09
1-GR	2	4	261.8	174.6	1.15
	3	7	227.3	180.5	1.44
2-GR	3	4	236.5	211.2	1.16

BC TABLE 9

FLC PHOSPHOLIPID

Remarks: data calculated on the basis of inorganic phosphorus determination and mean phospholipid molecular weight of 775, calculated on the basis of various parameters

(a) treatment group ~~versus control group~~
(b) relative molecular mass
(c) ~~values~~ not used to calculate mean quotient (see BC TABLE ...)

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	FAME (~~TOTAL AREA 1E6/1E6 FLC~~) ← (U/1EG FLC)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	5.76	–	–	3.94 83	3.06	3.3113	4.18	5.57	4.09	4.43
	2	5.57	–	–	3.79	2.7145	3.0145	3.7143	5.1384	3.26	3.87
	3	9.8586	3.3887	7.24	7.68	–	–	3.76	2.79	3.26	3.20
1-GR	1	2.99	–	–	~~3.15~~ 1.73	–	–	1.96	2.6988	2.4332	2.37
	2	2.85	–	–	–	2.58	–	1.50	–	5.01	–
	3	2.9988	2.05	2.40	1.64	2.78	2.32	1.36	2.2231	2.90	2.33
2-GR	3	–	4.5332	–	3.43	3.33	3.37	4.7143	3.24	3.81	3.76

BC TABLE 10

~~TOTAL AREA OF~~ FATTY ACID METHYL ESTERS FROM FLC

Remarks: 1 determination of a single pool, 5 rats per pool
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of cells per pool.

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A15 PS

GROUP	STATISTICAL PARAMETER	FAME (TOTAL AREA 1E6/1E6 FLC) (U/1EG FLC)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	1	1	3	2	2	3	3	3	3
	M	7.06	3.387	7.24	5.140	2.904	3.2119	3.89	4.50	3.54	3.83
	SE	1.40	–	–	1.279	–	–	0.145	0.86	0.28	0.36
	RSD (0/0)	34.3	–	–	42.9 / 43.8	–	–	6.45	33.23	13.5	16.1
1-GR	N	3	1	1	2	2	1	3	2	3	2
	M	2.94	2.05	2.40	2.40 / 1.69	2.68	2.32	1.61	2.465	3.454	2.35
	SE	0.05	–	–	–	–	–	0.18	–	0.7980	–
	RSD (0/0)	2.7	–	–	–	–	–	19.5	–	39.9 / 40.0	–
2-GR	X	–	4.532	–	3.43	3.33	3.37	4.743	3.24	3.81	3.76

BC TABLE 11

TOTAL AREA OF FATTY ACID METHYL ESTERS FROM FLC, STATISTICAL PARAMETERS

Remarks: 1 determination of a single pool, 5 rats per pool
Pool 1 and 2, 2 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of cells per pool.

3

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	PALMITIC ACID METHYL ESTER (ug/1E6 FLC)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	68.67	–	–	41.9	21.4	28.5	49.3	35.8	23.9	36.3
	2	47.7	–	–	38.5	17.6	23.7	53.4	32.23	21.67	30.56
	3	37.45	30.6	34.7	43.3	–	–	38.10	19.6	19.0	22.4
1-GR	1	40.7	–	–	17.3	–	–	25.6	22.6	18.5	21.3
	2	31.6	–	–	–	24.2	–	9.4	–	25.9	–
	3	20.6	18.9	19.5	18.3	33.0	27.1	22.0	20.7	26.8	23.9
2-GR	3	–	25.9	–	31.7	26.9	28.6	40.1	25.1	37.23	33.9

BC TABLE 12

PALMITIC ACID METHYL ESTER (16:0) FROM FLC

Remarks: 1 determination of a single pool, 5 rats per pool
Pool 1 and 2, 2 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of cells per pool.

3

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A17 PS

GROUP	STATISTICAL PARAMETER	PALMITIC ACID METHYL ESTER (ug/1E6 FLC) POOL 1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	1	1	3	2	2	3	3	3	3
	x̄	51.2330	30.6	34.7	41.23	19.50	26.10	46.930	29.203	21.503	29.737
	SE	9.18	–	–	1.43	–	–	4.5760	4.912	1.42	4.03
	RSD (0/0)	31.0	–	–	6.0	–	–	16.9 17.0	29.12	11.4	23.5
1-GR	N	3	1	1	2	2	1	3	2	3	2
	x̄	30.97	18.9	19.5	17.80	28.60	27.1	19.00	21.6570	23.73	22.60
	SE	5.81	–	–	–	–	–	4.91	–	2.63	–
	RSD (0/0)	32.5	–	–	–	–	–	44.8	–	19.2	–
2-GR	x	–	25.9	–	31.7	26.9	28.6	40.1	25.1	37.23	33.9

BC TABLE 13

PALMITIC ACID METHYL ESTER (16:0) FROM FLC, STATISTICAL PARAMETERS

Remarks: 1 determination of a single pool, 5 rats per pool
Pool 1 and 2, 2 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of cells per pool.

3

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A18 PS

GROUP	EXPERIMENT NO.	STEARIC ACID METHYL ESTER (ug/1E6 FLC)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	15.10	-	-	11.5	10.1	10.6	12.6	13.18	7.2	10.7
	2	12.5	-	-	11.5	8.34	9.23	12.3	~~14.0~~ 13.3	10.6	11.88
	3	1.6 ~~11.5~~ (α)	10.3 (α)	-	~~21.1~~	-	-	11.6	9.45	11.7	11.0
						12.1					
1-GR	1	6.9	-	-	4.7	-	-	5.9	9.1	8.6	8.0
	2	5.87	-	-	-	7.8	-	2.4	-	8.65	-
	3	5.6	6.6	6.2	4.8	8.5	7.0	3.5	5.8	9.32	6.9
2-GR	3	-	11.6	-	10.0	~~10.9~~ 11.0	10.6	8.8	10.0	12.43	11.21

BC TABLE 14

STEARIC ACID METHYL ESTER (18:0) FROM FLC

Remarks: 1 determination of a single pool, 5 rats per pool
Pool 1 and 2, 2 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of cells per pool.

3

(α) outlier not used in the calculation of means

20290273321

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A19 PS

GROUP	STATISTICAL PARAMETER	STEARIC ACID METHYL ESTER (ug/1E6 FLC)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	2	–	–	3	2	2	3	3	3	3
	x̄	13.80	–	–	11.70	9.20	9.90	12.67	12.37	9.83	11.20
	SE	–	–	–	0.20	–	–	0.30	1.45	1.35	0.36
	RSD (0/0)	–	–	–	3.0	–	–	4.2	20.3	23.9	5.6
1-GR	N	3	1	1	2	2	1	3	2	3	2
	x̄	6.03	6.6	6.2	4.75	8.15	7.0	3.93	7.45	8.83	7.45
	SE	0.42	–	–	–	–	–	1.03	–	0.22	–
	RSD (0/0)	12.4	11.9	–	–	–	–	45.5	–	4.6	–
2-GR	x	–	11.6	–	10.0	10.9 11.0	10.6	8.8	10.0	12.3	11.1

BC TABLE 15

STEARIC ACID METHYL ESTER (18:0) FROM FLC, STATISTICAL PARAMETERS

Remarks: 1 determination of a single pool, 5 rats per pool
Pool 1 and 2, 2 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of cells per pool.

3

20290273?2

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A20 PS

GROUP	POOL NO.	FAME ~~(ug/1E6 FLC)~~ ~~RATIO~~ ((g/1E6 FLC)/(g/1E6 FLC)) EXPERIMENT 1	2	3	STATISTICAL PARAMETER N	M	SE	RSD (0/0)
0-GR	1	4.5~~4~~8	3.82	# 23.44(a)	2	4.~~18~~20	–	–
	2	–	–	# 2.37(a)	–	–	–	–
	1 and 2	–	–	–	–	–	–	–
	3	3.64	3.35	3.58	3	3.52	0.09	4.3
	4	2.12	2.1~~2~~0	–	2	2.1~~2~~1	–	–
	3 and 4	2.6~~7~~9	2.5~~8~~5	–	2	2.6~~3~~2	–	–
	5	3.91	4.34	3.28	3	3.84	0.31	13.9
	6	2.~~61~~59	2.3~~0~~2	2.0~~9~~6	3	2.3~~3~~2	0.15	11.~~2~~4
	7	3.32	2.0~~4~~5	1.62	3	2.33	0.51	~~38.4~~ 37.9
	5, 6 and 7	3.39	2.5~~6~~9	2.04	3	2.6~~6~~7	0.39	25.~~6~~4

BC TABLE 16

RATIO OF PALMITIC ACID METHYL ESTER (16:0) VERSUS STEARIC ACID METHYL ESTER (18:0)

Einzelwerte und Mittelwerte auf getrennten Tabellen siehe Tab. 14/15

(a) outlier not used in the calculation of means

2029027323

GROUP	POOL NO.	FAME (~~ug/1E6 FLC~~) ~~RATIO~~ ((g/1E6 FLC)/(g/1E6 FLC))						
		EXPERIMENT			STATISTICAL PARAMETER			
		1	2	3	N	M	SE	RSD (0/0)
1-GR	1	5.90	5.64	3.68	3	5.07	0.706	23.96
	2	-	-	2.86	1	2.86	-	-
	1 and 2	-	-	3.15	1	3.15	-	-
	3	3.68	-	3.81	2	3.75	-	-
	4	-	3.10	3.88	2	3.49	-	-
	3 and 4	-	-	3.87	1	3.87	-	-
	5	4.34	3.92	6.29	3	4.85	0.73	26.1
	6	2.48	-	3.57	2	3.03	-	-
	7	2.15	3.05	2.91	3	2.70	0.28	17.39
	5, 6 and 7	2.66	-	3.46	2	3.06	-	-

BC TABLE 16 (continued)

RATIO OF PALMITIC ACID METHYL ESTER (16:0) VERSUS STEARIC ACID METHYL ESTER (18:0)

20290273324

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL NO.	FAME (~~ug/1E6 FLC~~) ~~RATIO~~ ((g /1E6 FLC)/(g /1E6 FLC))						
		EXPERIMENT			STATISTICAL PARAMETER			
		1	2	3	N	M	SE	RSD (0/0)
2-GR	1	-	-	-	-	-	-	-
	2	-	-	2.23	1	2.23	-	-
	1 and 2	-	-	-	-	-	-	-
	3	-	-	3.17	1	3.17	-	-
	4	-	-	2.4~~7~~5	1	2.4~~7~~5	-	-
	3 and 4	-	-	2.70	1	2.70	-	-
	5	-	-	4.56	1	4.56	-	-
	6	-	-	2.51	1	2.51	-	-
	7	-	-	3.0~~0~~3	1	3.0~~0~~3	-	-
	5, 6 and 7	-	-	3.0~~3~~5	1	3.0~~3~~5	-	-

BC TABLE 16 (continued)

RATIO OF PALMITIC ACID METHYL ESTER (16:0) VERSUS STEARIC ACID METHYL ESTER (18:0)

20290273325

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A30 PS

STATISTICAL PARAMETER	FAME ~~(ug/1E6 FLC)~~ ~~MEAN RATIO~~	((g/1EGFLC)/(g/1EGFLC))
	POOL 1, 3 ~~and~~ 5	POOL 2, 4, 6 ~~and~~ 7
N	18	20
M	4.19	2.62
SE	0.21	0.13
RSD (0/0)	21.04	22.01

BC TABLE 17

MEAN RATIO PALMITIC ACID METHYL VERSUS STEARIC ACID METHYL ESTER

Remarks: all groups pooled

20290273326

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A23 PS

no.

GROUP	EXP. NO.	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC)										
		POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)
0-GR	1	1	43.9			2	~~44.0~~ 42.9			-	-		
			46.1				42.4	43.83			-		
			46.4	45.47	3.0		46.2	~~44.20~~	4.~~3~~ 7		-	-	-
	2	1	60.9			2	53.2				-		
			61.0				62.6				-		
			55.7	59.20	5.1		58.0	~~52~~ 57.93	8.1	-	-	-	-
	3	1	223.6			2	374.2				-		
			220.8				374.2				-		
			209.1	217.83	3.5		377.0	375.13	0.4	-	-	-	-

BC TABLE 18

ACID PHOSPHATASE ACTIVITY

Remarks: M: the means of triplicate determination for each pool, 5 rats per pool

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) A24 PS

GROUP	EXP. NO.	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC)										
		POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)
0-GR	1	3	29.6			4	38.9			-	-		
			31.5				37.5				-		
			33.5 ~~32.2~~	31.~~10~~ 53	~~4.3~~ 6.2		42.9	39.77	7.0		-	-	-
	2	3	299.7			4	95.5			-	-		
			311.6				97.4				-		
			313.6	308.30	2.4		95.9	96.27	1.0		-	-	-
	3	3	395.7			4	776.7			-	-		
			397.3				831.7				-		
			412.0	401.67	2.24		773.7	794.03	4.1		-	-	-

BC TABLE 18 (continued)

ACID PHOSPHATASE ACTIVITY

Remarks: M: the means of triplicate determination for each pool, 5 rats per pool

20290273 28

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC)									
		POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)
0-GR	1	5	53.0 54.0 57.7	54.90	4.5	6	25.0 22.4 26.7	24.70	8.8	7	24.3 22.0 23.9	23.40	5.3
	2	5	355.0 342.1 354.1	350.40	2.06	6	323.8 311.8 311.8	315.80	2.2	7	334.3 341.3 352.8	342.80	2.7
	3	5	854.7 864.2 829.5	849.47	2.11	6	712.2 749.4 701.0	720.87	3.5	7	249.3 251.9 253.4	251.53	0.8

BC TABLE 18 (continued)

ACID PHOSPHATASE ACTIVITY

Remarks: M: the means of triplicate determination for each pool, 5 rats per pool

20290273 29

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC)										
		POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)
1-GR	1	1	2.8			2	19.2			-	-		
			2.6 3.3	2.40			19.7				-		
			-	3.05	-		19.1	19.33	1.7		-	-	-
	2	1	322.7			2	398.7			-	-		
			310.9				338.1				-		
			268.8	300.80	9.4		369.3	368.70	8.2		-	-	-
	3	1	424.4			2	557.4			-	-		
			410.5				545.0				-		
			450.4	428.43	4.7		553.2	551.87	1.1		-	-	-

BC TABLE 19

ACID PHOSPHATASE ACTIVITY

Remarks: M: the means of triplicate determination for each pool, 5 rats per pool

20290273330

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

NO.

GROUP	EXP. NO.	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC)										
		POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)
1-GR	1	3	5.9			4	26.0			-	-		
			4.9				26.5				-		
			5.1	5.30	10.0		41.2	31.23	27.6		-	-	-
	2	3	-			4	353.8			-	-		
			-				388.3				-		
			-	-	-		352.3	364.80	5.6		-	-	-
	3	3	254.9			4	903.4			-	-		
			246.7				989.9 ~~990.0~~				-		
			252.8	251.47	1.7		1000.5	964.60	5.5		-	-	-

BC TABLE 19 (continued)

ACID PHOSPHATASE ACTIVITY

Remarks: M: the means of triplicate determination for each pool, 5 rats per pool

20290273331

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC)										
		POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)	POOL	x	M	RSD (0/0)
1-GR	1	5	37.1			6	39.6			7	109.0		
			34.9				39.7				103.9		
			34.0	35.33	4.5		39.7	39.5	1.0		98.6	103.83	5.0
	2	5	132.9			6	-			7	569.2		
			129.8				-				548.2		
			131.7	131.47	1.2		-	-	-		521.9	546.43	4.3
	3	5	419.0			6	501.1			7	489.5		
			427.5				495.5				495.5		
			383.8 ~~405.5~~	~~417.33~~	~~2.7~~		503.1	499.90	0.8		479.6	488.20	1.6

BC TABLE 19 (continued)

ACID PHOSPHATASE ACTIVITY

Remarks: M: the means of triplicate determination for each pool, 5 rats per pool

20290273332

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC) POOL	x	M	RSD (0/0)
2-GR	3	1	487.7 505.3 477.30	490.00	2.9
		2	554.1 550.1 569.8	558.00	1.9
		3	709.9 686.5 664.0	686.80	3.3
		4	428.5 399.1 370.4	399.33	7.3
		5	766.0 783.6 810.9	786.83	2.9
		6	589.9 594.7 575.4	586.67	1.7
		7	332.0 308.1 324.0	321.37	3.8

BC TABLE 20

ACID PHOSPHATASE ACTIVITY

Remarks: M: the means of triplicate determination for each pool, 5 rats per pool

20290273333

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) B1 PS

GROUP	STATISTICAL PARAMETER	ACID PHOSPHATASE ACTIVITY (U/10E6 FLC) POOL 1	2	3	4	5	6	7
0-GR	N	3	3	3	3	3	3	3
	M	107.500	159.087	247.023	310.023	418.2573	353.7908	205.910
	SE	55.307	108.0942	111.2751	242.562	231.8683	201.8639	94.982 95.0
	RSD (0/0)	89.1	117.79	77.3 78.0	135.5	96.0	98.8	79.9
1-GR	N	3	3	2	3	3	2	3
	M	243.9 244.093	313.300	128.3854	453.553	194.710	269.700	379.4875
	SE	126.027	156.287	-	273.0791	114.718	-	138.850
	RSD (0/0)	89.4	86.4	-	104.3	102.0	-	63.4
2-GR	N							
	M	490.X0	558.0	686.8	399.3	786.8	586.7	321.374
	SE	-	-	-	-	-	-	-
	RSD (0/0)	-	-	-	-	-	-	-

BC TABLE 21

Means ACID PHOSPHATASE ACTIVITY, STATISTICAL PARAMETERS

Remarks: M: the means of 3 pools, 1 triplicate determination for each pool, 5 rats per pool

20290273334

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL NO.	STATISTICAL PARAMETER	ACID PHOSPHATASE (U/1E6 FLC) MEAN RATIO (a) EXPERIMENT 1	2	3	STATISTICAL PARAMETER M	SE	RSD (0/0)
1-GR	1		0.1	5.1	2.0	2.40	1.46	105.2
	2		0.4	6.4	1.5	2.77	1.84	115.5
	3		0.2	-	0.6	0.40	-	-
	4		0.8	3.8	1.2	1.93	0.94	84.3
	5		0.6	0.4	0.5	0.50	0.06	20.0
	6		1.6	-	0.7	1.15	-	-
	7		4.4	1.6	1.9	2.63	0.89	58.4
		N	7	5	7	7	5	5
		M	1.16	3.46	1.20	1.9468	0.761.04	67.876.7
		SE	0.57	0.10	0.24	-	-	-
		RSD (0/0)	130.9	71.2	51.8	-	-	-
2-GR	1		-	-	2.32	2.32	-	-
	2		-	-	1.5	1.5	-	-
	3		-	-	1.7	1.7	-	-
	4		-	-	0.5	0.5	-	-
	5		-	-	0.9	0.9	-	-
	6		-	-	0.8	0.8	-	-
	7		-	-	1.3	1.3	-	-
		N	-	-	7	7	-	-
		M	-	-	1.297	1.297	-	-
		SE	-	-	0.232	-	-	-
		RSD (0/0)	-	-	47.6 46.0	-	-	-

2 Tabellen

BC TABLE 22

RATIO OF ACID PHOSPHATASE ACTIVITY IN TREATMENT VERSUS CONTROL GROUP, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

(a) treatment group versus control group

20290273335

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF MACROPHAGES (0/0)						
		POOL						
		1	2	3	4	5	6	7
0-GR	1	98.5	98.6	99.2	99.2	98.9	98.8	99.4
	2	99.2	97.2	98.4	98.3	96.5	94.7	96.4
	3	96.6	96.7	97.3	99.4	94.9	97.8	98.3
1-GR	1	23.8	52.8	43.3	65.7	39.0	61.4	80.5
	2	28.7	54.2	30.3	66.7	23.2	46.6	66.9
	3	28.5	58.1	28.7	64.8	27.9	56.1	79.4
2-GR	3	86.5	91.5	68.7	89.3	77.4	89.8	97.5

BC TABLE 23

RELATIVE NUMBER OF MACROPHAGES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500

2029027336

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

RELATIVE NUMBER OF MACROPHAGES

GROUP	STATISTICAL PARAMETER	ACID PHOSPHATASE ACTIVITY (u/10E6 FLC) POOL						
		1	2	3	4	5	6	7
0-GR	N	3	3	3	3	3	3	3
	M (0/0)	98.10	97.50	98.30	98.97	96.77	97.10	98.03
	SE	0.78	0.57	0.55	0.34	1.16	1.23	0.88
	RSD (0/0)	1.4	1.0	1.0	0.6	2.1	2.2	1.5
1-GR	N	3	3	3	3	3	3	3
	M (0/0)	27.00	55.03	34.10	65.73	30.03	54.70	75.60
	SE	1.60	1.59	4.62	0.55	4.68	4.33	4.36
	RSD (0/0)	10.3	5.0	23.5	1.4	27.0	13.7	10.0
2-GR	x (%)	86.5	91.5	68.7	89.3	77.4	89.8	97.5

BC TABLE 24

RELATIVE NUMBER OF MACROPHAGES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETERS

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500

20290273337

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF MACROPHAGES (0/0)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	99.2	99.8	99.6	96.7	97.3	97.1	97.0	98.8	97.5	97.6
	2	97.8	99.4	98.8	98.6	99.4	99.2	98.9	98.3	98.3	98.4
	3	97.5	99.3	98.2	97.4	99.6	99.3	96.4	99.1	98.9	98.5
1-GR	1	41.3	79.5	67.7	37.1	72.1	53.1	45.4	74.0	92.3	76.1
	2	32.1	56.4	43.2	-	59.8	-	26.2	52.6	65.7	54.7
	3	46.3	52.0	49.9	37.2	67.3	55.2	24.8	45.7	73.5	53.8
2-GR	3	93.8	91.3	92.2	71.1	90.3	83.5	72.0	90.4	92.4	89.0

BC TABLE 25

RELATIVE NUMBER OF MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500 cells
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the means of these pools weighted according to the number of cells per pool.

20290273338

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	MEAN RELATIVE NUMBER OF MACROPHAGES									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	3	3	3	3	3	3	3	3	3
	M (0/0)	98.17	99.50	98.87	97.57	98.77	98.53	97.43	98.73	98.23	98.17
	SE	0.52	0.15	0.41	0.55	0.74	0.72	0.75	0.23	0.41	0.28
	RSD (0/0)	0.9	0.3	0.7	1.0	1.3	1.3	1.3	0.4	0.7	0.5
1-GR	N	3	3	3	2	3	2	3	3	3	3
	M (0/0)	39.90	62.63	53.60	37.15	66.40	54.15	32.13	57.43	77.17	61.53
	SE	4.16	8.53	7.31	-	3.58	-	6.65	8.52	7.89	7.29
	RSD (0/0)	18.1	23.6	23.6	-	9.3	-	35.8	25.7	17.7	20.5
2-GR	x (%)	93.8	91.3	92.2	71.1	90.3	83.5	72.0	90.4	92.4	89.0

BC TABLE 26

MEAN RELATIVE NUMBER OF MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETERS

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the means of these pools weighted according to the number of cells per pool.

2029027339

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF GRANULOCYTES (0/0)						
		POOL						
		1	2	3	4	5	6	7
0-GR	1	0.7	0.2	0.4	0.0	0.6	0.6	0.0
	2	0.0	0.0	0.8	0.4	2.7	3.8	3.0
	3	0.4	1.5	1.0	0.0	2.3	1.2	1.3
1-GR	1	75.0	46.8	56.0	34.3	60.5	38.0	19.5
	2	71.0	45.6	69.2	33.1	76.8	53.0	33.0
	3	70.5	39.3	69.5	34.3	71.1	42.7	19.7
2-GR	3	12.7	8.3	30.8	10.0	22.4	9.8	2.4

BC TABLE 27

RELATIVE NUMBER OF GRANULOCYTES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500

20290273 40

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	~~MEAN~~ RELATIVE NUMBER OF GRANULOCYTES POOL						
		1	2	3	4	5	6	7
0-GR	N	3	3	3	3	3	3	3
	M (0/0)	0.37	0.57	0.73	0.13	1.87	1.87	1.43
	SE	0.20	0.47	0.18	0.13	0.64	0.98	0.87
	RSD (0/0)	95.8	143.7	41.7	173.2	59.7	91.1	105.0
1-GR	N	3	3	3	3	3	3	3
	M (0/0)	72.17	43.90	64.90	33.90	69.47	44.57	24.07
	SE	1.42	2.33	4.45	0.40	4.78	4.43	4.47
	RSD (0/0)	3.4	9.2	11.9	2.0	11.9	17.2	32.1
2-GR	x (%)	12.7	8.3	30.8	10.0	22.4	9.8	2.4

BC TABLE 28

~~MEAN~~ RELATIVE NUMBER OF GRANULOCYTES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETER

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500

20290273341

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF GRANULOCYTES (0/0)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	0.4	0.0	0.1	1.3	0.4	0.7	0.8	0.0	0.0	0.3
	2	1.2	0.2	0.6	1.0	0.6	0.7	1.1	1.7	1.7	1.6
	3	1.1	0.2	0.7	0.9	0.0	0.1	1.7	0.6	0.9	0.9
1-GR	1	58.7	20.5	38.2	62.9	27.9	46.9	54.6	26.0	7.7	23.9
	2	67.6	43.6	56.6	-	39.5	-	73.3	47.2	33.9	44.9
	3	52.1	47.0	48.9	61.9	32.5	44.4	74.6	53.7	26.3	45.8
2-GR	3	5.5	8.5	7.4	28.3	8.8	15.7	27.2	9.1	7.0	10.4

BC TABLE 29

RELATIVE NUMBER OF GRANULOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the means of these pools weighted according to the number of cells per pool.

(BC FIGURE H0364, H0363)

20290273 42

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) B11 PS

GROUP	STATISTICAL PARAMETER	MEAN RELATIVE NUMBER OF GRANULOCYTES									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	3	3	3	3	3	3	3	3	3
	M (0/0)	0.90	0.13	0.~~35~~47	1.07	0.33	0.50	1.20	0.77	0.87	0.93
	SE	0.25	0.07	0.~~25~~19	0.12	0.18	0.20	0.26	0.50	0.49	0.38
	RSD (0/0)	48.4	86.6	~~101.0~~68.3	19.5	91.7	69.3	38.2	112.5	98.1	69.7
1-GR	N	3	3	3	2	3	2	3	3	3	3
	M (0/0)	59.47	37.03	47.90	62.40	33.30	~~40.8~~45.65	67.50	42.30	22.63	38.20
	SE	4.49	8.32	5.34	-	3.37	-	6.46	8.36	7.78	7.15
	RSD (0/0)	13.1	38.9	19.3	-	17.5	-	16.6	34.2	59.6	32.4
2-GR	x (%)	5.5	8.5	7.4	28.3	8.8	15.7	27.2	9.1	7.0	10.4

BC TABLE 30

MEAN RELATIVE NUMBER OF GRANULOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETERS

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the means of these pools weighted according to the number of cells per pool..

2029027343

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0080

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF LYMPHOCYTES (0/0) POOL						
		1	2	3	4	5	6	7
0-GR	1	0.7	1.2	0.4	0.8	0.6	0.6	0.6
	2	0.8	2.8	0.8	1.3	0.8	1.5	0.6
	3	3.0	1.8	1.7	0.6	2.7	1.0	0.4
1-GR	1	1.2	0.4	0.7	0.0	0.5	0.6	0.0
	2	0.4	0.2	0.6	0.2	0.0	0.4	0.2
	3	1.0	2.6	1.7	1.0	1.0	1.2	1.0
2-GR	3	0.8	0.2	0.5	0.8	0.2	0.4	0.1

BC TABLE 31

RELATIVE NUMBER OF LYMPHOCYTES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500

20290273 44

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD91 (R) B13 PS

GROUP	STATISTICAL PARAMETER	MEAN RELATIVE NUMBER OF LYMPHOCYTES						
		POOL						
		1	2	3	4	5	6	7
0-GR	N	3	3	3	3	3	3	3
	M (0/0)	1.50	1.93	0.97	0.90	1.37	1.03	0.53
	SE	0.75	0.47	0.38	0.21	0.67	0.26	0.07
	RSD (0/0)	86.7	41.8	68.9	40.1	84.8	43.6	21.7
1-GR	N	3	3	3	3	3	3	3
	M (0/0)	0.87	1.07	1.00	0.40	0.50	0.73	0.40
	SE	0.24	0.77	0.35	0.31	0.29	0.24	0.31
	RSD (0/0)	48.0	124.8	60.8	132.3	100.0	56.8	132.3
2-GR	x (%)	0.8	0.2	0.5	0.8	0.2	0.4	0.1

BC TABLE 32

MEAN RELATIVE NUMBER OF LYMPHOCYTES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETERS

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500

20290273A5

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF LYMPHOCYTES (0/0)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	0.4	0.2	0.3	1.9	2.3	2.2	2.2	1.2	2.5	2.1
	2	1.0	0.4	0.6	0.4	0.0	0.1	0.0	0.0	0.0	0.0
	3	1.4	0.6	1.1	1.6	0.4	0.6	1.9	0.4	0.2	0.5
1-GR	1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
	2	0.3	0.0	0.2	-	0.7	-	0.6	0.2	0.4	0.4
	3	1.7	1.0	1.3	0.9	0.2	0.5	0.6	0.7	0.2	0.4
2-GR	3	0.8	0.2	0.4	0.6	0.9	0.8	0.8	0.6	0.6	0.6

BC TABLE 33

RELATIVE NUMBER OF LYMPHOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the means of these pools weighted according to the number of cells per pool.

2029027346

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 1 GD91 (R) B15 PS

GROUP	STATISTICAL PARAMETER	MEAN RELATIVE NUMBER OF LYMPHOCYTES									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	3	3	3	3	3	3	3	3	3
	M (0/0)	0.93	0.40	0.67	1.30	0.90	0.97	1.37	0.53	0.90	0.87
	SE	0.29	0.12	0.23	0.46	0.71	0.63	0.69	0.35	0.80	0.63
	RSD (0/0)	53.9	50.0	60.6	61.1	136.5	113.5	87.3	114.6	154.4	126.6
1-GR	N	3	3	3	2	3	2	3	3	3	3
	M (0/0)	0.67	0.33	0.50	0.45	0.30	0.25	0.40	0.30	0.20	0.27
	SE	0.52	0.33	0.40	-	0.21	-	0.20	0.21	0.12	0.13
	RSD (0/0)	136.1	173.2	140.0	-	120.2	-	86.6	120.2	100	86.6
2-GR	x (%)	0.8	0.2	0.4	0.6	0.9	0.8	0.8	0.6	0.6	0.6

BC TABLE 34

MEAN RELATIVE NUMBER OF LYMPHOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETERS

Remarks: sum of macrophages, granulocytes and lymphocytes taken to be 100 percent
total count: approx. 500
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the means of these pools weighted according to the number of cells per pool.

2029027347

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	NUMBER OF MACROPHAGES/~~RAT (1E6)~~ (MEG/RAT)									
		POOL									
		1	2	1 + 2	3	4	3 + 4	5	6	7	5, 6 + 7
0-GR	1	0.59	1.00	1.59	0.77	1.45	2.22	1.21	0.65	1.24	3.10
	2	0.66	1.11	1.77	0.77	1.90	2.67	0.67	1.00	1.90	3.57
	3	0.45	0.31	0.76	0.23	1.28	1.51	0.53	1.01	1.71	3.25
1-GR	1	0.31	0.69	1.00	0.70	1.15	1.85	0.50	0.77	2.04	3.31
	2	0.44	0.65	1.09	0.54	1.71	2.25	0.17	0.38	1.20	1.75
	3	0.29	0.54	0.83	0.62	1.66	2.28	0.40	0.79	2.26	3.45
2-GR	3	0.16	0.28	0.44	0.79	1.83	2.62	0.25	0.71	1.31	2.27

BC TABLE 35

~~RELATIVE~~ NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER

Remarks: macrophages counted after centrifugation and resuspension

20290273 48

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	MEAN NUMBER OF MACROPHAGES/RAT (MEG)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N (MEG/RAT)	3	3	3	3	3	3	3	3	3	3
	M (0/0) (MEG)	0.567	0.807	1.373	0.590	1.543	2.133	0.803	0.887	1.617	3.307
	SE	0.062	0.250	0.311	0.180	0.185	0.338	0.207	0.118	0.196	0.139
	RSD (0/0)	18.9	53.8	39.2	52.8	20.8	27.4	44.7	23.1	21.0	7.3
1-GR	N (MEG/RAT)	3	3	3	3	3	3	3	3	3	3
	M (0/0) (MEG)	0.347	0.627	0.973	0.620	1.507	2.127	0.357	0.647	1.833	2.837
	SE	0.047	0.045	0.076	0.046	0.179	0.139	0.098	0.134	0.323	0.545
	RSD (0/0)	23.5	12.4	13.6	12.9	20.6	11.3	47.4	35.7	30.5	33.3
2-GR	x (MEG) (MEG/RAT)	0.16	0.28	0.44	0.79	1.83	2.62	0.25	0.71	1.31	2.27

BC TABLE 36

MEAN NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER, STATISTICAL PARAMETERS

Remarks: macrophages counted after centrifugation and resuspension

20290273 49

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	NUMBER OF ~~MACROPHAGES~~ GRANULOCYTES/~~RAT (1E3)~~ (1E3/RAT)									
		POOL									
		1	2	1 ~~and~~ + 2	3	4	3 ~~and~~ + 4	5	6	7	5, 6 ~~and~~ + 7
0-GR	1	2	0	2	11	6	17	10	0	0	10
	2	8	2	10	8	11	19	8	18	33	59
	3	5	1	6	2	0	2	9	6	16	31
1-GR	1	440	178	618	1189	446	1635	600/3	271	171	1045
	2	927	503	1430	-	1130	-	476	341	619	1437
	3	326	489	815	1033	803	1836	1205	928	809	2942
2-GR	3	9	26	36	314	178	492	94	71	99	265

BC TABLE 37

NUMBER OF GRANULOCYTES PER RAT, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: absolute number of granulocytes calculated on the basis of absolute and relative number of macrophages

2029027350

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

NO.

GROUP	STATISTICAL PARAMETER	~~MEAN~~ NUMBER OF GRANULOCYTES/~~RAT~~ (1E3)									
		POOL									
		1	2	1 ~~and~~ + 2	3	4	3 ~~and~~ + 4	5	6	7	5, 6 ~~and~~ + 7
0-GR	N	3	3	3	3	3	3	3	3	3	3
	M (1E3/RAT)	5.0	1.0	6.0	7.0	5.7	12.7	9.0	8.0	16.3	33.3
	SE	1.7	0.6	2.3	2.6	3.2	5.4	0.6	5.3	9.5	14.2
	RSD (0/0)	60.0	100.0	66.7	65.5	97.2	73.4	11.1	114.6	101.0	73.7
1-GR	N	3	3	3	2	3	2	3	3	3	3
	M (1E3/RAT)	564.3	390.0	954.3	1111.0	793.0	1735.5	760.3	513.3	533.0	1808.0
	SE	184.3	106.1	244.5	-	197.5	-	225.2	208.3	189.1	578.2
	RSD (0/0)	56.6	47.1	44.4	-	43.1	-	51.3	70.3	61.5	55.4
2-GR	x (1E3/RAT)	9	26	36	314	178	492	94	71	99	265

BC TABLE 38

~~MEAN~~ NUMBER OF GRANULOCYYTES PER RAT, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETERS

Remarks: absolute number of granulocytes calculated on the basis of absolute and relative number of macrophages

20290273S1

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	VIABILITY (0/0) POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	93.8	96.5	95.5	98.3	99.8	99.3	98.4	99.8	99.8	99.3
	2	98.7	98.8	98.8	99.4	100.0	99.8	98.4	99.4	99.6	99.3
	3	98.5	97.8	98.2	99.2	99.8	99.7	98.3	99.6	99.6	99.4
1-GR	1	93.7	96.7	95.8	98.5	99.1	98.9	97.0	99.2	98.6	98.5
	2	97.9	95.9	96.7	98.8	99.4	99.3	97.4	99.2	99.4	99.2
	3	96.2	96.6	96.5	98.8	97.1	97.6	98.7	98.6	98.9	98.8
2-GR	3	96.5	96.9	96.8	98.6	98.9	98.8	97.3	99.0	98.7	98.6

BC TABLE 39

VIABILITY OF MACROPHAGES, RESUSPENSION MEDIUM

Remarks: trypan blue method, approx. 500 macrophages counted in hemocytometer
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages per pool.

20290273S2

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	~~MEAN~~ VIABILITY POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	3	3	3	3	3	3	3	3	3
	M (0/0)	97.00	97.70	97.50	98.97	99.87	99.60	98.37	99.60	99.67	99.33
	SE	1.60	0.67	1.01	0.34	0.07	0.15	0.03	0.12	0.07	0.03
	RSD (0/0)	2.9	1.2	1.80	0.6	0.1	0.3	0.1	0.2	0.1	0.1
1-GR	N	3	3	3	3	3	3	3	3	3	3
	M (0/0)	95.93	96.40	96.33	98.70	98.53	98.60	97.70	99.00	98.97	98.83
	SE	1.22	0.25	0.27	0.10	0.72	0.51	0.51	0.20	0.23	0.20
	RSD (0/0)	2.2	0.5	0.5	0.2	1.3	0.9	0.9	0.3	0.4	0.4
2-GR	x (%)	96.5	96.9	96.8	98.6	98.9	98.8	97.3	99.0	98.7	98.6

BC TABLE 40

~~MEAN~~ VIABILITY OF MACROPHAGES, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

Remarks: trypan blue method, approx. 500 macrophages counted in hemocytometer
Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages per pool.

20290273S3

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	NUMBER OF NONVIABLE MACROPHAGES/~~RAT (1E3)~~ (1E3/RAT)									
		POOL									
		1	2	1 + ~~and~~ 2	3	4	3 + ~~and~~ 4	5	6	7	5, 6 + ~~and~~ 7
0-GR	1	37	35	72	13	3	16	19	1	2	~~22~~ 23
	2	9	13	~~21~~22	5	0	5	11	6	8	~~25~~24
	3	7	7	14	2	3	~~8~~4	9	4	7	20
1-GR	1	20	23	42	11	10	~~20~~21	15	6	29	50
	2	9	27	36	6	10	~~16~~17	4	3	7	~~14~~15
	3	11	18	29	7	48	~~55~~56	5	11	25	41
2-GR	3	6	9	14	11	20	31	7	7	17	~~32~~31

BC TABLE 41

NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM

Remarks: absolute number of nonviable macrophages calculated on the basis of viability and absolute number of macrophages

20290273S4

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	MEAN NUMBER OF NONVIABLE MACROPHAGES/RAT (1E3)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	3	3	3	3	3	3	3	3	3
	M (1E3/RAT)	17.7	18.3	~~35.7~~ 36.0	6.7	2.0	8.~~7~~3	13.0	3.7	5.7	22.3
	SE	9.7	8.5	18.~~8~~1	3.3	1.0	3.~~7~~8	3.1	1.5	1.9	1.~~5~~2
	RSD (0/0)	94.9	80.4	~~88.8~~ 87.3	85.3	86.6	~~73.3~~ 79.9	40.7	68.6	56.7	~~11.3~~ 9.3
1-GR	N	3	3	3	3	3	3	3	3	3	3
	M (1E3/RAT)	13.3	22.7	35.7	8.0	22.7	~~30.3~~ 31.3	8.0	6.7	20.3	35.0~~3~~
	SE	3.4	2.6	3.8	1.5	12.7	12.4	3.5	2.3	6.8	10.~~8~~5
	RSD (0/0)	44.0	19.9	18.2	33.1	96.8	~~70.7~~ 68.5	76.0	60.6	57.6	~~53.5~~ 51.4
2-GR	x (1E3/RAT)	6	9	14	11	20	31	7	7	17	~~32~~31

BC TABLE 42

MEAN NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

Remarks: absolute number of nonviable macrophages calculated on the basis of viability and absolute number of macrophages

20290273S5

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF MULTINUCLEATED MACROPHAGES (0/0)									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	1	1.17	1.56	1.42	0.60	1.40	1.12	0.61	1.18	1.21	0.97
	2	1.02	0.96	0.98	0.82	-	-	0.96	1.38	1.57	1.40
	3	0.49	2.98	1.51	0.56	0.75	0.72	0.39	1.33	1.13	1.07
1-GR	1	2.30	6.19	4.98	7.53	4.65	5.74	6.28	4.43	7.16	6.39
	2	5.13	3.77	4.32	-	3.36	-	4.28	4.14	5.65	5.19
	3	2.96	7.22	5.73	7.23	7.72	7.59	6.36	2.50	2.18	2.74
2-GR	3	3.01	2.91	2.95	2.24	3.51	3.13	1.35	1.85	2.59	2.22

BC TABLE 43

RELATIVE NUMBER OF MULTINUCLEATED MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION

Remarks: Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages per pool.

20290273356

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	MEAN RELATIVE NUMBER OF MULTINUCLEATED MACROPHAGES									
		POOL									
		1	2	1 and 2	3	4	3 and 4	5	6	7	5, 6 and 7
0-GR	N	3	3	3	3	2	2	3	3	3	3
	M (0/0)	0.893	1.833	1.303	0.660	1.075	0.920	0.653	1.297	1.303	1.175
	SE	0.296	0.599	0.164	0.081	-	-	0.166	0.060	0.135	0.130
	RSD (0/0)	40.0	56.6	21.8	21.2	-	-	44.0	8.0	18.0	19.6
1-GR	N	3	3	3	2	3	2	3	3	3	3
	M (0/0)	3.463	5.727	5.010	7.380	5.243	6.650	5.640	3.690	4.995	4.773
	SE	0.858	1.023	0.407	-	1.293	-	0.680	0.601	1.474	1.074
	RSD (0/0)	42.8	30.9	14.1	-	42.7	-	20.9	28.2	51.1	39.0
2-GR	x (%)	3.01	2.91	2.95	2.24	3.51	3.13	1.35	1.85	2.59	2.22

BC TABLE 44

MEAN RELATIVE NUMBER OF MULTINUCLEATED MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, STATISTICAL PARAMETER

Remarks: Pool 1 and 2, 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages per pool.

20290273357

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	NUMBER OF MULTINUCLEATED MACROPHAGES/~~[illegible]~~ (RESPIRAT) POOL 1	2	1 + 2	3	4	3 + 4	5	6	7	5, 6 + 7
0-GR	1	6.9	15.6 (2.0) (a)	22.5	4.6	20.3	24.9	7.4	7.6	15.0	30.0
	2	6.7	10.6	17.4	6.3	–	–	6.4	13.8	29.7	49.9
	3	2.2	9.2	11.4	1.3	9.6	10.9	2.1	13.4 (~~2.0~~ 1.9) (a)	19.3	34.8
1-GR	1	7.1	42.7	49.8	52.7 (7.5) (a)	53.5	106.2	31.4	34.1	146.0 (4.2) (a)	211.5
	2	22.6	24.5	47.1	–	57.5	–	7.3	15.7	67.7 (3.2) (a)	90.7
	3	8.6	39.0	47.6	44.8	128.1	172.9	25.4	19.8	49.3 (4.9) (a)	94.5
2-GR	3	4.8	8.1	13.0	17.7	64.3	82.0	3.4 (0.7) (a)	13.1	34.0	50.4

BC TABLE 45

NUMBER OF MULTINUCLEATED MACROPHAGES, RESUSPENSION MEDIUM

Remarks: absolute number of multinucleated macrophages calculated from differential counts and absolute macrophage number

(a) number of macrophages with .GT.2 nuclei

20290273S8

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	MEAN NUMBER OF MULTINUCLEATED MACROPHAGES (~~HEC~~) POOL 1	2	1 + 2	3	4	3 + 4	5	6	7	5, 6 + 7
0-GR	N	3	3	3	3	2	2	3	3	3	3
	M (AE3)	5.27	11.80	17.10	4.07	15.0	17.90	5.30	11.60	21.33	38.23
	SE	1.53	1.94	3.21	1.47	-	-	1.63	2.00	4.36	6.00
	RSD (0/0)	50.5	28.5	32.5	62.5	-	-	53.1	29.9	35.4	27.2
1-GR	N	3	3	3	2	3	2	3	3	3	3
	M (AE3)	12.77	35.40	48.17	48.8	79.70	139.5	21.37	23.20	87.67	132.23
	SE	4.94	5.55	0.83	-	24.23	-	7.24	5.58	29.65	39.65
	RSD (0/0)	67.0	27.2	3.0	-	52.7	-	58.7	41.6	58.6	51.9
2-GR	x (AE3)	4.8	8.1	13.0	17.7	64.3	82.0	3.4	13.1	34.0	50.4

BC TABLE 46

MEAN NUMBER OF MULTINUCLEATED MACROPHAGES, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

Remarks: absolute number of multinucleated macrophages calculated from differential counts and absolute macrophage number

2029027359

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	~~MEAN~~ MACROPHAGE AREA (~~um2~~) POOL 3	4	3 ~~AND~~ 4 and	5	6	7	5, 6 ~~AND~~ 7 and
0-GR	N	107	133		94	187	188	
	M (μm^2)	204	211	210	232	214	227	224
	SE	6	5		5	4	5	
	RSD (0/0)	28.7	28.3		22.8	26.4	30.6	
1-GR	N	88	77		99	164	134	
	M (μm^2)	335	376	365	306	274	267	273
	SE	13	15		13	9	8	
	RSD (0/0)	37.2	35.8		43.1	42.3	34.2	
2-GR	N	100	90		81	68	81	
	M (μm^2)	242	267	259	201	262	297	275
	SE	8	10		8	12	10	
	RSD (0/0)	34.4	35.2		37.3	37.2	31.1	

BC TABLE 47

MEAN MACROPHAGE AREA, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

Remarks: Pool 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages/pool.

2029027360

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	DISTRIBUTION OF MACROPHAGES (0/0) AREA CLASS											
		0	1	2	3	4	5	6	7	8	9	10	11
0-GR	3	0.0	0.0	1.9	1.9	24.3	37.4	28.0	6.5	0.0	0.0	0.0	0.0
	4	0.0	0.0	0.0	2.3	18.8	45.9	24.8	6.8	1.5	0.0	0.0	0.0
	5	0.0	0.0	0.0	0.0	8.5	42.6	38.3	9.6	1.1	0.0	0.0	0.0
	6	0.0	0.0	0.0	0.5	19.8	42.2	29.4	7.5	0.5	0.0	0.0	0.0
	7	0.0	0.0	0.0	1.6	15.4	35.6	33.0	12.2	1.6	0.5	0.0	0.0
1-GR	3	0.0	0.0	0.0	1.1	3.4	12.5	26.1	38.6	10.2	6.8	1.1	0.0
	4	0.0	0.0	0.0	0.0	5.2	5.2	23.4	26.0	27.3	13.0	0.0	0.0
	5	0.0	0.0	0.0	1.0	5.1	24.2	28.3	24.2	11.1	5.1	1.0	0.0
	6	0.0	0.0	1.2	4.9	4.9	29.9	26.2	21.3	9.8	1.8	0.0	0.0
	7	0.0	0.0	0.0	1.5	8.2	23.9	41.8	15.7	9.0	0.0	0.0	0.0
2-GR	3	0.0	0.0	0.0	3.0	15.0	31.0	29.0	19.0	3.0	0.0	0.0	0.0
	4	0.0	0.0	0.0	2.2	12.2	20.0	33.3	25.6	5.6	1.1	0.0	0.0
	5	0.0	0.0	0.0	9.9	23.5	42.0	16.0	4.9	3.7	0.0	0.0	0.0
	6	0.0	0.0	0.0	4.4	5.9	27.9	33.8	20.6	5.9	1.5	0.0	0.0
	7	0.0	0.0	0.0	0.0	2.5	19.8	38.3	29.6	8.6	1.2	0.0	0.0

BC TABLE 48

RELATIVE DISTRIBUTION OF MACROPHAGES ACCORDING TO AREA

Remarks: (for number of cells per determination see BC TABLE 47.) classification on the basis of
10 equal steps on logarithmic scale
range: 50 to 1000 um2
classes 0 and 11 out of range

20290273361

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) A3 WS

GROUP	STATISTICAL PARAMETER	~~MEAN~~ NUCLEUS AREA (~~um2~~) POOL 3	4	3 and ~~AND~~ 4	5	6	7	5, 6 and ~~AND~~ 7
0-GR	N	107	134		95	192	192	
	M (μm^2)	60.6	64.7	64.1	64.7	64.7	65.7	65.2
	SE	1.1	0.8		1.0	0.8	0.7	
	RSD (0/0)	19.0	14.7		15.2	17.6	15.1	
1-GR	N	94	78		103	171	136	
	M (μm^2)	62.2	68.2	66.6	60.7	58.1	60.8	60.2
	SE	1.3	1.3		1.2	1.0	1.2	
	RSD (0/0)	20.3	17.2		19.6	23.2	23.5	
2-GR	N	100	90		84	69	81	
	M (μm^2)	61.8	66.2	64.9	60.5	63.6	70.7	67.4
	SE	1.4	1.2		1.3	1.3	1.2	
	RSD (0/0)	22.7	17.6		19.1	16.8	~~70.7~~ 15.4	

BC TABLE 49

MEAN NUCLEUS AREA, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

Remarks: Pool 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages/pool.

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	DISTRIBUTION OF NUCLEI (0/0) AREA CLASS											
		0	1	2	3	4	5	6	7	8	9	10	11
0-GR	3	0.0	0.9	3.7	10.3	27.1	33.6	13.1	5.6	3.7	0.0	0.0	1.9
	4	0.0	0.7	0.7	1.5	20.1	31.3	28.4	9.7	5.2	0.7	0.7	0.7
	5	0.0	0.0	0.0	5.3	16.8	32.6	30.5	8.4	2.1	1.1	3.2	0.0
	6	1.0	0.0	0.5	4.7	21.9	25.0	28.1	13.0	1.6	1.0	1.6	1.6
	7	0.0	0.5	0.0	2.1	18.8	28.1	32.8	11.5	2.6	1.0	1.6	1.0
1-GR	3	0.0	0.0	8.5	8.5	21.3	22.3	19.1	9.6	7.4	2.1	0.0	1.1
	4	0.0	0.0	0.0	2.6	15.4	29.5	19.2	14.1	10.3	5.1	2.6	1.3
	5	0.0	3.9	3.9	9.7	24.3	24.3	16.5	13.6	1.9	1.0	1.0	0.0
	6	1.2	2.3	13.5	14.6	17.5	24.6	13.5	5.8	5.3	1.2	0.0	0.6
	7	1.5	2.2	5.1	16.2	15.4	25.7	16.9	9.6	3.7	0.7	2.2	0.7
2-GR	3	0.0	3.0	7.0	10.0	19.0	20.0	22.0	10.0	4.0	3.0	0.0	2.0
	4	0.0	0.0	2.2	8.9	15.6	20.0	24.4	17.8	3.3	6.7	1.1	0.0
	5	0.0	0.0	6.0	16.7	21.4	28.6	9.5	9.5	4.8	3.6	0.0	0.0
	6	0.0	1.4	0.0	11.6	18.8	20.3	26.1	15.9	5.8	0.0	0.0	0.0
	7	0.0	0.0	0.0	2.5	11.1	16.0	25.9	24.7	12.3	4.9	1.2	1.2

BC TABLE 50

RELATIVE DISTRIBUTION OF NUCLEI ACCORDING TO AREA

Remarks: (for number of cells per determination see BC TABLE 47) classification on the basis of
10 equal steps on logarithmic scale
range: 30 to 100 um2
classes 0 and 11 out of range

20290273G3

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) A5 WS

GROUP	STATISTICAL PARAMETER	~~MEAN~~ VACUOLE AREA (~~um2~~) POOL 3	4	3 ~~AND~~ 4	5	6	7	5, 6 ~~AND~~ 7
0-GR	N	172	321		384	653	396	
	M (μm^2)	0.73	0.59	0.61	0.67	0.74	0.91	0.82
	SE	0.09	0.03		0.03	0.03	0.11	
	RSD (0/0)	163.2	94.3		110.7	110.7	230.7	
1-GR	N	392	646		997	1228	615	
	M (μm^2)	3.35	1.78	2.21	2.51	2.51	2.18	2.29
	SE	0.24	0.12		0.32	0.24	0.16	
	RSD (0/0)	141.8	167.3		401.7	330.8	181.1	
2-GR	N	553	524		315	391	488	
	M (μm^2)	0.99	1.04	1.02	1.18	0.90	0.97	0.97
	SE	0.12	0.07		0.10	0.10	0.06	
	RSD (0/0)	275.4	161.5		155.0	213.5	136.5	

BC TABLE 51

MEAN VACUOLE AREA, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

Remarks: Pool 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages/pool.

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) A6 WS

GROUP	STATISTICAL PARAMETER	~~MEAN~~ VACUOLE AREA PER MACROPHAGE ~~CELL~~ ~~(0/0)~~ POOL 3	4	3 and 4 ~~AND~~	5	6	7	5, 6 and 7 ~~AND~~
0-GR	N	172	321	-	384	653	396	-
	M (%)	0.53	0.63	0.61	1.08	1.13	0.67	0.88
	SE	-	-	-	-	-	-	
	RSD (0/0)	-	-	-	-	-	-	
1-GR	N	392	646	-	997	1228	615	
	M (%)	4.01	3.74	3.81	6.36	5.72	3.21	4.15
	SE	-	-	-	-	-	-	
	RSD (0/0)	-	-	-	-	-	-	
2-GR	N	553	524	-	315	391	488	
	M (%)	1.97	1.96	1.96	2.14	1.69	1.73	1.76
	SE	-	-	-	-	-	-	
	RSD (0/0)	-	-	-	-	-	-	

BC TABLE 52

~~MEAN~~ VACUOLE AREA PER MACROPHAGE ~~CELL~~, RESUSPENSION MEDIUM, STATISTICAL PARAMETERS

Remarks: Pool 3 and 4, 5, 6 and 7 represents the mean of these pools weighted according to the number of macrophages/pool.

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	DISTRIBUTION OF VACUOLES (0/0) AREA CLASS											
		0	1	2	3	4	5	6	7	8	9	10	11
0-GR	3	0.0	6.4	29.7	37.8	14.5	7.0	2.9	0.6	1.2	0.0	0.0	0.0
	4	0.0	1.9	25.2	47.0	20.2	2.5	2.8	0.3	0.0	0.0	0.0	0.0
	5	0.0	7.6	25.8	30.2	24.5	9.4	2.3	0.3	0.0	0.0	0.0	0.0
	6	0.0	2.1	23.3	39.1	21.3	10.7	2.9	0.5	0.2	0.0	0.0	0.0
	7	0.0	0.3	15.4	41.9	25.0	13.1	3.0	0.8	0.3	0.0	0.3	0.0
1-GR	3	0.0	0.3	3.3	13.5	23.0	24.5	13.3	9.7	8.4	3.3	0.8	0.0
	4	0.0	0.5	13.5	29.7	23.5	14.7	8.8	4.3	3.9	1.1	0.0	0.0
	5	0.0	0.6	9.7	25.8	24.6	17.9	10.0	6.0	3.7	0.7	0.7	0.3
	6	0.0	0.5	7.0	20.8	24.8	22.5	11.4	7.8	3.3	1.3	0.5	0.2
	7	0.0	0.2	7.8	21.1	25.7	22.9	11.1	6.2	2.8	1.8	0.5	0.0
2-GR	3	0.0	1.4	21.3	37.4	25.0	9.2	3.4	1.4	0.2	0.2	0.2	0.2
	4	0.0	3.4	21.4	25.6	26.9	14.5	5.5	1.7	0.6	0.4	0.0	0.0
	5	0.0	1.3	16.5	34.2	19.0	18.1	6.7	2.5	1.0	0.6	0.0	0.0
	6	0.0	3.6	19.2	35.8	23.8	13.3	3.6	0.0	0.5	0.0	0.3	0.0
	7	0.0	2.9	16.0	35.5	25.4	12.7	5.7	1.2	0.4	0.2	0.0	0.0

BC TABLE 53

DISTRIBUTION OF VACUOLES ACCORDING TO SIZE

Remarks: (for number of cells per determination see BC TABLE 47) classification on the basis of
10 equal steps on logarithmic scale
range: 0.1 to 50 um2
classes 0 and 11 out of range

20290273366

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	VIABILITY NUMBER OF MACROPHAGES VIABLE ABS.	VIABLE REL. (b) (0/0)	INTERMEDIATE (a) ABS.	INTERMEDIATE (a) REL. (b) (0/0)	NONVIABLE ABS.	NONVIABLE REL. (b) (0/0)	TOTAL
0-GR	1	1	4589	51.1	2297	25.6	2096	23.3	8982
		3	32880	90.5	1406	3.9	2037	5.6	36323
	2	1	5957	36.9	7751	48.1	2417	15.0	16125
		3	18044	91.2	650	3.3	1093	5.5	19787
	3	1	6242	95.7	71	1.1	212	3.2	6525
		3	19354	96.5	314	1.6	398	2.0	20066
	4	1	12915	92.9	209	1.5	779	5.6	13903
		3	22216	95.0	488	2.1	686	2.9	23390
	5	1	2559	89.5	113	4.0	186	6.5	2858
		3	21998	96.8	285	1.3	438	1.9	22721

BC TABLE 54 (FIGURE HO 3356, 3524, 3525, 3526, 3527)

VIABILITY OF MACROPHAGES DETERMINED BY FCM, INDIVIDUAL VALUES

Remarks: total number of signals analyzed: 19400 to 67400
no data from experiment 2 due to laser failure

(a) for explanation see ...
(b) to total

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	VIABILITY NUMBER OF MACROPHAGES VIABLE ABS.	VIABLE REL. (b) (0/0)	INTERMEDIATE (a) ABS.	INTERMEDIATE (a) REL. (b) (0/0)	NONVIABLE ABS.	NONVIABLE REL. (b) (0/0)	TOTAL
0-GR	6	1	10208	92.8	279	2.5	513	4.7	11000
		3	15769	93.1	503	3.0	672	4.0	16944
	7	1	4072	95.1	88	2.1	120	2.8	4280
		3	15708	93.8	334	2.0	709	4.2	16751
1-GR	1	1	1039	50.8	417	20.4	588	28.8	2044
		3	11563	86.8	877	6.6	884	6.6	13324
	2	1	5075	59.2	1735	20.2	1764	20.6	8574
		3	15637	90.4	634	3.7	1033	6.0	17304
	3	1	2682	82.7	353	10.9	209	6.4	3244
		3	7393	94.8	169	2.2	238	3.1	7800

BC TABLE 54 (continued)

VIABILITY OF MACROPHAGES DETERMINED BY FCM, INDIVIDUAL VALUES

Remarks: total number of signals analyzed: 19400 to 67400
no data from experiment 2 due to laser failure

(a) for explanation see ...
(b) to total

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	VIABILITY NUMBER OF MACROPHAGES VIABLE ABS.	VIABLE REL. (b) (0/0)	INTERMEDIATE (a) ABS.	INTERMEDIATE (a) REL. (b) (0/0)	NONVIABLE ABS.	NONVIABLE REL. (b) (0/0)	TOTAL
1-GR	4	1	13226	87.6	655	4.3	1214	8.0	15095
		3	14333	91.2	466	3.0	916	5.8	15715
	5	1	599	51.8	245	21.2	312	27.0	1156
		3	6805	87.7	421	5.4	529	6.8	7755
	6	1	12333	83.5	957	6.5	1479	10.0	14769
		3	23273	92.3	745	3.0	1189	4.7	25207
	7	1	17949	85.4	1340	6.4	1735	8.3	21024
		3	20285	90.7	775	3.5	1306	5.8	22366
2-GR	1	3	13177	91.5	484	3.4	747	5.2	14408
	2	3	18112	91.0	643	3.2	1158	5.8	19913

BC TABLE 54 (continued)

VIABILITY OF MACROPHAGES DETERMINED BY FCM, INDIVIDUAL VALUES

Remarks: total number of signals analyzed: 19400 to 67400
no data from experiment 2 due to laser failure

(a) for explanation see ...
(b) to total

2029022369

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	VIABILITY NUMBER OF MACROPHAGES VIABLE ABS.	VIABLE REL. (b) (0/0)	INTERMEDIATE (a) ABS.	INTERMEDIATE REL. (b) (0/0)	NONVIABLE ABS.	NONVIABLE REL. (b) (0/0)	TOTAL
2-GR	3	3	16062	86.3	1735	9.3	824	4.4	18621
	4	3	15083	87.1	1326	7.7	905	5.2	17314
	5	3	19154	86.6	1803	8.1	1172	5.3	22129
	6	3	12988	88.8	1111	7.6	531	3.6	14630
	7	3	13096	87.1	758	5.0	1190	7.9	15044

BC TABLE 54 (continued)

VIABILITY OF MACROPHAGES DETERMINED BY FCM, INDIVIDUAL VALUES

Remarks: total number of signals analyzed: 19400 to 67400
no data from experiment 2 due to laser failure

(a) for explanation see
(b) to total

20290273?0

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) A12 WS

GROUP	POOL	EXPERIMENT NO.	RELATIVE NUMBER OF MACROPHAGES (0/0)		
			VIABLE	INTERMEDIATE	NONVIABLE
0-GR	1 and 2	1	42.2	39.7	18.1
		3	90.8	3.6	5.6
	3 and 4	1	93.8	1.4	4.8
		3	95.2	2.0	2.8
	5, 6 and 7	1	92.5	2.9	4.6
		3	94.1	2.2	3.8
1-GR	1 and 2	1	56.6	20.3	23.1
		3	89.1	4.7	6.2
	3 and 4	1	85.8	6.8	7.4
		3	92.2	2.8	5.1
	5, 6 and 7	1	79.9	8.6	11.5
		3	90.7	3.6	5.7
2-GR	1 and 2	3	91.1	3.3	5.6
	3 and 4	3	86.9	8.2	5.0
	5, 6 and 7	3	87.5	6.2	6.3

BC TABLE 55 [Figure HO 3355, 3616, 3617]

RELATIVE NUMBER OF VIABLE, INTERMEDIATE AND NONVIABLE MACROPHAGES, WEIGHTED MEAN OF POOLS

Remarks: mean weighted according to number of macrophages/pool/5 rats, re- ~~(see BC TABLE ...)~~ suspension medium, hemocytometer

keine Tabelle mit diesen Werten, nur auf Tab. Protokoll

2029027371

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	GREEN FLUORESCENCE SIGNALS			
		N	MEAN (CH. NO.)	CV (0/0)	MEAN REL. TO 0-GR (a) (0/0)
0-GR	1	30598	296.7	47.4	100
	2	17842	268.1	49.4	100
	3	17877	328.5	47.4	100
	4	21995	304.3	48.6	100
	5	20648	255.6	49.9	100
	6	15570	287.9	52.7	100
	7	15665	291.7	51.8	100
1-GR	1	11329	379.3	- (b)	127.8
	2	15797	406.2	- (b)	151.5
	3	7367	379.4	- (b)	115.5
	4	14404	367.9	- (b)	120.9
	5	6732	306.0	56.4	119.7
	6	23583	353.5	- (b)	122.8
	7	20296	385.8	- (b)	132.3
2-GR	1	12963	280.3	60.7	94.5
	2	18219	289.5	61.8	108.0
	3	15895	271.1	58.8	82.5
	4	14810	291.5	55.4	95.8
	5	18126	243.4	57.2	95.2
	6	12945	320.6	53.6	111.4
	7	12880	306.7	51.6	105.1

BC TABLE 56

GREEN FLUORESCENCE SIGNALS FROM MACROPHAGES CORRESPONDING TO ESTERASE ACTIVITY, EXPERIMENT 3

Remarks: .LE.2.8 0/0 of signals out of range, included in calculation of mean

(a) corresponding pool
(b) CV not meaningful, .GT.1 0/0 of signals out of range

20290272372

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) A14 WS

GROUP	~~WEIGHTED~~ MEAN ESTERASE ACTIVITY					
	POOL					
	ABS. (CH. NO.)			REL. TO 0-GR (0/0) (a)		
	1 ~~AND~~ 2	3 AND 4	5, 6 AND 7	1 AND 2	3 AND 4	5, 6 AND 7
0-GR	284.9	308.0	284.6	100	100	100
1-GR	396.9	371.0	369.2	139.3	120.5	129.7
2-GR	286.1	285.4	304.1	100.4	92.7	106.9

BC TABLE 57

ESTERASE ACTIVITY OF MACROPHAGES, WEIGHTED MEAN OF POOLS, EXPERIMENT 3 [ES. Tab 57]

Remarks: mean of pools weighted according to number of macrophages/pool

(a) corresponding pool

20290273?3

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	RATIO GNF/AXL N	MEAN (CH. NO.)	CV (0/0)	MEAN REL. TO 0-GR (a) (0/0)
0-GR	1	30597	268.9	42.2	100
	2	17842	237.7	43.6	100
	3	17877	295.9	41.7	100
	4	21995	274.6	42.8	100
	5	20666	229.9	46.0	100
	6	15572	264.5	49.5	100
	7	15667	267.2	48.4	100
1-GR	1	11327	258.4	43.1	96.1
	2	15796	287.2	41.9	120.8
	3	7365	267.1	41.5	90.3
	4	14397	272.5	41.6	99.2
	5	6730	214.8	40.6	93.4
	6	23577	260.5	40.8	98.5
	7	20296	286.7	40.5	107.3
2-GR	1	12960	232.6	54.2	86.5
	2	18216	239.1	53.7	100.6
	3	15911	231.8	51.6	78.3
	4	14815	253.4	48.4	92.3
	5	18123	193.2	50.7	84.0
	6	12946	275.6	47.4	104.2
	7	12880	265.9	46.0	99.5

BC TABLE 58

RATIO OF GREEN FLUORESCENCE/AXL, MACROPHAGES, ESTERASSE ACTIVITY, EXPERIMENT 3

Remarks: ratio calculated on a cell by cell basis

(a) corresponding pool

202902737A

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) A16 WS

GROUP	STATISTICAL PARAMETER					
	ESTERASE ACTIVITY					
	GREEN FLUORESCENCE			RATIO GREEN FLUORESCENCE/AXL		
	M (CH. NO.)	SE	RSD (0/0)	M (CH. NO.)	SE	RSD (0/0)
0-GR	292.50	7.76	4.6	264.71	6.69	4.4
1-GR	379.02	8.94	4.1	273.54	1.87	1.2
2-GR	291.88	6.13	3.6	248.21	7.04	4.9

BC TABLE 59

ESTERASE ACTIVITY OF MACROPHAGES, WEIGHTED MEAN OF POOLS, EXPERIMENT 3, STATISTICAL PARAMETERS

(Figure 403357)

20290273?5

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	WEIGHTED MEAN RATIO POOL					
	ABS. (CH. NO.)			REL. TO 0-GR (0/0) (a)		
	1 AND 2	3 AND 4	5, 6 AND 7	1 AND 2	3 AND 4	5, 6 AND 7
0-GR	256.0	277.9	260.3	100	100	100
1-GR	277.2	271.0	272.4	108.3	97.5	104.7
2-GR	236.7	246.9	261.0	92.5	88.9	100.3

BC TABLE 60

RATIO OF GREEN FLUORESCENCE/AXL, MACROPHAGES, ESTERASE ACTIVITY, EXPERIMENT 3, WEIGHTED MEAN OF POOLS

Remarks: mean of pools weighted according to number of macrophages/pool

(a) corresponding pool

20290273?6

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	CONCENTRATION OF MACROPHAGES (1E6/ml) POOL			VIABILITY (0/0)		
		8	9	10	8	9	10
0-GR	1	0.67	1.12	0.93	75.6	99.6	98.5
	2	1.00	0.83	1.02	95.3	97.5	97.4
	3	-	-	1.19	-	-	99.8
1-GR	1	-	0.93	0.83	-	97.0	98.2
	2	-	0.88	1.15	-	96.3	96.4 -> 96.4
	3	-	0.82	0.95	-	97.1	97.8
2-GR	3	-	0.83	1.05	-	96.4	96.1

BC TABLE 61

CONCENTRATION AND VIABILITY OF MACROPHAGES AFTER PHAGOCYTOSIS

Remarks: determination immediately after 40 min incubation
nominal concentration of macrophages at start of phagocytosis experiment: 1E6/ml
viability determined by trypan blue method

2029027377

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF MACROPHAGES						
			NONPHAGOCYTIZING		PHAGOCYTIZING				
					NO. OF BEADS/CELL				TOTAL
					.GE.1		.GE.3		
			ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	
0-GR	8	1 (c)	8215	85.0	1445	15.0	763	7.9	9660
		1 (d)	3364	72.9	1252	27.1	690	14.9	4616
		2	7626	69.1	3414	30.9	1877	17.0	11040
		3	-	-	-	-	-	-	-
	9	1	8402	73.9	2963	26.1	1564	13.8	11365
		2	6884	64.2	3832	35.8	2250	21.0	10716
		3	-	-	-	-	-	-	-
	10	1	8038	77.1	2381	22.9	1259	12.1	10419
		2	7488	61.1	4770	38.9	2554	20.8	12258
		3	10726	55.8	8491	44.2	4545	23.7	19217

BC TABLE 62

NUMBER OF NONPHAGOCYTIZING AND PHAGOCYTIZING MACROPHAGES, INDIVIDUAL VALUES

Remarks: total number of signals counted: 80000

(a) per sample determined by FCM
(b) to total
(c) atypical FWD versus AXL distribution of macrophages (see) values for total macrophages
(d) values for macrophage subpopulation at approx. typical location in FWD versus AXL cytogram

20290273?8

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF MACROPHAGES						
			NONPHAGOCYTIZING		PHAGOCYTIZING				TOTAL
					NO. OF BEADS/CELL				
					.GE.1		.GE.3		
			ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	
1-GR	8	1	-	-	-	-	-	-	-
		2	-	-	-	-	-	-	-
		3	-	-	-	-	-	-	-
	9	1	10044	48.1	10850	51.9	7103	34.0	20894
		2	4671	47.1	5248	52.9	3304	33.3	9919
		3	3514	42.2	4809	57.8	3084	37.1	8323
	10	1	9847	43.0	13056	57.0	8296	36.2	22903
		2	7286	42.7	9773	57.3	6197	36.3	17059
		3	8550	41.2	12205	58.8	7827	37.7	20755

BC TABLE 62 (continued)

NUMBER OF NONPHAGOCYTIZING AND PHAGOCYTIZING MACROPHAGES, INDIVIDUAL VALUES

Remarks: total number of signals counted: 80000

(a) per sample determined by FCM
(b) to total

20290273?9

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF MACROPHAGES						
			NONPHAGOCYTIZING		PHAGOCYTIZING				
					NO. OF BEADS/CELL				TOTAL
					.GE.1		.GE.3		
			ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	
2-GR	8	3	-	-	-	-	-	-	-
	9	3	7783	46.2	9076	53.8	5670	33.6	16859
	10	3	7361	50.3	7287	49.7	4186	28.6	14648

BC TABLE 62 (continued)

NUMBER OF NONPHAGOCYTIZING AND PHAGOCYTIZING MACROPHAGES, INDIVIDUAL VALUES

Remarks: total number of signals counted: 80000

(a) per sample determined by FCM
(b) to total

2029027380

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF MACROPHAGES POOL 8	9	10
0-GR	N	2	2	3
	M (0/0)	77.1 (a)	69.1	64.7
	SE	-	-	6.4
	RSD (0/0)	-	-	17.2
1-GR	N	-	3	3
	M (0/0)	-	45.8	42.3
	SE	-	1.8	0.6
	RSD (0/0)	-	6.9	2.3
2-GR	N	-	1	1
	M (0/0)	-	46.2	50.3
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 63

RELATIVE NUMBER OF NONPHAGOCYTIZING MACROPHAGES, STATISTICAL PARAMETERS

Remarks: see BC TABLE 62 and BC FIGURE [vergl. Tab. 63] [H03353]

2029027381

(a) mean calculated using value for total macrophages, experiment 1 (see BC TABLE 62), mean value using macrophage subpopulation: 71.0

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF MACROPHAGES		
		POOL		
		8	9	10
0-GR	N	2	2	3
	M (0/0)	23.0 (a)	31.0	35.3
	SE	-	-	6.4
	RSD (0/0)	-	-	31.4
1-GR	N	-	3	3
	M (0/0)	-	54.2	57.7
	SE	-	1.8	0.6
	RSD (0/0)	-	5.8	1.7
2-GR	N	-	1	1
	M (0/0)	-	53.8	49.7
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 64

RELATIVE NUMBER OF MACROPHAGES PHAGOCYTIZING .GE.1 BEAD, STATISTICAL PARAMETERS

Remarks: see BC TABLE 62 and BC FIGURE

[H03513]

(a) mean calculated using value for total macrophages, experiment 1 (see BC TABLE 62), mean value using macrophage subpopulation: 29.0

2029027382

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF MACROPHAGES POOL 8	9	10
0-GR	N	2	2	3
	M (0/0)	12.5 (a)	17.4	18.9
	SE	-	-	3.5
	RSD (0/0)	-	-	32.0
1-GR	N	-	3	3
	M (0/0)	-	34.8	36.7
	SE	-	1.2	0.5
	RSD (0/0)	-	5.8	2.3
2-GR	N	-	1	1
	M (0/0)	-	33.6	28.6
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 65

RELATIVE NUMBER OF MACROPHAGES PHAGOCYTIZING .GE.3 BEADS, STATISTICAL PARAMETERS

Remarks: see BC TABLE 62 and BC FIGURE [H03512]

(a) mean calculated using value for total macrophages, experiment 1 (see BC TABLE 62), mean value using macrophage subpopulation: 16.0

2029027383

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	MEAN FLUORESCENCE (CH. NO.) NUMBER OF BEADS PER MACROPHAGE 0	1	.GE.0	MEAN NUMBER OF BEADS PER MACROPHAGE
0-GR	8	1 (a)	15.4	51.2	45.6	0.84
		1 (b)	15.7	51.2	73.5	1.63
		2	11.3	48.2	82.9	1.94
		3	-	-	-	-
	9	1	12.4	48.6	65.1	1.46
		2	11.3	48.5	104.8	2.51
		3	-	-	-	-
	10	1	11.4	48.8	61.4	1.34
		2	11.3	48.0	93.9	2.25
		3	10.3	55.6	115.6	2.32
1-GR	8	1	-	-	-	-
		2	-	-	-	-
		3	-	-	-	-
	9	1	11.7	46.9	164.4	4.34
		2	9.3	44.6	162.8	4.35
		3	11.8	54.9	188.1	4.09
	10	1	6.5	41.2	171.9	4.77
		2	7.8	42.7	156.1	4.25
		3	10.7	53.3	200.8	4.46
2-GR	8	3	-	-	-	-
	9	3	12.9	56.7	198.5	4.24
	10	3	9.9	53.5	158.3	3.40

BC TABLE 66

MEAN NUMBER OF PHAGOCYTIZED BEADS PER MACROPHAGE, INDIVIDUAL VALUES

Remarks: calculation: (a - b)/(c - b),
a = mean fluorescence, .GE.0 beads per macrophage
b = mean fluorescence, 0 beads per macrophage
c = mean fluorescence, 1 bead per macrophage

(a) atypical FWD versus AXL distribution of macrophages (see), values for total macrophages
(b) values for macrophage subpopulation at approx. typical location in FWD versus AXL cytogram

2029027384

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	MEAN NUMBER OF BEADS PER MACROPHAGE		
		POOL		
		8	9	10
0-GR	N	2	2	3
	M	1.39 (a)	1.99	1.97
	SE	-	-	0.32
	RSD (0/0)	-	-	27.8
1-GR	N	-	3	3
	M	-	4.26	4.49
	SE	-	0.09	0.15
	RSD (0/0)	-	3.5	5.8
2-GR	N	-	1	1
	M	-	4.24	3.40
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 67

MEAN NUMBER OF PHAGOCYTIZED BEADS PER MACROPHAGE, STATISTICAL PARAMETERS

Remarks: see BC TABLE 66 and BC FIGURE

[H0335-2]

(a) mean calculated using value for total macrophages, experiment 1 (see BC TABLE 66), mean value using macrophage subpopulation: 1.79

2029027385

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF GRANULOCYTES						
			NONPHAGOCYTIZING		PHAGOCYTIZING				
					NO. OF BEADS/CELL				TOTAL
					.GE.1		.GE.3		
			ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	
0-GR	8	1 (c)	-	-	-	-	-	-	-
		2	423	94.4	25	5.6	16	3.6	448
		3	-	-	-	-	-	-	-
	9	1	542	92.2	46	7.8	33	5.6	588
		2	237	91.2	23	8.8	19	7.3	260
		3	-	-	-	-	-	-	-
	10	1 (d)	-	-	-	-	-	-	-
		2	116	89.9	13	10.1	9	7.0	129
		3	65	64.4	36	35.6	31	30.7	101

BC TABLE 68

NUMBER OF NONPHAGOCYTIZING AND PHAGOCYTIZING GRANULOCYTES, INDIVIDUAL VALUES

Remarks: total number of signals counted: 80000

(a) per sample determined by FCM
(b) to total
(c) evaluation not possible, atypical FWD versus AXL distribution (see BC FIGURE ..)
(d) insufficient number of cells

20290273B6

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF GRANULOCYTES						
			NONPHAGOCYTIZING		PHAGOCYTIZING				
					NO. OF BEADS/CELL				TOTAL
					.GE.1		.GE.3		
			ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	ABS. (a)	REL. (b) (0/0)	
1-GR	8	1	-	-	-	-	-	-	-
		2	-	-	-	-	-	-	-
		3	-	-	-	-	-	-	-
	9	1	8095	46.6	9278	53.4	6088	35.0	17373
		2	5369	48.4	5732	51.6	3614	32.6	11101
		3	6951	39.3	10740	60.7	6560	37.1	17691
	10	1	2746	46.6	3145	53.4	2217	37.6	5891
		2	5195	42.5	7032	57.5	4581	37.5	12227
		3	3796	37.7	6265	62.3	3970	39.5	10061
2-GR	8	3	-	-	-	-	-	-	-
	9	3	486	40.4	718	59.6	519	43.1	1204
	10	3	192	38.7	304	61.3	233	47.0	496

BC TABLE 68 (continued)

NUMBER OF NONPHAGOCYTIZING AND PHAGOCYTIZING GRANULOCYTES, INDIVIDUAL VALUES

Remarks: total number of signals counted: 80000

(a) per sample determined by FCM
(b) to total

20290273B7

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF GRANULOCYTES		
		POOL		
		8	9	10
0-GR	N	1	2	2
	M (0/0)	94.4	91.7	77.2
	SE	-	-	-
	RSD (0/0)	-	-	-
1-GR	N	-	3	3
	M (0/0)	-	44.77	42.27
	SE	-	2.78	2.57
	RSD (0/0)	-	10.8	10.5
2-GR	N	-	1	1
	M (0/0)	-	40.4	38.7
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 69

RELATIVE NUMBER OF NONPHAGOCYTIZING GRANULOCYTES, STATISTICAL PARAMETERS

Remarks: see BC TABLE 68. and BC FIGURE

[H03353]

2029027388

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF GRANULOCYTES		
		POOL		
		8	9	10
0-GR	N	1	2	2
	M (0/0)	5.6	8.3	22.9
	SE	-	-	-
	RSD (0/0)	-	-	-
1-GR	N	-	3	3
	M (0/0)	-	55.2	57.7
	SE	-	2.8	2.6
	RSD (0/0)	-	8.7	7.7
2-GR	N	-	1	1
	M (0/0)	-	59.6	61.3
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 70

RELATIVE NUMBER OF GRANULOCYTES PHAGOCYTIZING .GE.1 BEAD, STATISTICAL PARAMETERS

Remarks: see BC TABLE 69 and BC FIGURE

[403513]

2029027389

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF GRANULOCYTES POOL 8	9	10
0-GR	N	1	2	2
	M (0/0)	3.6	6.5	18.9
	SE	-	-	-
	RSD (0/0)	-	-	-
1-GR	N	-	3	3
	M (0/0)	-	34.9	38.2
	SE	-	1.3	0.7
	RSD (0/0)	-	6.5	3.0
2-GR	N	-	1	1
	M (0/0)	-	43.1	47.0
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 71

RELATIVE NUMBER OF GRANULOCYTES PHAGOCYTIZING .GE.3 BEADS, STATISTICAL PARAMETERS

Remarks: see BC TABLE 68 and BC FIGURE

[H03512]

2029027390

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	MEAN FLUORESCENCE (CH. NO.) NUMBER OF BEADS PER GRANULOCYTE 0	1	.GE.0	MEAN NUMBER OF BEADS PER GRANULOCYTE
0-GR	8	1 (a)	-	-	-	-
		2	6.1	42.8	14.5	0.23
		3	-	-	-	-
	9	1	7.1	45.1	27.6	0.54
		2	6.3	42.0	31.2	0.70
		3	-	-	-	-
	10	1 (a)	-	-	-	-
		2	7.8	42.0	31.1	0.68
		3	8.2	55.6	137.5	2.73
1-GR	8	1	-	-	-	-
		2	-	-	-	-
		3	-	-	-	-
	9	1	8.2	45.7	123.8	3.08
		2	7.5	45.3	108.4	2.67
		3	9.3	55.1	136.3	2.77
	10	1	8.4	46.0	142.1	3.56
		2	8.1	45.4	126.0	3.16
		3	9.7	54.8	168.2	3.51
2-GR	8	3	-	-	-	-
	9	3	8.4	54.0	170.4	3.55
	10	3	8.5	53.0	188.7	4.05

BC TABLE 72

MEAN NUMBER OF PHAGOCYTIZED BEADS PER GRANULOCYTE, INDIVIDUAL VALUES

Remarks: for calculation see BC TABLE 66

(a) evaluation not possible, atypical FWD versus AXL distribution (see BC FIGURE ..)

20290273 91

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	MEAN NUMBER OF BEADS PER GRANULOCYTE		
		POOL		
		8	9	10
0-GR	N	1	2	2
	M	0.23	0.62	1.71
	SE	-	-	-
	RSD (0/0)	-	-	-
1-GR	N	-	3	3
	M	-	2.84	3.41
	SE	-	0.12	0.13
	RSD (0/0)	-	7.53	6.39
2-GR	N	-	1	1
	M	-	3.55	4.05
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 73

MEAN NUMBER OF PHAGOCYTIZED BEADS PER GRANULOCYTE, STATISTICAL PARAMETERS

Remarks: see BC TABLE 72 and BC FIGURE

[H03352]

2029027392

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXPERIMENT NO.	AUTOFLUORESCENCE SIGNALS		
			N	MEAN (a)	CV (0/0)
0-GR	8	1	8133	0.41	22.5
		2	4696	0.29	20.9
		3	-	-	-
	9	1	6371	0.37	21.8
		2	4186	0.29	19.9
		3	-	-	-
	10	1	9631	0.37	19.6
		2	6347	0.29	20.1
		3	8513	0.31	23.3
1-GR	8	1	-	-	-
		2	-	-	-
		3	-	-	-
	9	1	6715	0.53	40.4
		2	2933	0.47	39.9
		3	4933	0.51	53.2
	10	1	6513	0.54	36.3
		2	4440	0.49	44.5
		3	7676	0.51	51.7
2-GR	8	3	-	-	-
	9	3	6122	0.36	32.2
	10	3	7876	0.38	33.4

BC TABLE 74

AUTOFLUORESCENCE SIGNALS FROM MACROPHAGES, PHAGOCYTOSIS ASSAY

Remarks: sample taken at time = 0 of incubation
macrophages associated with latex beads (mean: 12.9 0/0, RSD: 47.5 0/0) excluded from evaluation

(a) normalized to fluorescence of standard latex beads

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXPERIMENT NO.	RATIO AUTOFLUORESCENCE/AXL		
			N	MEAN (a)	CV (0/0)
0-GR	8	1 (b)	-	-	-
		2	5533	10.2	16.0
		3	-	-	-
	9	1	6336	12.4	16.2
		2	4235	11.1	15.8
		3	-	-	-
	10	1	9540	12.5	13.6
		2	6397	11.0	15.0
		3	8510	15.2	16.8
1-GR	8	1	-	-	-
		2	-	-	-
		3	-	-	-
	9	1	6515	13.9	23.3
		2	2932	14.1	24.5
		3	4961	18.9	31.2
	10	1	6503	14.5	21.9
		2	4394	14.5	25.0
		3	7720	19.1	30.1
2-GR	8	3	-	-	-
	9	3	6106	16.4	20.4
	10	3	7835	17.0	19.3

BC TABLE 75

RATIO AUTOFLUORESCENCE/AXL OF MACROPHAGES, PHAGOCYTOSIS ASSAY

Remarks: ratio calculated for each macrophage
sample taken at time = 0 of incubation
macrophages associated with latex beads (mean: 12.9 0/0, RSD: 47.5 0/0) excluded from evaluation

(a) normalized to standard latex beads
(b)

2029027394

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXPERIMENT NO.	RED FLUORESCENCE SIGNAL AREA			
			N	MEAN		
				ABSOLUTE (CH. NO.)	RELATIVE TO G0/1 OF	
					MACROPHAGES	GRANULOCYTES
0-GR	8	1	1069	410.7	1.85	2.09
		2	719	436.7	1.83	2.11
		3	178	508.9	1.82	2.13
	9	1	744	399.2	1.83	2.09
		2	693	434.1	1.83	2.12
		3	586	522.2	1.84	2.15
	10	1	1024	407.8	1.83	2.10
		2	634	431.6	1.84	2.09
		3	743	528.6	1.82	2.08
1-GR	8	1	602	442.2	1.80	2.23
		2	290	428.5	1.81	2.17
		3	591	565.1	1.79	2.24
	9	1	1069	422.7	1.78	2.16
		2	444	470.9	1.83	2.29
		3	493	512.6	1.76	2.18
	10	1	1087	426.2	1.79	2.18
		2	740	477.2	1.83	2.31
		3	407	529.0	1.75	2.17
2-GR	8	3	-	-	-	-
	9	3	607	510.0	1.81	2.13
	10	3	540	543.6	1.82	2.15

BC TABLE 76

RED FLUORESCENCE SIGNAL AREA OF MACROPHAGES IN G2 (a), DNA ASSAY

(a) region (see BC FIGURE ..)

20290273395

Source: https://www.industrydocuments.ucsf.edu/docs/ynd10000

GROUP	POOL	STATISTICAL PARAMETER			
		CV			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	7.83	0.55	12.1
	9	3	7.47	0.44	10.1
	10	3	7.30	0.42	9.9
	8 to 10	9	7.53	0.25	9.9
1-GR	8	3	12.17	0.47	6.6
	9	3	11.63	0.44	6.5
	10	3	11.60	0.47	7.1
	8 to 10	9	11.80	0.25	6.3
2-GR	8	-	-	-	-
	9	1	8.6	-	-
	10	1	9.2	-	-
	9 and 10	2	8.9	-	-

BC TABLE 77

CV OF RED FLUORESCENCE SIGNAL AREA DISTRIBUTION OF MACROPHAGES IN G0/1, DNA ASSAY, STATISTICAL PARAMETERS

2029027396

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	RED FLUORESCENCE SIGNALS (a)				
			N	AREA		HEIGHT	
				MEAN (CH. NO.)	CV (0/0)	MEAN (CH. NO.)	CV (0/0)
0-GR	8	1	517	196.6	3.3	296.1	6.2
		2	808	206.7	4.0	368.6	8.9
		3	173	238.4	3.4	466.6	5.8
	9	1	326	190.9	3.4	288.6	6.3
		2	942	204.7	4.1	372.5	7.7
		3	718	243.1	3.8	471.4	5.8
	10	1	163	193.8	3.6	291.5	6.5
		2	589	206.3	4.0	375.1	7.2
		3	672	254.0	4.0	476.6	7.4
1-GR	8	1	13927	198.1	3.4	298.5	7.1
		2	15039	197.6	3.9	363.7	8.1
		3	13814	252.93	3.8	478.2	7.9
	9	1	10627	195.7	3.5	294.6	7.0
		2	18624	206.0	3.9	373.6	8.3
		3	19390	234.9	4.5	452.4	8.5
	10	1	4380	195.5	3.7	292.1	7.2
		2	13598	206.4	3.7	371.6	8.0
		3	8216	243.8	4.3	467.0	8.2
2-GR	8	3	-	-	-	-	-
	9	3	4397	239.0	4.0	458.3	7.9
	10	3	3101	253.4	4.0	474.4	8.3

BC TABLE 78

RED FLUORESCENCE SIGNALS OF GRANULOCYTES, DNA ASSAY

(a) region 3 in BC FIGURE

20290273397

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	RELATIVE MEAN RED FLUORESCENCE SIGNAL (0/0) AREA	HEIGHT
1-GR	8	1	100.8	100.8
		2	95.6	98.7
		3	105.8	102.5
	9	1	102.5	102.1
		2	100.6	100.3
		3	96.6	96.0
	10	1	100.9	100.2
		2	100.0	99.1
		3	96.0	98.0
2-GR	8	3	-	-
	9	3	98.3	97.2
	10	3	99.8	99.5

BC TABLE 79

MEAN RED FLUORESCENCE OF GRANULOCYTES, 1-GR AND 2-GR, RELATIVE TO 0-GR, DNA ASSAY, INDIVIDUAL VALUES

[1] Remarks: see BC TABLE 78

202902739B

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER						
		RELATIVE MEAN RED FLUORESCENCE SIGNAL						
		N	AREA			HEIGHT		
			M (0/0)	SE	RSD (0/0)	M (0/0)	SE	RSD (0/0)
1-GR	8	3	100.7	3.0	5.1	100.7	1.1	1.9
	9	3	99.9	1.7	3.0	99.4	1.8	3.2
	10	3	99.0	1.5	2.6	99.1	0.6	1.1
	8 to 10	9	99.9	1.1	3.3	99.7	0.7	2.1
2-GR	8	-	-	-	-	-	-	-
	9	1	98.3	-	-	97.2	-	-
	10	1	99.8	-	-	99.6	-	-
	9 and 10	2	99.0	-	-	98.4	-	-

BC TABLE 80

MEAN RED FLUORESCENCE OF GRANULOCYTES, 1-GR AND 2-GR, RELATIVE TO 0-GR, DNA ASSAY, STATISTICAL PARAMETERS

[?] Remarks: see BC TABLE 79

20290273399

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) B12 WS

GROUP	POOL 1ST	POOL 2ND	RELATIVE MEAN RED FLUORESCENCE SIGNAL (0/0) (a) EXPERIMENT NO. AREA 1	AREA 2	AREA 3	HEIGHT 1	HEIGHT 2	HEIGHT 3
0-GR	8	9	103.0	101.0	98.1	102.6	99.0	99.0
	8	10	101.4	100.2	93.9	101.6	98.3	97.9
	9	10	98.5	99.2	95.7	99.0	99.3	98.9
1-GR	8	9	101.2	95.9	107.4	101.3	97.4	105.7
	8	10	101.3	95.7	103.5	102.2	97.9	102.4
	9	10	100.1	99.8	96.3	100.9	100.5	96.9
2-GR	8	9	-	-	-	-	-	-
	8	10	-	-	-	-	-	-
	9	10	-	-	94.3	-	-	96.6

BC TABLE 81

MEAN RED FLUORESCENCE OF GRANULOCYTES, COMPARISON OF POOLS, DNA ASSAY, INDIVIDUAL VALUES

[?] Remarks: see BC TABLE 78

(a) 1st pool relative to 2nd pool

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) B13 WS

GROUP	POOL 1ST	POOL 2ND	STATISTICAL PARAMETER RELATIVE MEAN RED FLUORESCENCE SIGNAL (a) N	AREA M (0/0)	AREA SE	AREA RSD (0/0)	HEIGHT M (0/0)	HEIGHT SE	HEIGHT RSD (0/0)
0-GR	8	9	3	100.7	1.4	2.5	100.2	1.2	2.1
	8	10	3	98.5	2.3	4.1	99.3	1.2	2.0
	9	10	3	97.8	1.1	1.9	99.1	0.1	0.2
1-GR	8	9	3	101.5	3.3	5.7	101.5	2.4	4.1
	8	10	3	100.2	2.3	4.0	100.8	1.5	2.5
	9	10	3	98.8	1.2	2.1	99.4	1.3	2.2
2-GR	8	9	–	–	–	–	–	–	–
	8	10	–	–	–	–	–	–	–
	9	10	1	94.3	–	–	96.6	–	–
0-GR to 2-GR	8	9	6	101.1	1.6	3.9	100.8	1.2	3.0
	8	10	6	99.3	1.5	3.7	100.0	0.9	2.2
	9	10	7	97.7	0.9	2.3	98.9	0.6	1.7

BC TABLE 82

MEAN RED FLUORESCENCE OF GRANULOCYTES, COMPARISON OF POOLS, DNA ASSAY, STATISTICAL PARAMETERS

[?] Remarks: see BC TABLE 81

(a) 1st pool relative to 2nd pool

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Source: https://www.industrydocuments.ucsf.edu/docs/ndf0000

SUBREPORT P 0500/3057 GD103 (R) B14 WS

GROUP	POOL	STATISTICAL PARAMETER CV N	AREA M (0/0)	AREA SE	AREA RSD (0/0)	HEIGHT M (0/0)	HEIGHT SE	HEIGHT RSD (0/0)
0-GR	8	3	3.57	0.22	10.6	6.97	0.97	24.2
	9	3	3.77	0.20	9.3	6.60	0.57	14.9
	10	3	3.87	0.13	6.0	7.03	0.27	6.7
	8 to 10	9	3.73	0.10	8.4	6.87	0.34	14.9
1-GR	8	3	3.70	0.15	7.2	7.70	0.31	6.9
	9	3	3.97	0.29	12.7	7.93	0.47	10.3
	10	3	3.90	0.20	8.9	7.80	0.31	6.8
	8 to 10	9	3.86	0.12	9.2	7.81	0.19	7.2
2-GR	8	-	-	-	-	-	-	-
	9	1	4.0	-	-	7.9	-	-
	10	1	4.0	-	-	8.3	-	-
	9 and 10	2	4.0	-	-	8.1	-	-

BC TABLE 83

CV OF RED FLUORESCENCE SIGNALS OF GRANULOCYTES, DNA ASSAY, STATISTICAL PARAMETERS

[7] Remarks: see BC TABLE 78

2029027402

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) B15 WS

GROUP	POOL	EXP. NO.	RED FLUORESCENCE SIGNALS						
			N	AREA			HEIGHT		
				MEAN (CH. NO.)	CV (0/0)	SHIFT (a) (0/0)	MEAN (CH. NO.)	CV (0/0)	SHIFT (a) (0/0)
0-GR	8	1	34993	222.0	6.9	12.9	276.0	8.0	-6.79
		2	38226	238.2	8.8	15.2	357.8	10.7	-2.93
		3	10390	278.9	7.8	17.0	436.0	10.0	-6.56
	9	1	29029	218.3	6.6	14.4	278.6	8.1	-3.47
		2	36776	236.6	8.0	15.6	360.7	10.9	-3.17
		3	35461	284.5	7.8	17.0	442.3	10.1	-6.17
	10	1	35119	222.3	6.5	14.7	283.8	8.1	-2.64
		2	32709	234.5	7.9	13.7	360.3	10.8	-3.95
		3	39269	290.3	7.5	14.3	441.7	10.9	-7.32

BC TABLE 84

RED FLUORESCENCE SIGNALS OF MACROPHAGES IN G0/1 (b), DNA ASSAY

(a) of the mean, relative to the mean position of the corresponding signals from granulocytes (see BC TABLE 78)
(b) region 3 (see BC FIGURE ..)

2029027403

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) B16 WS

GROUP	POOL	EXP. NO.	RED FLUORESCENCE SIGNALS						
			N	AREA			HEIGHT		
				MEAN (CH. NO.)	CV (0/0)	SHIFT (a) (0/0)	MEAN (CH. NO.)	CV (0/0)	SHIFT (a) (0/0)
1-GR	8	1	10004	245.4	11.3	23.9	292.8	9.3	-1.91
		2	5363	236.7	12.9	19.8	357.4	12.1	-1.73
		3	13518	315.8	12.3	25.2	456.5	10.4	-4.54
	9	1	16691	237.2	11.1	21.2	285.7	9.6	-3.02
		2	13807	256.9	12.5	24.7	373.7	11.6	0.03
		3	12239	291.8	11.3	24.2	429.5	10.7	-5.06
	10	1	18670	237.7	10.7	21.6	290.6	9.2	-0.68
		2	18500	260.6	12.3	26.3	375.1	11.8	0.94
		3	10831	301.6	11.8	23.7	446.5	10.7	-4.89
2-GR	8	3	-	-	-	-	-	-	-
	9	3	18468	281.6	8.6	17.8	422.0	10.8	-7.92
	10	3	24329	299.5	9.2	18.2	443.6	10.4	-6.49

BC TABLE 84 (continued)

RED FLUORESCENCE SIGNALS OF MACROPHAGES IN G0/1 (b), DNA ASSAY

(a) of the mean, relative to the mean position of the corresponding signals from granulocytes (see BC TABLE ..)
(b) region 3 (see BC FIGURE ..)

2029027404

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER SHIFT			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	15.05	1.18	13.6
	9	3	15.65	0.77	8.6
	10	3	14.22	0.30	3.7
	8 to 10	9	14.98	0.47	9.3
1-GR	8	3	22.95	1.62	12.2
	9	3	23.38	1.09	8.1
	10	3	23.85	1.35	9.8
	8 to 10	9	23.39	0.70	9.0
2-GR	8	-	-	-	-
	9	1	17.8	-	-
	10	1	18.2	-	-
	9 and 10	2	18.01	-	-

BC TABLE 85

SHIFT OF MEAN RED FLUORESCENCE SIGNAL AREA OF MACROPHAGES IN G0/1, DNA ASSAY, STATISTICAL PARAMETERS

Remarks: shift relative to mean red fluorescence signal area of granulocytes (see BC TABLE 78.)
See BC TABLE 84

2029027405

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER SHIFT (0/0)			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	-5.43	1.25	39.9
	9	3	-4.27	0.96	38.8
	10	3	-4.64	1.39	52.1
	8 to 10	9	-4.78	0.63	39.6
1-GR	8	3	-2.73	0.91	57.6
	9	3	-2.69	1.48	95.4
	10	3	-1.38	1.58	198.4
	8 to 10	9	-2.26	0.71	94.4
2-GR	8	-	-	-	-
	9	1	-7.92	-	-
	10	1	-6.49	-	-
	9 and 10	2	-7.21	-	-

BC TABLE 86

SHIFT OF MEAN RED FLUORESCENCE SIGNAL HEIGHT OF MACROPHAGES IN G0/1, DNA ASSAY, STATISTICAL PARAMETERS

Remarks: shift relative to mean red fluorescence signal height of granulocytes (see BC TABLE 78)

see BC TABLE 84

20290274O6

Source: https://www.industrydocuments.ucsf.edu/docs/ynd0000

GROUP	POOL	EXP. NO.	NUMBER OF SIGNALS CORRESPONDING DNA CONTENT					
			G0/1	S	G2	2 x G0/1	.GT.2 x G0/1	TOTAL
0-GR	8	1	34993	604	1069	453	49	37168
		2	38226	363	719	567	69	39944
		3	10390	192	178	295	38	11093 (a)
	9	1	29029	437	744	443	40	30693
		2	36776	500	693	700	94	38763
		3	35461	707	586	1837	280	38871
	10	1	35119	559	1024	735	55	37492
		2	32709	475	634	460	72	34350 (b)
		3	39269	607	743	808	66	41493

BC TABLE 87

NUMBER OF RED FLUORESCENCE SIGNALS FROM MACROPHAGES CORRESPONDING TO VARIOUS DNA CONTENTS, ALL GROUPS

Remarks: total number of signals analyzed: 50000, unless otherwise noted
signals in region 1 to 5 (see BC FIGURE ..)

(a) 13602 signals analyzed
(b) 43900 signals analyzed

2029027407

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF SIGNALS CORRESPONDING DNA CONTENT					
			G0/1	S	G2	2 x G0/1	.GT.2 x G0/1	TOTAL
1-GR	8	1	10004	660	602	560	141	11967 (a)
		2	5363	449	290	438	98	6638
		3	13518	882	591	700	132	15823
	9	1	16691	618	1069	777	88	19243
		2	13807	835	475	765	170	16052
		3	12239	933	493	898	221	14784
	10	1	18670	539	1087	726	84	21106
		2	18500	854	740	1314	193	21601
		3	10831	410	407	550	73	12271
2-GR	8	3	-	-	-	-	-	-
	9	3	18468	466	607	402	86	20029 (b)
	10	3	24329	420	540	517	69	25875

BC TABLE 87 (continued)

NUMBER OF RED FLUORESCENCE SIGNALS FROM MACROPHAGES CORRESPONDING TO VARIOUS DNA CONTENTS, ALL GROUPS

Remarks: total number of signals analyzed: 50000, unless otherwise noted
signals in region 1 to 5 (see BC FIGURE ..)

(a) 41496 signals analyzed
(b) 30105 signals analyzed

2029027408

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) B21 WS

GROUP	POOL	EXP. NO.	RELATIVE NUMBER OF SIGNALS (0/0) CORRESPONDING DNA CONTENT				
			G0/1	S	G2	2 x G0/1	.GT.2 x G0/1
0-GR	8	1	94.1	1.6	2.9	1.2	0.1
		2	95.7	0.9	1.8	1.4	0.2
		3	93.7	1.7	1.6	2.7	0.3
	9	1	94.6	1.4	2.4	1.4	0.1
		2	94.9	1.3	1.8	1.8	0.2
		3	91.2	1.8	1.5	4.7	0.7
	10	1	93.7	1.5	2.7	2.0	0.1
		2	95.2	1.4	1.8	1.3	0.2
		3	94.6	1.5	1.8	1.9	0.2

BC TABLE 88

RELATIVE NUMBER OF RED FLUORESCENCE SIGNALS FROM MACROPHAGES CORRESPONDING TO VARIOUS DNA CONTENTS, INDIVIDUAL VALUES

Remarks: signals in region 1 to 5 (see BC FIGURE ..)
see BC TABLE 87

2029027409

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	RELATIVE NUMBER OF SIGNALS (0/0) CORRESPONDING DNA CONTENT				
			G0/1	S	G2	2 x G0/1	.GT.2 x G0/1
1-GR	8	1	83.6	5.5	5.0	4.7	1.2
		2	80.8	6.8	4.4	6.6	1.5
		3	85.4	5.6	3.7	4.4	0.8
	9	1	86.7	3.2	5.6	4.0	0.5
		2	86.0	5.2	3.0	4.8	1.1
		3	82.8	6.3	3.3	6.1	1.5
	10	1	88.5	2.6	5.2	3.4	0.4
		2	85.6	4.0	3.4	6.1	0.9
		3	88.3	3.3	3.3	4.5	0.6
2-GR	8	3	-	-	-	-	-
	9	3	92.2	2.3	3.0	2.0	0.4
	10	3	94.0	1.6	2.1	2.0	0.3

BC TABLE 88 (continued)

RELATIVE NUMBER OF RED FLUORESCENCE SIGNALS FROM MACROPHAGES CORRESPONDING TO VARIOUS DNA CONTENTS, INDIVIDUAL VALUES

Remarks: signals in region 1 to 5 (see BC FIGURE ..)
see BC TABLE 8?

2029027410

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF SIGNALS CORRESPONDING DNA CONTENT					
			G0	S	G2	2 x G0/1	.GT.2 x G0/1	TOTAL
0-GR	8	1	517	1	0	0	0	518
		2	808	4	0	0	0	812
		3	173	3	0	0	0	176 (a)
	9	1	326	1	0	0	0	327
		2	942	7	0	0	0	949
		3	718	0	0	0	0	718
	10	1	163	1	0	0	0	164
		2	589	4	0	0	0	593 (b)
		3	672	6	0	1	0	679

BC TABLE 89

NUMBER OF RED FLUORESCENCE SIGNALS FROM GRANULOCYTES CORRESPONDING TO VARIOUS DNA CONTENTS, INDIVIDUAL VALUES

Remarks: total number of signals analyzed: 50000, unless otherwise noted
signals in region 1 to 5 (see BC FIGURE ..)

(a) 13602 signals analyzed
(b) 43900 signals analyzed

20290274 11

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF SIGNALS CORRESPONDING DNA CONTENT G0	S	G2	2 x G0/1	.GT.2 x G0/1	TOTAL
1-GR	8	1	13927	6	6	2	0	13941 (a)
		2	15039	25	2	4	1	15071
		3	13814	21	4	6	2	13847
	9	1	10627	8	0	0	0	10635
		2	18624	39	2	1	0	18666
		3	19390	24	4	8	0	19426
	10	1	4380	0	0	1	0	4381
		2	13816	41	12	7	0	13876
		3	8216	9	0	2	2	8229
2-GR	8	3	-	-	-	-	-	-
	9	3	4397	23	0	1	0	4421 (b)
	10	3	3101	25	0	2	0	3128

BC TABLE 89 (continued)

NUMBER OF RED FLUORESCENCE SIGNALS FROM GRANULOCYTES CORRESPONDING TO VARIOUS DNA CONTENTS, INDIVIDUAL VALUES

Remarks: total number of signals analyzed: 50000, unless otherwise noted
signals in region 1 to 5 (see BC FIGURE ..)

(a) 41496 signals analyzed
(b) 30105 signals analyzed

20290274 12

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD103 (R) B25 WS

GROUP	POOL	EXP. NO.	RELATIVE NUMBER OF SIGNALS (0/0) CORRESPONDING DNA CONTENT				
			G0	S	G2	2 x G0	.GT.2 x G0
0-GR	8	1	99.8	0.2	0.0	0.0	0.0
		2	99.5	0.5	0.0	0.0	0.0
		3	98.3	1.7	0.0	0.0	0.0
	9	1	99.7	0.3	0.0	0.0	0.0
		2	99.3	0.7	0.0	0.0	0.0
		3	100.0	0.0	0.0	0.0	0.0
	10	1	99.4	0.6	0.0	0.0	0.0
		2	99.3	0.7	0.0	0.0	0.0
		3	99.0	0.9	0.0	0.1	0.0

BC TABLE 90

RELATIVE NUMBER OF RED FLUORESCENCE SIGNALS FROM GRANULOCYTES CORRESPONDING TO VARIOUS DNA CONTENTS, INDIVIDUAL VALUES

Remarks: see BC TABLE 89

20290274 13

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	RELATIVE NUMBER OF SIGNALS (0/0) CORRESPONDING DNA CONTENT				
			G0	S	G2	2 x G0	.GT.2 x G0
1-GR	8	1	99.9	0.0	0.0	0.0	0.0
		2	99.8	0.2	0.0	0.0	0.0
		3	99.8	0.2	0.0	0.0	0.0
	9	1	99.9	0.1	0.0	0.0	0.0
		2	99.8	0.2	0.0	0.0	0.0
		3	99.8	0.1	0.0	0.0	0.0
	10	1	100.0	0.0	0.0	0.0	0.0
		2	99.6	0.3	0.1	0.1	0.0
		3	99.98	0.1	0.0	0.0	0.0
2-GR	8	3	-	-	-	-	-
	9	3	99.5	0.5	0.0	0.0	0.0
	10	3	99.1	0.8	0.0	0.1	0.0

BC TABLE 90 (continued)

RELATIVE NUMBER OF RED FLUORESCENCE SIGNALS FROM GRANULOCYTES CORRESPONDING TO VARIOUS DNA CONTENTS, INDIVIDUAL VALUES

20290274 14

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER RELATIVE NUMBER OF GRANULOCYTES N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	99.21	0.46	0.8
	9	3	99.65	0.21	0.4
	10	3	99.23	0.13	0.2
	8 to 10	9	99.36	0.17	0.5
1-GR	8	3	99.82	0.04	0.1
	9	3	99.83	0.04	0.1
	10	3	99.80	0.12	0.2
	8 to 10	9	99.82	0.04	0.1
2-GR	8	-	-	-	-
	9	1	99.5	-	-
	10	1	99.1	-	-
	9 and 10	2	99.30	-	-

BC TABLE 91

RELATIVE NUMBER OF GRANULOCYTES IN REGION CORRESPONDING TO G0 DNA CONTENT, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 90

202902741S

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	0.79	0.46	100.8
	9	3	0.35	0.21	106.2
	10	3	0.72	0.08	19.7
	8 to 10	9	0.62	0.16	79.1
1-GR	8	3	0.12	0.04	58.3
	9	3	0.14	0.04	48.7
	10	3	0.14	0.09	111.0
	8 to 10	9	0.13	0.03	68.9
2-GR	8	-	-	-	-
	9	1	0.5	-	-
	10	1	0.8	-	-
	9 and 10	2	0.66	-	-

BC TABLE 92

RELATIVE NUMBER OF GRANULOCYTES IN REGION CORRESPONDING TO S PHASE DNA CONTENT, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 90

202927416

Source: https://www.industrydocuments.ucsf.edu/docs/lyndl0000

GROUP	POOL	STATISTICAL PARAMETER			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	0.00	0.00	-
	9	3	0.00	0.00	-
	10	3	0.00	0.00	-
	8 to 10	9	0.00	0.00	-
1-GR	8	3	0.03	0.01	57.3
	9	3	0.01	0.01	100.0
	10	3	0.03	0.05	173.2
	8 to 10	9	0.02	0.01	130.8
2-GR	8	-	-	-	-
	9	1	0.0	-	-
	10	1	0.0	-	-
	9 and 10	2	0.00	-	-

BC TABLE 93.1

RELATIVE NUMBER OF GRANULOCYTES IN REGION CORRESPONDING TO G2 DNA CONTENT, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 90

2029027417

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	0.00	0.00	-
	9	3	0.00	0.00	-
	10	3	0.05	0.05	173.2
	8 to 10	9	0.02	0.02	300.0
1-GR	8	3	0.03	0.01	57.3
	9	3	0.02	0.01	124.9
	10	3	0.03	0.01	57.7
	8 to 10	9	0.02	0.01	68.2
2-GR	8	-	-	-	-
	9	1	0.0	-	-
	10	1	0.1	-	-
	9 and 10	2	0.04	-	-

BC TABLE 93.2

RELATIVE NUMBER OF GRANULOCYTES IN REGION CORRESPONDING TO 2 x ~~G2/G~~ G0 DNA CONTENT, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 90

2029027418

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	0.00	0.00	-
	9	3	0.00	0.00	-
	10	3	0.00	0.00	-
	8 to 10	9	0.00	0.00	-
1-GR	8	3	0.01	0.00	86.6
	9	3	0.00	0.00	-
	10	3	0.01	0.01	173.2
	8 to 10	9	0.00	0.00	163.5
2-GR	8	-	-	-	-
	9	1	0.0	-	-
	10	1	0.0	-	-
	9 and 10	2	0.00	-	-

BC TABLE 94

RELATIVE NUMBER OF GRANULOCYTES IN REGION CORRESPONDING TO .GT.2 x G0 DNA CONTENT, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 90

2029027419

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER RELATIVE NUMBER OF MACROPHAGES (a) N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	94.50	0.61	1.1
	9	3	93.56	1.17	2.2
	10	3	94.51	0.45	0.8
	8 to 10	9	94.19	0.43	1.4
1-GR	8	3	83.27	1.35	2.8
	9	3	85.18	1.21	2.5
	10	3	87.46	0.91	1.8
	8 to 10	9	85.30	0.84	3.0
2-GR	8	-	-	-	-
	9	1	92.21	-	-
	10	1	94.03	-	-
	8 to 10	2	93.12	-	-

BC TABLE 95

RELATIVE NUMBER OF MACROPHAGES IN G0, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 88 and BC FIGURE

[H0335-4]

(a) relative to all macrophages in regions 1 to 5 (see BC FIGURE ..)

2029027420

Source: https://www.industrydocuments.ucsf.edu/docs/ynd10000

GROUP	POOL	STATISTICAL PARAMETER RELATIVE NUMBER OF MACROPHAGES (a)			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	2.09	0.40	32.9
	9	3	1.91	0.27	24.4
	10	3	2.12	0.30	24.8
	8 to 10	9	2.04	0.17	24.6
1-GR	8	3	4.38	0.37	14.7
	9	3	3.95	0.81	35.6
	10	3	3.97	0.59	25.9
	8 to 10	9	4.10	0.32	23.2
2-GR	8	-	-	-	-
	9	1	3.03	-	-
	10	1	2.09	-	-
	8 to 10	2	2.56	-	-

BC TABLE 97

RELATIVE NUMBER OF MACROPHAGES IN G2, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 88 and BC FIGURE
[H0335-4]

(a) relative to all macrophages in regions 1 to 5 (see BC FIGURE ..)

2029027421

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER			
		RELATIVE NUMBER OF MACROPHAGES (a)			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	1.42	0.26	31.4
	9	3	1.51	0.16	18.3
	10	3	1.44	0.03	3.9
	8 to 10	9	1.46	0.09	18.3
1-GR	8	3	5.95	0.41	11.8
	9	3	4.91	0.91	32.0
	10	3	3.28	0.41	21.4
	8 to 10	9	4.71	0.50	31.6
2-GR	8	-	-	-	-
	9	1	2.33	-	-
	10	1	1.62	-	-
	8 to 10	2	1.98	-	-

BC TABLE 96

RELATIVE NUMBER OF MACROPHAGES IN S, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 88 and BC FIGURE
[1103354]

(a) relative to all macrophages in regions 1 to 5 (see BC FIGURE ..)

20290274422

Source: https://www.industrydocuments.ucsf.edu/docs/ynd10000

GROUP	POOL	STATISTICAL PARAMETER RELATIVE NUMBER OF MACROPHAGES (a) N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	1.77	0.45	44.2
	9	3	2.66	1.04	67.8
	10	3	1.75	0.21	20.3
	8 to 10	9	2.06	0.36	53.2
1-GR	8	3	5.23	0.69	22.8
	9	3	4.96	0.59	20.7
	10	3	4.67	0.77	28.5
	8 to 10	9	4.95	0.35	21.5
2-GR	8	-	-	-	-
	9	1	2.01	-	-
	10	1	2.00	-	-
	8 to 10	2	2.01	-	-

BC TABLE 98

RELATIVE NUMBER OF MACROPHAGES IN REGION CORRESPONDING TO 2 x G0 DNA CONTENT, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 88 and BC FIGURE
[H0335-4]

(a) relative to all macrophages in regions 1 to 5 (see BC FIGURE ..)

2029027423

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER			
		RELATIVE NUMBER OF MACROPHAGES (a)			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	3	0.21	0.06	52.3
	9	3	0.36	0.18	86.4
	10	3	0.17	0.02	18.5
	8 to 10	9	0.25	0.06	75.4
1-GR	8	3	1.16	0.19	28.0
	9	3	1.00	0.30	51.6
	10	3	0.63	0.14	39.4
	8 to 10	9	0.93	0.14	43.7
2-GR	8	-	-	-	-
	9	1	0.43	-	-
	10	1	0.27	-	-
	8 to 10	2	0.35	-	-

BC TABLE 99

RELATIVE NUMBER OF MACROPHAGES IN REGION CORRESPONDING TO .GT.2 x G0 DNA CONTENT, STATISTICAL PARAMETERS

Remarks: signals in region (see BC FIGURE ..)
see BC TABLE 88 and BC FIGURE
[H03354]

(a) relative to all macrophages in regions 1 to 5 (see BC FIGURE ..)

2029027424

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

MP.

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF MACROPHAGES POOL		
		8	9	10
0-GR	1	1.44	1.16	1.20
	2	0.98	-	1.51
	3	2.23	0.69	1.19
1-GR	1	5.02	5.51	6.34
	2	4.18	-	5.20
	3	5.94	7.57	2.28
2-GR	3	2.94	3.13	2.37

BC TABLE 100

RELATIVE NUMBER OF MULTINUCLEATED MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, WEIGHTED MEAN, POOL 8 TO 10

Remarks: microscope counting

pool 8 to 10 weighted according to composition 3:7 of macrophages (see BC PAGE 4-1), equivalent to pools 8 to 10 used for phagocytosis and DNA assay [vergl. Tab. 109]

2029027425

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	FORWARD SCATTER SIGNALS N	MEAN (CH. NO.)	CV (0/0)	MEAN REL. TO 0-GR (0/0) (a)
0-GR	1	32828	463.4	19.2	100
	2	17953	459.4	18.3	100
	3	19431	449.9	20.9	100
	4	22050	440.9	15.9	100
	5	21909	439.7	18.7	100
	6	15534	450.7	15.6	100
	7	15639	449.1	15.2	100
1-GR	1	11278	441.6	20.3	95.3
	2	15844	463.1	18.9	100.8
	3	7343	433.8	20.4	96.4
	4	14395	425.2	18.0	96.4
	5	6826	444.7	18.7	101.1
	6	23795	452.5	16.6	100.4
	7	20408	465.8	16.9	103.7
2-GR	1	13292	468.5	24.5	101.1
	2	18338	490.8	23.2	106.8
	3	16382	475.7	24.5	105.7
	4	14832	464.7	23.2	105.4
	5	18173	511.3	24.1	116.3
	6	12743	465.0	24.3	103.2
	7	12765	463.9	23.2	103.2

BC TABLE 101

FORWARD SCATTER SIGNALS FROM MACROPHAGES, VIABILITY AND ESTERASE ASSAY, EXPERIMENT 3, INDIVIDUAL VALUES

Remarks: viable macrophages only (see [redacted]
total number of signals analyzed: [redacted]
.LE.0.2 0/0 of macrophage signals out of range

(a) corresponding pool

2029027426

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	MEAN FORWARD SCATTER					
	POOL					
	1 AND 2		3 AND 4		5, 6 AND 7	
	ABS. (CH. NO.)	REL. (a) (0/0)	ABS. (CH. NO.)	REL. (a) (0/0)	ABS. (CH. NO.)	REL. (a) (0/0)
0-GR	461.7	100	442.3	100	448.1	100
1-GR	455.6	98.7	427.5	96.7	460.3	102.7
2-GR	482.6	104.5	468.0	105.8	469.4	104.8

BC TABLE 102

FORWARD SCATTER OF MACROPHAGES, VIABILITY AND ESTERASE ASSAY, EXPERIMENT 3, WEIGHTED MEAN OF POOLS

Remarks: mean weighted according to number of macrophages/pool/~~(see BC TABLE ..)~~, 5 rats, resuspension medium, hemocytometer
equivalent to combined respective pools
no data from experiment 2 due to laser failure
See BC TABLE 101

[Werte in keine Tabelle, nur auf Zählprotokolle]

(a) to 0-GR .

20290274 27

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER					
	MEAN FORWARD SCATTER					
	ABSOLUTE			RELATIVE TO 0-GR		
	M (CH. NO.)	SE	RSD (0/0)	M (0/0)	SE	RSD (0/0)
0-GR	450.7	5.8	2.22	100	-	-
1-GR	447.8	10.2	3.96	99.4	1.8	3.11
2-GR	473.4	4.7	1.70	105.0	0.4	0.66

BC TABLE 103

FORWARD SCATTER OF MACROPHAGES, VIABILITY AND ESTERASE ASSAY, EXPERIMENT 3, STATISTICAL PARAMETERS

Remarks: N = 3, statistical parameters for weighted mean of pools (see BC TABLE 102)

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	FORWARD SCATTER SIGNALS N	MEAN (a)	CV (0/0)	MEAN REL. TO 0-GR (0/0) (b)
0-GR	8	1 (c)	9600	.GT.19.3	- (d)	-
		2	5540	14.8	21.0	-
		3 (e)	-	-	-	-
	9	1	6411	14.0	- (d)	100
		2	4164	12.7	17.4	100
		3 (e)	-	-	-	-
	10	1	9628	13.0	13.6	100
		2	6385	12.2	16.1	100
		3 (e)	-	-	-	-
1-GR	8	1	-	-	-	-
		2	-	-	-	-
		3	-	-	-	-
	9	1	6723	15.0	- (d)	107.2
		2	2974	13.2	18.8	103.9
		3 (e)	-	-	-	-
	10	1	6494	14.5	- (d)	111.7
		2	4418	13.2	17.0	107.7
		3 (e)	-	-	-	-
2-GR	8	3 (e)	-	-	-	-
	9	3 (e)	-	-	-	-
	10	3 (e)	-	-	-	-

BC TABLE 104

FORWARD SCATTER SIGNALS FROM MACROPHAGES, PHAGOCYTOSIS ASSAY

Remarks: sample taken at time = 0 of incubation
macrophages associated with latex beads (mean: 12.9 0/0, RSD: 47.5 0/0) excluded from evaluation
total number of signals analyzed: 80000

(a) normalized to standard latex beads
(b) corresponding pool, same experiment
(c) atypical FWD distribution, 50.9 0/0 of signals out of range
(d) not meaningful, .GT.1 0/0 of signals out of range
(e) data not evaluated, poor instrument alignment for this parameter

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	AXIAL LIGHT LOSS SIGNALS N	MEAN (CH. NO.)	CV (0/0)	MEAN REL. TO 0-GR (0/0) (a)
0-GR	1	32828	325.6	11.9	100
	2	17953	333.3	10.8	100
	3	19431	328.6	11.9	100
	4	22050	327.9	11.0	100
	5	21909	329.4	10.8	100
	6	15534	323.9	9.5	100
	7	15639	324.3	9.5	100
1-GR	1	11278	419.6	22.0	128.9
	2	15844	406.7	20.7	122.0
	3	7343	413.5	22.4	125.8
	4	14395	390.1	19.7	119.0
	5	6826	407.6	22.0	123.7
	6	23795	389.7	20.4	120.3
	7	20408	388.5	19.8	119.8
2-GR	1	13292	356.2	15.4	109.4
	2	18338	354.9	15.0	106.5
	3	16382	341.9	15.3	104.1
	4	14832	338.8	13.6	103.3
	5	18173	369.9	16.9	112.3
	6	12743	345.1	15.5	106.6
	7	12765	342.7	14.9	105.7

BC TABLE 105

AXIAL LIGHT LOSS SIGNALS FROM MACROPHAGES, VIABILITY AND ESTERASE ASSAY, EXPERIMENT 3, INDIVIDUAL VALUES

Remarks: viable macrophages only (see
total number of signals analyzed: 19400 to 67400
.LE.2 macrophage signals out of range

(a) corresponding pool

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	WEIGHTED MEAN AXIAL LIGHT LOSS					
		POOL					
		1 AND 2		3 AND 4		5, 6 AND 7	
		ABS. (CH. NO.)	REL. (a) (0/0)	ABS. (CH. NO.)	REL. (a) (0/0)	ABS. (CH. NO.)	REL. (a) (0/0)
0-GR	1 (b)	-	-	-	-	-	-
	3	328.8	100	328.0	100	325.0	100
1-GR	1 (b)	-	-	-	-	-	-
	3	411.2	125.1	396.4	120.9	391.0	120.3
2-GR	3	355.4	108.1	339.7	103.6	346.4	106.6

BC TABLE 106

AXIAL LIGHT LOSS OF VIABLE MACROPHAGES, WEIGHTED MEAN OF POOL 1 AND 2, 3 AND 4 AND 5, 6 AND 7

Remarks: mean weighted according to number of macrophages/pool/~~(see BC TABLE ..);~~ Srab, resuspension medium, hemocytometer
equivalent to combined respective pools
no data from experiment 2 due to laser failure
see BC TABLE 105 [s. Tab 102]

(a) to 0-GR
(b) not calculated, excessive number of signals out of range (see BC TABLE

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER					
	MEAN AXIAL LIGHT LOSS					
	ABSOLUTE			RELATIVE TO 0-GR		
	M (CH. NO.)	SE	RSD (0/0)	M (0/0)	SE	RSD (0/0)
0-GR	327.3	1.2	0.61	100	-	-
1-GR	399.5	6.0	2.62	122.1	1.5	2.13
2-GR	347.2	4.5	2.26	106.1	1.3	2.17

BC TABLE 107

AXIAL LIGHT LOSS OF MACROPHAGES, VIABILITY AND ESTERASE ASSAY, EXPERIMENT 3, STATISTICAL PARAMETERS

Remarks: N = 3, statistical parameters for weighted mean of pools (see BC TABLE 106)

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	AXIAL LIGHT LOSS SIGNALS N	MEAN (a)	CV (0/0)	MEAN REL. TO 0-GR (0/0) (b)
0-GR	8	1	9600	15.0	- (c)	-
		2	5540	14.2	12.9	-
		3	-	-	-	-
	9	1	6411	14.9	13.1	100
		2	4164	13.3	11.2	100
		3	-	-	-	-
	10	1	9628	14.8	12.8	100
		2	6385	13.1	11.5	100
		3	8534	10.2	10.6	100
1-GR	8	1	-	-	-	-
		2	-	-	-	-
		3	-	-	-	-
	9	1	6723	.GT. 18.2 (d)	- (c)	.GT. 122.8
		2	2974	16.3	19.5	123.2
		3	4995	12.9	22.7	-
	10	1	6494	.GT. 18.0 (e)	- (c)	.GT. 122.2
		2	4418	16.4	- (c)	125.2
		3	7817	12.7	21.8	124.0
2-GR	8	3	-	-	-	-
	9	3	6169	10.8	15.4	-
	10	3	7886	11.1	17.0	108.8

BC TABLE 108

AXIAL LIGHT LOSS SIGNALS FROM MACROPHAGES, PHAGOCYTOSIS ASSAY

Remarks: sample taken at time = 0 of incubation
macrophages associated with latex beads (mean: 12.9 0/0, RSD: 47.5 0/0) excluded from evaluation
total number of signals analyzed: 80000

(a) normalized to standard latex beads
(b) corresponding pool, same experiment
(c) not meaningful, .GT.1 0/0 of signals out of range
(d) 10 0/0 of signals out of range but included in calculation of mean
(e) 7.5 0/0 of signals out of range but included in calculation of mean

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	WEIGHTED MEAN AXIAL LIGHT LOSS					
		POOL					
		8		9		10	
		ABS. (CH. NO.)	REL. (a) (0/0)	ABS. (CH. NO.)	REL. (a) (0/0)	ABS. (CH. NO.)	REL. (a) (0/0)
0-GR	1 (b)	-	-	-	-	-	-
	3	331.0	100	328.1	100	325.4	100
1-GR	1 (b)	-	-	-	-	-	-
	3	410.6	124.0	397.1	121.0	393.2	120.8
2-GR	3	355.3	107.3	339.7	103.5	349.5	107.4

BC TABLE 109

AXIAL LIGHT LOSS OF VIABLE MACROPHAGES, WEIGHTED MEAN, POOL 8 TO 10

Remarks: mean weighted according to composition 3 : 7 of macrophages (see BC PAGE 4-1.), equivalent to pools used for phagocytosis assay
see BC TABLE 105

(a) to 0-GR
(b) not calculated, excessive number of signals out of range (see BC TABLE

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF RIGHT ANGLE SCATTER SIGNALS				
			TOTAL	.GE.14 x STANDARD (a)		OUT OF RANGE	
				ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)
0-GR	8	1	9216	5480	59.5	1610	17.5
		2	5464	1676	30.7	231	4.2
		3	-	-	-	-	-
	9	1	6233	1314	21.1	362	5.8
		2	4164	580	13.9	85	2.0
		3	-	-	-	-	-
	10	1	9628	2225	23.1	594	6.2
		2	6385	1064	16.7	143	2.2
		3	8534	1551	18.2	169	2.0
1-GR	8	1	-	-	-	-	-
		2	-	-	-	-	-
		3	-	-	-	-	-
	9	1	6723	4892	72.8	3337	49.6
		2	2974	1993	67.0	648	21.8
		3	4995	3465	69.4	1207	24.2
	10	1	6494	4634	71.4	2894	44.6
		2	4418	2970	67.2	930	21.1
		3	7817	5051	64.6	1529	19.6
2-GR	8	3	-	-	-	-	-
	9	3	6169	2664	43.2	340	5.5
	10	3	7886	3905	49.5	614	7.8

BC TABLE 110

NUMBER OF RIGHT ANGLE SCATTER SIGNALS FROM MACROPHAGES, PHAGOCYTOSIS ASSAY

Remarks: sample taken at time = 0 of incubation, macrophages associated with latex beads (mean: 12.9 0/0, RSD: 47.5 0/0) excluded from evaluation

(a) standard latex beads

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	STATISTICAL PARAMETER			
		RELATIVE NUMBER OF RAS SIGNALS			
		N	M (0/0)	SE	RSD (0/0)
0-GR	8	1 (a)	30.7	-	-
	9	2	17.5	-	-
	10	3	19.3	1.9	17.31
1-GR	8	-	-	-	-
	9	3	69.7	1.7	4.18
	10	3	67.7	2.0	5.07
2-GR	8	-	-	-	-
	9	1	43.2	-	-
	10	1	49.5	-	-

BC TABLE 111

RELATIVE NUMBER OF RIGHT ANGLE SCATTER SIGNALS FROM MACROPHAGES .GT.14 x STANDARD, PHAGOCYTOSIS ASSAY, STATISTICAL PARAMETERS

Remarks: see BC TABLE 110 and BC FIGURE .. [Figure 3]

(a) value from experiment 1 not used for calculation of mean (see

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE NUMBER OF CELLS (0/0)					
		MACROPHAGES			GRANULOCYTES		
		POOL			POOL		
		8	9	10	8	9	10
0-GR	1	99.9	99.3	100.0	0.1	0.7	0.0
	2	100.0	99.5	96.7	0.0	0.5	3.3
	3	99.5	99.7	99.2	0.5	0.3	0.8
1-GR	1	62.3	56.1	85.9	37.7	43.9	14.1
	2	46.0	-	61.3	54.0	-	38.7
	3	50.7	54.4	62.4	49.3	45.6	37.6
2-GR	3	92.3	84.2	92.3	7.7	15.8	7.7

BC TABLE 112

RELATIVE NUMBER OF MACROPHAGES AND GRANULOCYTES IN RESUSPENSION MEDIUM, WEIGHTED MEAN, POOL 8 TO 10

Remarks: microscope counting, cytocentrifuge preparation
pool 8 to 10 calculated according to the composition 3 : 7 of macrophages
(see BC PAGE 4-1)
sum of macrophages and granulocytes taken to be 100 0/0

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[Fig. H03358, H03610, H03608, H03611, H03609]

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES.		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
0-GR	1	1 (b)	-	-	-	-	-
		3	36268	96.7	1224	3.3	37492
			35404	95.6	1634	4.4	37038
			36323	96.9	1146	3.1	37469
	2	1 (b)	-	-	-	-	-
		3	19874	98.9	218	1.1	20092
			19571	99.1	174	0.9	19745
			19787	98.5	294	1.5	20081
	3	1	6525	87.7	913	12.3	7438
			6588	88.2	882	11.8	7470
		3	20103	90.5	2121	9.5	22224
			20283	91.7	1830	8.3	22113
			20066	89.6	2323	10.4	22389
	4	1	13903	96.5	505	3.5	14408
			13832	96.7	466	3.3	14298
		3	23408	99.3	171	0.7	23579
			23152	99.4	134	0.6	23286
			23390	99.2	178	0.8	23568
	5	1	2858	90.2	312	9.8	3170
			2816	90.7	289	9.3	3105
		3	22728	86.8	3446	13.2	26174
			22423	87.0	3363	13.0	25786
			22721	86.9	3422	13.1	26143
	6	1	11000	98.6	157	1.4	11157
			10958	98.5	162	1.5	11120
		3	16876	98.8	210	1.2	17086
			16758	98.4	271	1.6	17029
			16944	99.0	179	1.0	17123

BC TABLE 113

NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, VIABILITY AND ESTERASE ASSAY, INDIVIDUAL VALUES

Remarks: values from 2 to 3 evaluations of data from 1 FCM determination
total counts: ..... to .....

(a) sum of macrophages and granulocytes
(b) atypical FWD versus AXL distribution (see BC FIGURE ..), evaluation not possible

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
0-GR	7	1	4280	97.7	103	2.3	4383
			4290	97.8	96	2.2	4388
		3	16803	99.2	133	0.8	16936
			16767	99.3	121	0.7	16888
			16751	99.0	169	1.0	16920
1-GR	1	1	2044	25.9	5847	74.1	7891
			1883	25.5	5494	74.5	7377
		3	13162	39.7	20018	60.3	33180
			12909	40.8	18731	59.2	31640
			13324	40.4	19641	59.6	32965
	2	1	8574	69.0	3858	31.0	12432
			7749	68.3	3603	31.7	11352
		3	17296	76.3	5375	23.7	22671
			17431	77.7	4996	22.3	22427
			17304	75.7	5560	24.3	22864
	3	1	3244	35.4	5913	64.6	9157
			3132	36.0	5580	64.0	8712
		3	7873	26.5	21887	73.5	29760
			7728	25.7	22290	74.3	30018
			7800	25.8	22484	74.2	30284
	4	1	15095	69.0	6787	31.0	21882
			14626	69.1	6550	30.9	21176
		3	15687	59.6	10646	40.4	26333
			15501	59.1	10710	40.9	26211
			15715	59.4	10735	40.6	26450
	5	1	1156	29.5	2769	70.5	3925
			1186	30.4	2718	69.6	3904

BC TABLE 113 (continued)

NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, VIABILITY AND ESTERASE ASSAY, INDIVIDUAL VALUES

Remarks: values from 2 to 3 evaluations of data from 1 FCM determination
total counts: ..... to .....

(a) sum of macrophages and granulocytes

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
1-GR	5	3	7608	22.9	25676	77.1	33284
			7400	22.5	25485	77.5	32885
			7755	22.8	26213	77.2	33968
	6	1	14769	71.7	5815	28.3	20584
			14947	72.6	5655	27.4	20602
		3	25236	56.9	19080	43.1	44316
			25123	57.4	18674	42.6	43797
			25207	56.5	19380	43.5	44587
	7	1	21024	88.3	2792	11.7	23816
			21010	89.3	2505	10.7	23515
		3	22067	76.7	6708	23.3	28775
			21913	77.2	6465	22.8	28378
			22366	77.3	6551	22.7	28917
2-GR	1	3	14342	89.2	1733	10.8	16075
			14284	89.8	1629	10.2	15913
			14408	88.9	1799	11.1	16207
	2	3	19993	95.4	957	4.6	20950
			19994	95.9	859	4.1	20853
			19913	95.2	995	4.8	20908
	3	3	18776	69.8	8138	30.2	26914
			18105	69.9	7796	30.1	25901
			18621	69.5	8173	30.5	26794
	4	3	17192	92.3	1430	7.7	18622
			17405	93.1	1292	6.9	18697
			17314	91.6	1593	8.4	18907
	5	3	21471	78.4	5925	21.6	27396
			21590	80.2	5326	19.8	26916
			22129	78.3	6116	21.7	28245

BC TABLE 113 (continued)

NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, VIABILITY AND ESTERASE ASSAY, INDIVIDUAL VALUES

Remarks: values from 2 to 3 evaluations of data from 1 FCM determination
total counts: ..... to .....

(a) sum of macrophages and granulocytes

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
2-GR	6	3	14342	90.3	1535	9.7	15877
			14328	91.3	1370	8.7	15698
			14630	90.1	1610	9.9	16240
	7	3	14913	94.3	899	5.7	15812
			15050	94.6	853	5.4	15903
			15044	93.7	1013	6.3	16057

BC TABLE 113 (continued)

NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM FWD VERSUS AXL/VIABILITY AND ESTERASE ASSAY, INDIVIDUAL VALUES

Remarks: values from 2 to 3 evaluations of data from 1 FCM determination
total counts: ..... to .....

(a) sum of macrophages and granulocytes

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	STATISTICAL PARAMETER RELATIVE NUMBER OF CELLS MACROPHAGES M (0/0)	SE	RSD (0/0)	GRANULOCYTES M (0/0)	SE	RSD (0/0)
0-GR	1	1	-	-	-	-	-	-
		3	96.4	0.4	0.8	3.6	0.4	20.4
	2	1	-	-	-	-	-	-
		3	98.9	0.2	0.3	1.1	0.2	25.7
	3	1	88.0	-	-	12.0	-	-
		3	90.6	0.6	1.2	9.4	0.6	11.2
	4	1	96.6	-	-	3.4	-	-
		3	99.3	0.1	0.1	0.7	0.1	14.0
	5	1	90.4	-	-	9.6	-	-
		3	86.9	0.0	0.1	13.1	0.0	0.5
	6	1	98.6	-	-	1.4	-	-
		3	98.7	0.2	0.3	1.3	0.2	21.3
	7	1	97.7	-	-	2.3	-	-
		3	99.2	0.1	0.1	0.8	0.1	17.4
1-GR	1	1	25.7	-	-	74.3	-	-
		3	40.3	0.3	1.4	59.7	0.3	1.0
	2	1	68.6	-	-	31.4	-	-
		3	76.6	0.6	1.4	23.4	0.6	4.5
	3	1	35.7	-	-	64.3	-	-
		3	26.0	0.2	1.6	74.0	0.2	0.5
	4	1	69.0	-	-	31.0	-	-
		3	59.4	0.1	0.4	40.6	0.1	0.5
	5	1	29.9	-	-	70.1	-	-
		3	22.7	0.1	0.9	77.3	0.1	0.3
	6	1	72.2	-	-	27.9	-	-
		3	56.9	0.2	0.7	43.1	0.2	1.0

BC TABLE 114

RELATIVE NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, VIABILITY AND ESTERASE ASSAY, STATISTICAL PARAMETERS FOR MULTIPLE EVALUATION

Remarks: statistical parameters for N = 3 (see BC TABLE 113)

[Fig. H03614, H03612]

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	STATISTICAL PARAMETER					
			RELATIVE NUMBER OF CELLS					
			MACROPHAGES			GRANULOCYTES		
			M (0/0)	SE	RSD (0/0)	M (0/0)	SE	RSD (0/0)
1-GR	7	1	88.8	-	-	11.2	-	-
		3	77.1	0.2	0.5	22.9	0.2	1.5
2-GR	1	3	89.3	0.3	0.5	10.7	0.3	4.1
	2	3	95.5	0.2	0.3	4.5	0.2	7.3
	3	3	69.7	0.1	0.3	30.3	0.1	0.7
	4	3	92.3	0.4	0.8	7.7	0.4	9.9
	5	3	79.0	0.6	1.4	21.0	0.6	5.1
	6	3	90.6	0.4	0.7	9.4	0.4	6.6
	7	3	94.2	0.3	0.5	5.8	0.3	8.3

BC TABLE 114 (continued)

RELATIVE NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, VIABILITY AND ESTERASE ASSAY, STATISTICAL PARAMETERS FOR MULTIPLE EVALUATION

Remarks: statistical parameters for N = 3 (see BC TABLE 113)

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	WEIGHTED MEAN RELATIVE NUMBER OF GRANULOCYTES POOL					
		1 AND 2	3 AND 4	5, 6 AND 7	8	9	10
0-GR	1	-	6.6	5.1	-	6.2	2.0
	3	2.6	2.1	3.2	1.9	3.5	1.0
1-GR	1	54.7	~~48.9~~ 49.0	34.2	54.3	46.1	16.9
	3	41.6	~~56.0~~ 55.9	43.2	39.7	57.2	30.3
2-GR	3	7.1	15.9	8.8	6.4	15.9	6.9

BC TABLE 115

RELATIVE NUMBER OF GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, VIABILITY AND ESTERASE ASSAY, WEIGHTED MEAN OF POOLS

Remarks: pool 1 to 7 weighted according to number of macrophages/~~and granulocytes per~~ pool/5 rats, resuspension medium, hemocytometer ~~(see BC-TABLE ...)~~, equivalent to combined respective pools
pool 8 to 10 weighted according to composition 3 : 7 of macrophages (see BC PAGE 4-1), equivalent to pool 8 to 10 used for phagocytosis and DNA assay

[vgl. Tab. 102]

[fig. H03358]

20290274444

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	WEIGHTED MEAN RELATIVE NUMBER OF MACROPHAGES POOL					
		1 AND 2	3 AND 4	5, 6 AND 7	8	9	10
0-GR	1	-	93.4	94.9	-	93.8	98.0
	3	97.4	97.9	96.8	98.1	96.5	99.0
1-GR	1	45.3	51.10	65.8	45.7	53.9	83.1
	3	58.4	44.01	56.8	60.3	42.8	69.7
2-GR	3	92.9	84.1	91.2	93.6	84.1	93.1

BC TABLE 116

RELATIVE NUMBER OF MACROPHAGES DETERMINED FROM FWD VERSUS AXL, VIABILITY AND ESTERASE ASSAY, WEIGHTED MEAN OF POOLS

Remarks: pool 1 to 7 weighted according to number of macrophages/~~and granulocytes per~~ pool/5 vials, resuspension medium hemocytometer ~~(see BC TABLE ..)~~, equivalent to combined respective pools [vgl. Tab. 102]
pool 8 to 10 weighted according to composition 3 : 7 of macrophages (see BC PAGE 4-1), equivalent to pool 8 to 10 used for phagocytosis and DNA assay

20290274S

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
0-GR	8	1	37363	98.4	611	1.6	37974
		2	40211	97.5	1044	2.5	41255
		3	11145	98.3	198	1.7	11343 (b)
	9	1	31037	98.0	633	2.0	31670
		2	39029	97.0	1205	3.0	40234
		3	39088	97.8	865	2.2	39953
	10	1	37610	99.4	240	0.6	37850
		2	34517	98.0	699	2.0	35216 (c)
		3	41535	98.1	807	1.9	42342
1-GR	8	1	12014	45.9	14177	54.1	26191 (d)
		2	6807	30.2	15754	69.8	22561
		3	15883	52.5	14388	47.5	30271
	9	1	19583	64.0	11004	36.0	30587
		2	16216	45.9	19085	54.1	35301
		3	14849	42.9	19730	57.1	34579
	10	1	21280	81.4	4852	18.6	26132
		2	21728	60.3	14319	39.7	36047
		3	12378	59.0	8599	41.0	20977
2-GR	8	3	-	-	-	-	-
	9	3	20142	81.5	4581	18.5	24723 (e)
	10	3	25999	88.5	3385	11.5	29384

BC TABLE 117

NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, DNA ASSAY, INDIVIDUAL VALUES

Remarks: number of signals in region ..... (macrophages) and ..... (granulocytes) (see BC FIGURE ..)
total number of signals analyzed: 50000, unless otherwise noted

(a) sum of macrophages and granulocytes
(b) total number of signals analyzed: 13602
(c) total number of signals analyzed: 43900
(d) total number of signals analyzed: 41496
(e) total number of signals analyzed: 30105

[Fig. H036 10]

2029027446

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF GRANULOCYTES (0/0) POOL 8	9	10
0-GR	N	3	3	3
	M (0/0)	1.96	2.39	1.51
	SE	0.29	0.31	0.44
	RSD (0/0)	25.2	22.2	50.4
1-GR	N	3	3	3
	M (0/0)	57.16	49.03	33.09
	SE	6.61	6.58	7.27
	RSD (0/0)	20.0	23.3	38.1
2-GR	N	-	1	1
	M (0/0)	-	18.5	11.5
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 118

RELATIVE NUMBER OF GRANULOCYTES DETERMINED FROM FWD VERSUS AXL, DNA ASSAY, STATISTICAL PARAMETERS

Remarks: see BC TABLE 117

20290274A7

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF MACROPHAGES (0/0)		
		POOL		
		8	9	10
0-GR	N	3	3	3
	M (0/0)	98.04	97.61	98.49
	SE	0.29	0.31	0.44
	RSD (0/0)	0.5	0.5	0.8
1-GR	N	3	3	3
	M (0/0)	42.84	50.97	66.91
	SE	6.61	6.58	7.27
	RSD (0/0)	26.7	22.4	18.2
2-GR	N	-	1	1
	M (0/0)	-	81.5	88.5
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 119

RELATIVE NUMBER OF MACROPHAGES DETERMINED FROM FWD VERSUS AXL CYTOGRAM, DNA ASSAY, STATISTICAL PARAMETERS

Remarks: see BC TABLE 117

20290274 48

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
0-GR	8	1	-	-	-	-	-
		2	6080	96.8	199	3.2	6279
			5109	96.0	211	4.0	5320
			6001	96.6	210	3.4	6211
		3	-	-	-	-	-
	9	1	7318	97.9	156	2.1	7474
			7344	95.8	325	4.2	7669
			7382	96.5	270	3.5	7652
		2	4566	96.8	153	3.2	4719
			4599	96.8	152	3.2	4751
			4575	96.4	172	3.6	4747
		3	-	-	-	-	-
	10	1	10517	99.7	31	0.3	10548
			10639	99.3	75	0.7	10714
			10630	99.7	36	0.3	10666
		2	6668	99.2	54	0.8	6722
			6655	99.0	70	1.0	6725
			6694	98.8	84	1.2	6778
		3	10825	98.6	159	1.4	10984
			10824	97.7	250	2.3	11074
			10846	98.5	165	1.5	11011
1-GR	8	1	-	-	-	-	-
		2	-	-	-	-	-
		3	-	-	-	-	-

BC TABLE 120

NUMBER OF MACROPHAGES AND GRANULOCYTES, PHAGOCYTOSIS ASSAY, INDIVIDUAL VALUES

Remarks: number obtained by 3 independent evaluations of list data from 1 FCM determination
total number of signals counted: 80000

(a) sum of macrophages and granulocytes

2029027449

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
1-GR	9	1	7751	50.2	7679	49.8	15430
			7835	49.8	7889	50.2	15724
			7972	49.5	8123	50.5	16095
		2	3131	33.4	6245	66.6	9376
			3211	34.4	6125	65.6	9336
			3233	34.0	6283	66.0	9516
		3	6179	39.9	9315	60.1	15494
			6198	40.2	9220	59.8	15418
			6266	40.2	9339	59.8	15605
	10	1	7549	79.3	1970	20.7	9519
			7601	78.7	2053	21.3	9654
			7645	78.7	2067	21.3	9712
		2	4763	54.7	3948	45.3	8711
			4882	55.3	3948	44.7	8830
			4896	55.1	3984	44.9	8880
		3	9381	63.6	5358	36.4	14739
			9384	63.8	5326	36.2	14710
			9546	64.2	5315	35.8	14861
2-GR	8	3	-	-	-	-	-
	9	3	7643	76.7	2323	23.3	9966
			7721	76.9	2323	23.1	10044
			7768	76.3	2409	23.7	10177
	10	3	9552	87.5	1370	12.5	10922
			9718	88.5	1258	11.5	10976
			9709	87.7	1361	12.3	11070

BC TABLE 120 (continued)

NUMBER OF MACROPHAGES AND GRANULOCYTES, PHAGOCYTOSIS ASSAY, INDIVIDUAL VALUES

Remarks: number obtained by 3 independent evaluations of list data from 1 FCM determination
total number of signals counted: 80000

(a) sum of macrophages and granulocytes

202902745O

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	STATISTICAL PARAMETER RELATIVE NUMBER OF CELLS MACROPHAGES M (0/0)	SE	RSD (0/0)	GRANULOCYTES M (0/0)	SE	RSD (0/0)
0-GR	8	1	-	-	-	-	-	-
		2	96.5	0.2	0.4	3.5	0.2	11.8
		3	-	-	-	-	-	-
	9	1	96.7	0.6	1.1	3.3	0.6	33.3
		2	96.6	0.1	0.2	3.4	0.1	6.9
		3	-	-	-	-	-	-
	10	1	99.6	0.1	0.2	0.4	0.1	50.5
		2	99.0	0.1	0.2	1.0	0.1	21.5
		3	98.3	0.3	0.5	1.7	0.3	26.1
1-GR	8	1	-	-	-	-	-	-
		2	-	-	-	-	-	-
		3	-	-	-	-	-	-
	9	1	49.9	0.2	0.7	50.1	0.2	0.7
		2	33.9	0.3	1.5	66.1	0.3	0.8
		3	40.1	0.1	0.4	59.9	0.1	0.3
	10	1	78.9	0.2	0.4	21.1	0.2	1.6
		2	55.0	0.2	0.6	45.0	0.2	0.7
		3	63.9	0.2	0.5	36.1	0.2	0.9
2-GR	8	3	-	-	-	-	-	-
	9	3	76.6	0.2	0.4	23.4	0.2	1.2
	10	3	87.9	0.3	0.6	12.1	0.3	4.7

BC TABLE 121

RELATIVE NUMBER OF MACROPHAGES AND GRANULOCYTES, PHAGOCYTOSIS ASSAY, STATISTICAL PARAMETERS FOR MULTIPLE EVALUATION

Remarks: statistical parameters for N = 3 (see BC TABLE 139)
pool 3 and 4 represents the mean of pool 3 and pool 4 weighted according to the absolute number of cells per pool (see BC TABLE ..)
pool 9 represents the mean of pool 3 and pool 4 weighted according to composition 3 : 7 of macrophages (see

[Fig. 1103608

2029027451

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF GRANULOCYTES (0/0) POOL 8	9	10
0-GR	N	1	2	3
	M (0/0)	3.5	3.4	1.0
	SE	-	-	0.4
	RSD (0/0)	-	-	63.0
1-GR	N	-	3	3
	M (0/0)	-	58.7	34.1
	SE	-	4.7	7.0
	RSD (0/0)	-	13.7	35.5
2-GR	N	-	1	1
	M (0/0)	-	23.4	12.1
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 122

RELATIVE NUMBER OF GRANULOCYTES, PHAGOCYTOSIS ASSAY, STATISTICAL PARAMETERS FOR MULTIPLE DETERMINATIONS

Remarks: see BC TABLE 121

20290274S2

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF ~~GRANULOCYTES~~ MACROPHAGES (0/0)		
		POOL		
		8	9	10
0-GR	N	1	2	3
	M (0/0)	96.5	96.7	99.0
	SE	-	-	0.4
	RSD (0/0)	-	-	0.7
1-GR	N	-	3	3
	M (0/0)	-	41.3	65.9
	SE	-	4.7	7.0
	RSD (0/0)	-	19.5	18.3
2-GR	N	-	1	1
	M (0/0)	-	76.6	87.9
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 123

RELATIVE NUMBER OF MACROPHAGES, PHAGOCYTOSIS ASSAY, STATISTICAL PARAMETERS FOR MULTIPLE DETERMINATIONS

Remarks: see BC TABLE 121

2029027453

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
0-GR	1	1 (b)	-	-	-	-	-
		3	30598	98.59	439	1.41	31037
	2	1 (b)	-	-	-	-	-
		3	17842	99.48	94	0.52	17936
	3	1	5615	98.42	90	1.58	5705
		3	17877	98.47	277	1.53	18154
	4	1	13174	99.17	110	0.83	13284
		3	21995	99.77	51	0.23	22046
	5	1	2480	98.14	47	1.86	2527
		3	20648	97.80	465	2.20	21113
	6	1	10371	99.30	73	0.70	10444
		3	15570	98.98	161	1.02	15731
	7	1	4161	99.26	31	0.74	4192
		3	15665	99.33	105	0.67	15770
1-GR	1	1	1013	16.09	5281	83.91	6294
		3	11239	37.93	18538	62.07	29867
	2	1	5050	65.94	2608	34.06	7658
		3	15797	76.49	4856	23.51	20653
	3	1	2733	33.08	5529	66.92	8262
		3	7367	24.93	22187	75.07	29554
	4	1	13325	68.15	6227	31.85	19552
		3	14404	57.55	10625	42.45	25029
	5	1	787	23.00	2634	77.00	3421
		3	6732	21.01	25310	78.99	32042

BC TABLE 124

NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM ESTERASE ACTIVITY, VIABILITY AND ESTERASE ASSAY, INDIVIDUAL VALUES

Remarks: for details of determination (see

(a) sum of macrophages and granulocytes
(b)

[Fig. H036.13]

2029027454

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
1-GR	6	1	13417	72.10	5192	27.90	18609
		3	23583 ~~25538~~	56.03	18508	43.97	42091
	7	1	18746	89.04	2308	10.96	21054
		3	20296	76.17	6350	23.83	26646
2-GR	1	3	12960	93.66	878	6.34	13838
	2		18219	96.52	656	3.48	18875
	3		15895	70.96	6504	29.04	22399
	4		14810	94.36	886	5.64	15696
	5		18126	79.93	4550	20.07	22676
	6		12945	91.89	1143	8.11	14088
	7		12880	94.77	711	5.23	13591

BC TABLE 124 (continued)

NUMBER OF MACROPHAGES AND GRANULOCYTES DETERMINED FROM ESTERASE ACTIVITY, VIABILITY AND ESTERASE ASSAY, INDIVIDUAL VALUES

Remarks: for details of determination (see

(a) sum of macrophages and granulocytes

2029027455

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B8 WS

GROUP	EXP. NO.	WEIGHTED MEAN RELATIVE NUMBER OF GRANULOCYTES (0/0) POOL					
		1 AND 2	3 AND 4	5, 6 AND 7	8	9	10
0-GR	1	-	1.1	1.2	-	1.1	0.7
	3	1.0	0.4	1.0	0.8	0.6	0.8
1-GR	1	66.3	51.4	40.1	65.8	48.3	16.8
	3	43.5	57.5	45.0	41.4	58.7	31.2
2-GR	3	4.5	14.1	8.0	4.4	14.1	6.1

BC TABLE 125

RELATIVE NUMBER OF GRANULOCYTES DETERMINED FROM ESTERASE ACTIVITY, VIABILITY AND ESTERASE ASSAY, WEIGHTED MEAN OF POOLS

Remarks: pools 1 to 7 weighted according to number of macrophages /~~and granulocytes per~~ pool/Ratsuspensionsmedium, hemocytometer ~~(see BC TABLE ..)~~, equivalent to combined respective pools
pools 8 to 10 weighted according to composition 3 : 7 of macrophages (see BC PAGE 4-1), equivalent to pools 8 to 10 used for phagocytosis and DNA assays

see BC TABLE 124

[Fig H036/11]

20290274S6

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXP. NO.	WEIGHTED MEAN RELATIVE NUMBER OF MACROPHAGES (0/0) POOL					
		1 AND 2	3 AND 4	5, 6 AND 7	8	9	10
0-GR	1	-	98.9	98.8	-	98.9	99.3
	3	99.0	99.6	99.0	99.2	99.4	99.2
1-GR	1	33.7	48.6	59.9	34.2	51.7	83.2
	3	56.5	42.5	55.0	58.6	41.3	68.8
2-GR	3	95.5	85.9	92.0	95.6	85.9	93.9

BC TABLE 126

RELATIVE NUMBER OF MACROPHAGES DETERMINED FROM ESTERASE ACTIVITY, VIABILITY AND ESTERASE ASSAY, WEIGHTED MEAN OF POOLS

Remarks: pools 1 to 7 weighted according to number of macrophages ~~and granulocytes per~~ pool (Brutt, resuspension medium, hemocytometer ~~(see BC TABLE ..)~~), equivalent to combined respective pools
pools 8 to 10 weighted according to composition 3 : 7 of macrophages (see BC PAGE 4-1), equivalent to pools 8 to 10 used for phagocytosis and DNA assays
see BC TABLE 124

20290274S7

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	EXP. NO.	NUMBER OF CELLS				
			MACROPHAGES		GRANULOCYTES		TOTAL (a)
			ABSOLUTE	REL. TO TOTAL (0/0)	ABSOLUTE	REL. TO TOTAL (0/0)	
0-GR	8	1	37168	98.6	518	1.4	37686
		2	39944	98.0	812	2.0	40756
		3	11093	98.4	176	1.6	11269 (b)
	9	1	30693	98.9	327	1.1	31020
		2	38763	97.6	949	2.4	39712
		3	38871	98.2	718	1.8	39589
	10	1	37492	99.6	164	0.4	37656
		2	34350	98.3	593	1.7	34943 (c)
		3	41493	98.4	679	1.6	42172
1-GR	8	1	11967	46.2	13941	53.8	25908 (d)
		2	6638	30.6	15071	69.4	21709
		3	15823	53.3	13847	46.7	29670
	9	1	19243	64.4	10635	35.6	29878
		2	16052	46.2	18666	53.8	34718
		3	14784	43.2	19426	56.8	34210
	10	1	21106	82.8	4381	17.2	25487
		2	21601	60.9	13876	39.1	35477
		3	12271	59.9	8229	40.1	20500
2-GR	8	3	-	-	-	-	-
	9	3	20029	81.9	4421	18.1	24450 (e)
	10	3	25875	89.2	3128	10.8	29003

BC TABLE 127

NUMBER OF MACROPHAGES AND GRANULOCYTES, DNA-GATED SIGNALS, DNA ASSAY, INDIVIDUAL VALUES

Remarks: number of signals in region (see BC FIGURE)
total number of signals analyzed: 50000, unless otherwise noted

(a) sum of macrophages and granulocytes
(b) total number of signals analyzed: 13602
(c) total number of signals analyzed: 43900
(d) total number of signals analyzed: 41496
(e) total number of signals analyzed: 30105

[Fig. H03603, H03615

20290274

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF GRANULOCYTES (0/0) POOL 8	9	10
0-GR	N	3	3	3
	M	1.64	1.75	1.25
	SE	0.18	0.39	0.41
	RSD (0/0)	19.4	38.4	56.2
1-GR	N	3	3	3
	M	56.63	48.71	32.15
	SE	6.72	6.62	7.48
	RSD (0/0)	20.5	23.5	40.3
2-GR	N	-	1	1
	M	-	18.1	10.8
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 128

RELATIVE NUMBER OF GRANULOCYTES, DNA-GATED SIGNALS, DNA ASSAY, STATISTICAL PARAMETERS

Remarks: number of signals in region (see BC FIGURE)
see BC TABLE 127

20290274S9

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	STATISTICAL PARAMETER	RELATIVE NUMBER OF ~~GRANULOCYTES~~ MACROPHAGES (0/0)		
		POOL		
		8	9	10
0-GR	N	3	3	3
	M	98.36	98.25	98.75
	SE	0.18	0.39	0.41
	RSD (0/0)	0.3	0.7	0.7
1-GR	N	3	3	3
	M	43.37	51.29	67.85
	SE	6.72	6.62	7.48
	RSD (0/0)	26.8	22.3	19.1
2-GR	N	-	1	1
	M	-	81.9	89.2
	SE	-	-	-
	RSD (0/0)	-	-	-

BC TABLE 129

RELATIVE NUMBER OF MACROPHAGES, DNA-GATED SIGNALS, DNA ASSAY, STATISTICAL PARAMETERS

Remarks: number of signals in region (see BC FIGURE)

see BC TABLE 127

20290277460

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	EXPERIMENT NO.	RELATIVE DIFFERENCE (0/0) POOL		
		8	9	10
0-GR	1	14.29	81.82	50.0
	2	25.0	25.0	17.65
	3	6.25	22.22	18.75
1-GR	1	0.56	1.12	8.14
	2	0.58	0.56	1.53
	3	1.71	0.53	2.24
2-GR	3	-	2.21	6.48

BC TABLE 130

RELATIVE DIFFERENCE BETWEEN THE RELATIVE NUMBER OF GRANULOCYTES OBTAINED WITHOUT AND WITH DNA-GATING, DNA ASSAY

Remarks: relative number of ungated (see BC TABLE 117) minus relative number of DNA-gated (see BC ~~FIGURE~~ Table 127) granulocyte signals relative to relative number of DNA-gated granulocyte signals

[Fig. 403615]

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

GROUP	POOL	RATIO (COUNTS/MACROPHAGE) COUNTS REGION ... (a) EXP. 1	EXP. 3	(RED) BLOOD CELLS (b) EXP. 1	EXP. 3
0-GR	1	-	0.42	0.51	0.14
	2	-	0.05	0.03	0.00
	3	~~2.19~~ 1,46	1.63	1.21	0.94
	4	0.20	0.04	0.09	0.02
	5	0.80	1.17	0.22	1.07
	6	0.06	0.05	0.01	0.04
	7	0.04	0.02	0.01	0.01
1-GR	1	0.92	0.58	1.53	0.03
	2	0.18	0.07	0.07	0.10
	3	0.11	1.29	0.06	0.92
	4	0.01	~~0.68~~ 0.10	0.01	0.17
	5	0.89	2.29	0.56	0.90
	6	0.05	0.21	0.04	0.15
	7	0.02	0.04	0.02	0.01
2-GR	1	-	0.82	-	0.34
	2	-	0.14	-	0.16
	3	-	0.51	-	0.26
	4	-	0.05	-	0.03
	5	-	0.47	-	0.73
	6	-	0.08	-	0.04
	7	-	0.04	-	0.02

BC TABLE 131

RATIO OF RBC PER MACROPHAGE DERIVED FROM FCM AND MICROSCOPIC DATA

Remarks: counts in region ... considered to represent mainly red blood cells

(a) FCM (see .....and BC FIGURE..)
(b) microscope counting, cytocentrifuge preparation

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FIGURES

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SUBREPORT P 0500/3057 GD151 (R) B15 WS M

BC FIGURE 1

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 0-GR

Remarks: A: pool 1, B: pool 2
representative field, resuspension medium, experiment 1

SUBREPORT P 0500/3057 GD151 (R) B15 WS M

BC FIGURE 2

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 1-GR

Remarks: A: pool 1, B: pool 2
representative field, resuspension medium, experiment 1

SUBREPORT P 0500/3057 GD151 (R) B15 WS M

BC FIGURE 3

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 0-GR

Remarks: A: pool 3, B: pool 4
representative field, resuspension medium, experiment 3

SUBREPORT P 0500/3057 GD151 (R) B15 WS M

BC FIGURE 4

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 0-GR

Remarks: A: pool 5, B: pool 6, C: pool 7
representative field, resuspension medium, experiment 3

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

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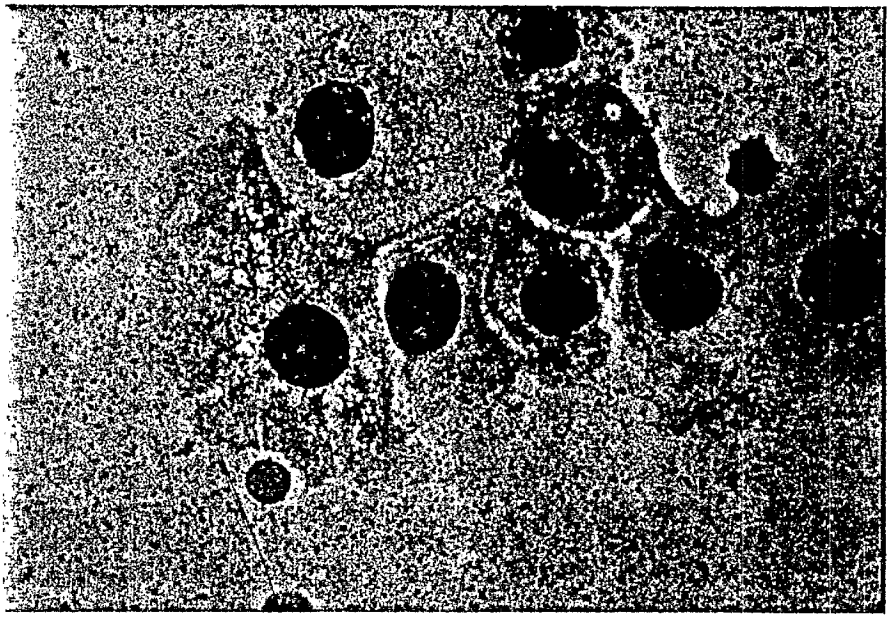

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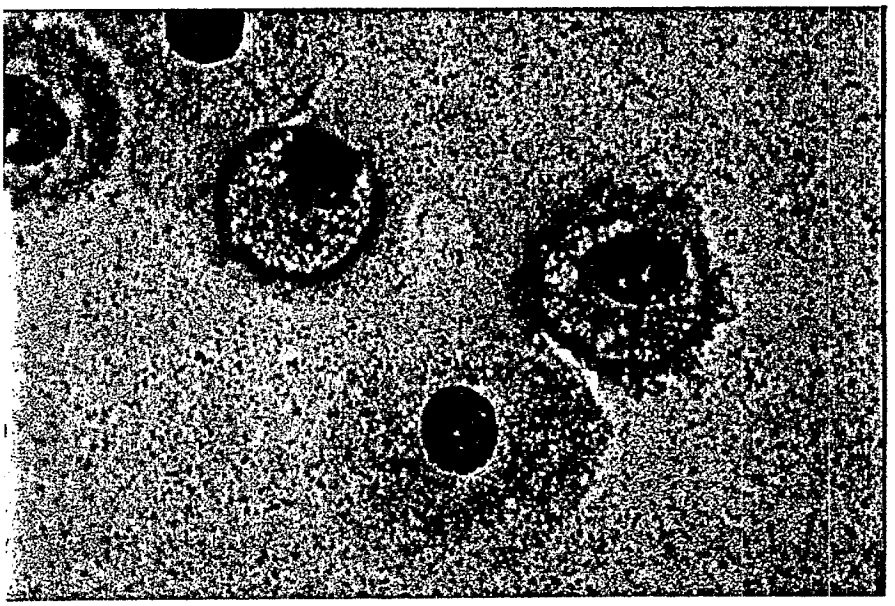

B

3C FIGURE 1

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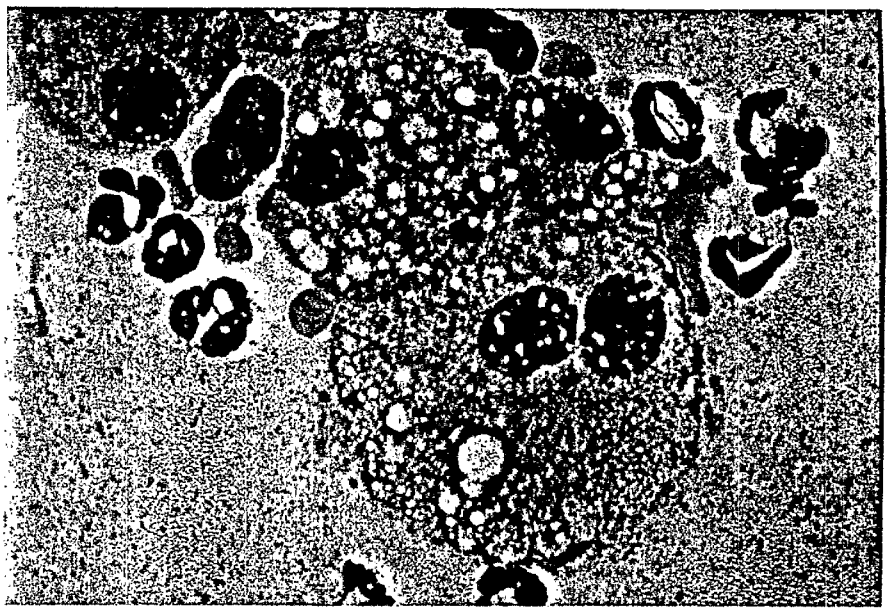

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BC FIGURE 2

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

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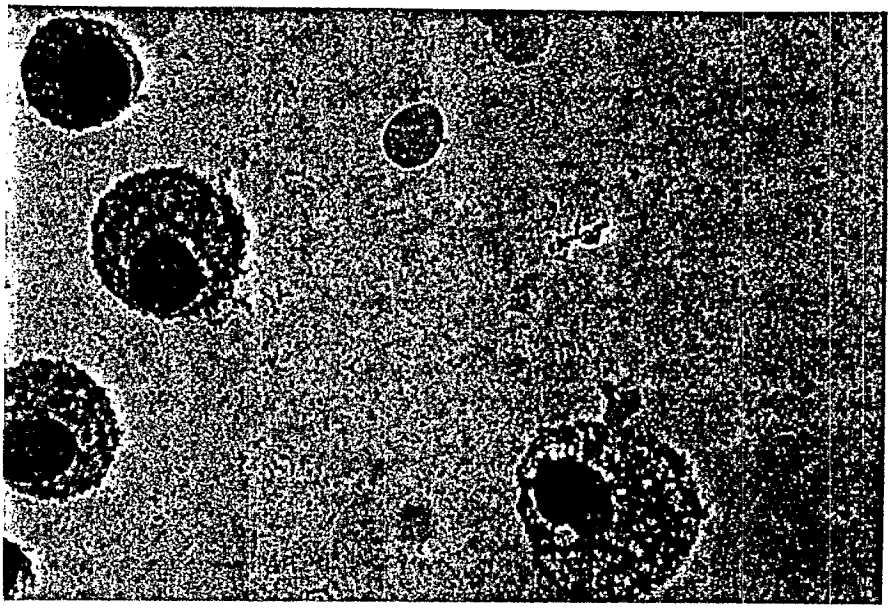

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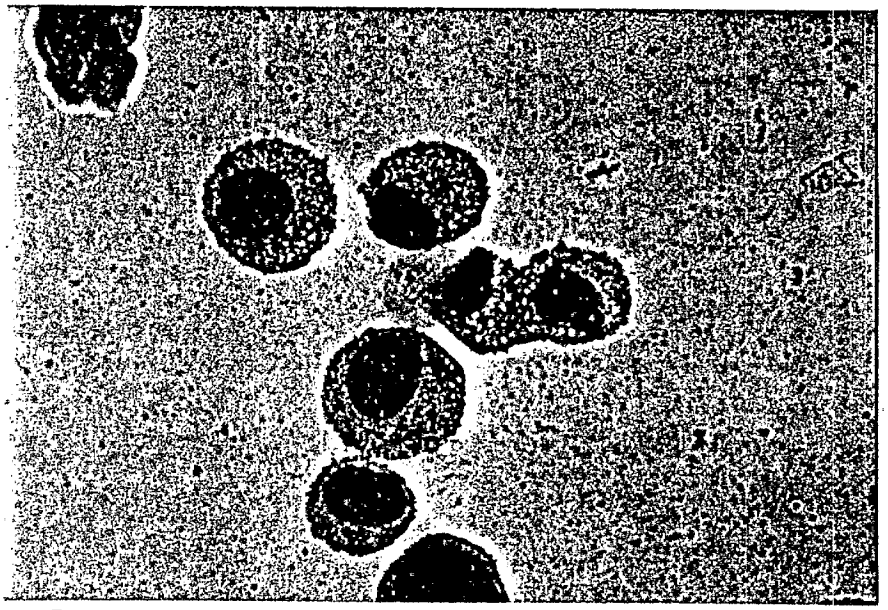

B

BC FIGURE 3

45. KW 83

Fortsetzung auf Blatt:

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

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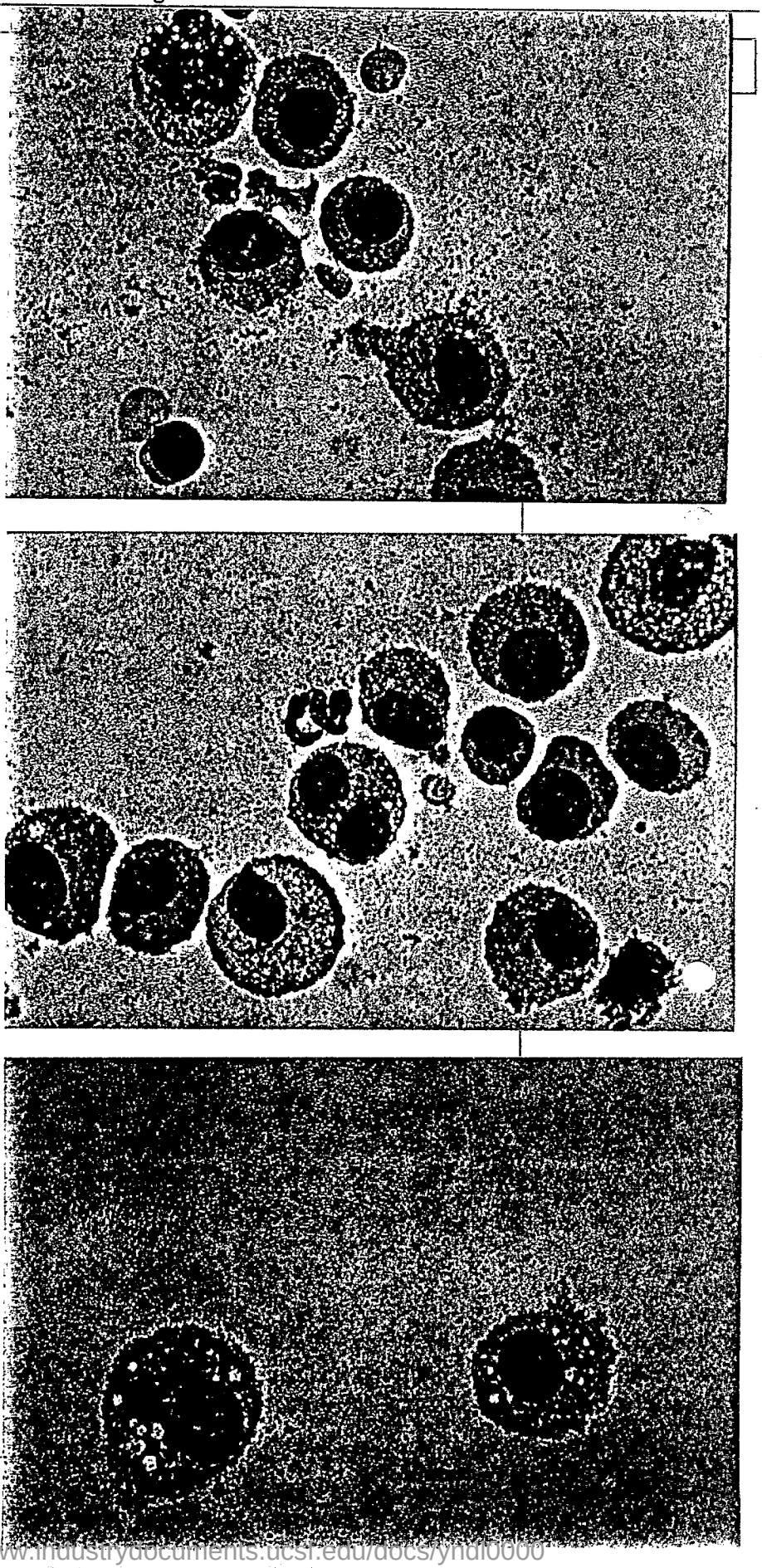
C

FIGURE 4



Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B16 WS M

BC FIGURE 5

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 1-GR

Remarks: A: pool 3, B: pool 4
representative field, resuspension medium, experiment 3

SUBREPORT P 0500/3057 GD151 (R) B16 WS M

BC FIGURE 6

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 1-GR

Remarks: A: pool 5, B: pool 6, C: pool 7
representative field, resuspension medium, experiment 3

SUBREPORT P 0500/3057 GD151 (R) B16 WS M

BC FIGURE 7

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 2-GR

Remarks: A: pool 3, B: pool 4
representative field, resuspension medium, experiment 3

SUBREPORT P 0500/3057 GD151 (R) B16 WS M

BC FIGURE 8

FREE LUNG CELLS IN CYTOCENTRIFUGE PREPARATIONS, 0-GR

Remarks: A: pool 5, B: pool 6, C: pool 7
representative field, resuspension medium, experiment 3

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

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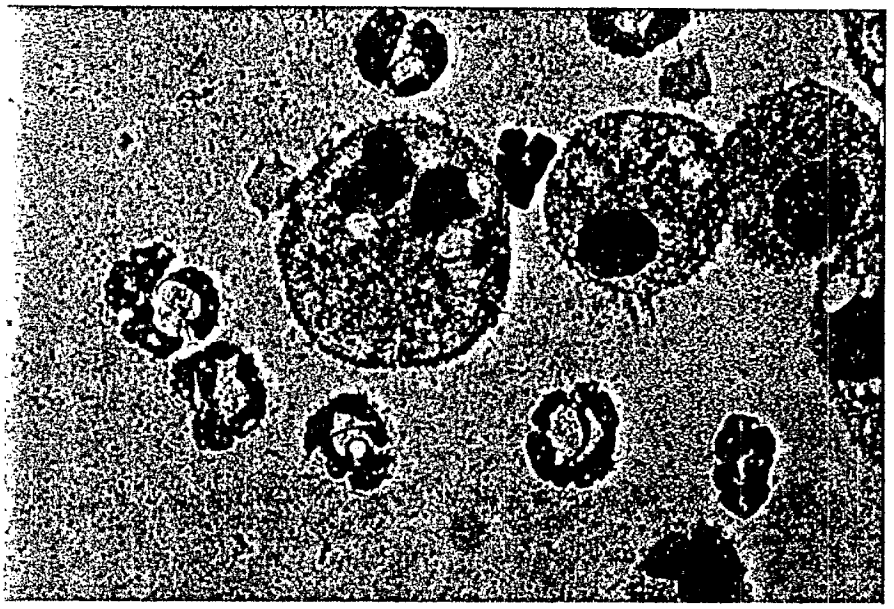

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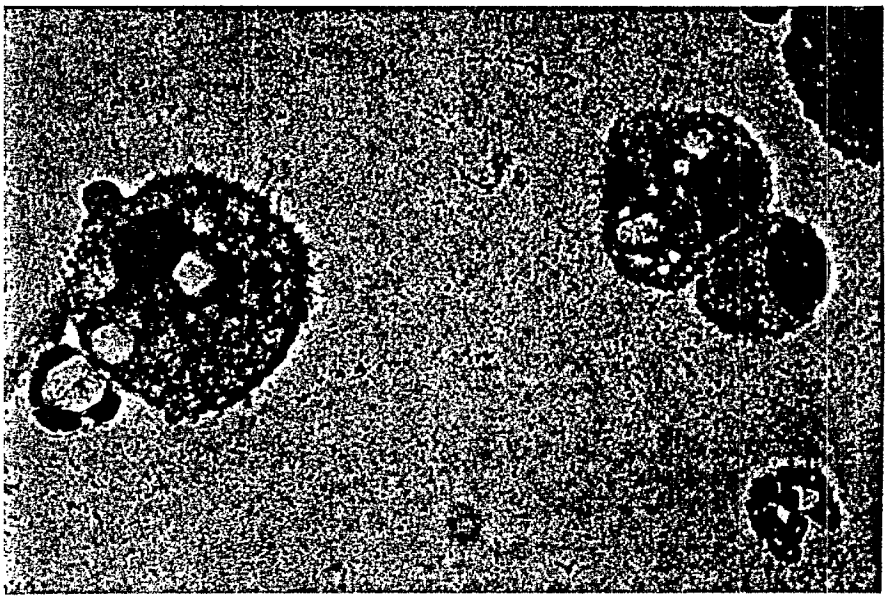

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1-6R

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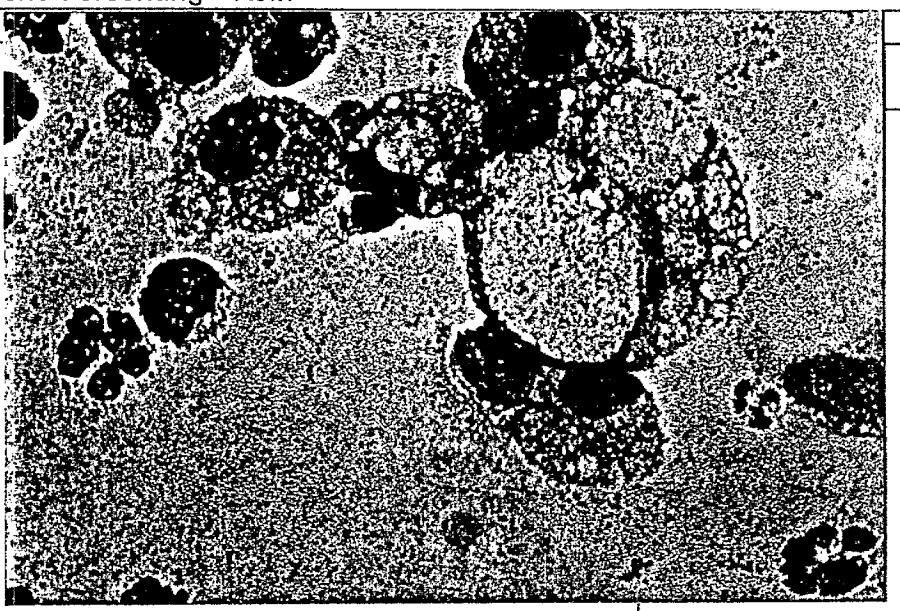
BC FIGURE 6



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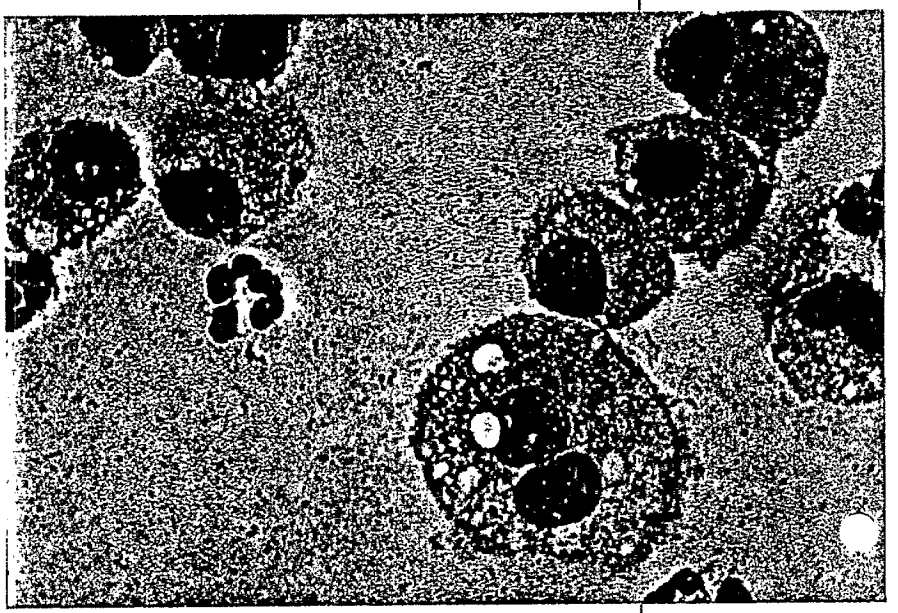

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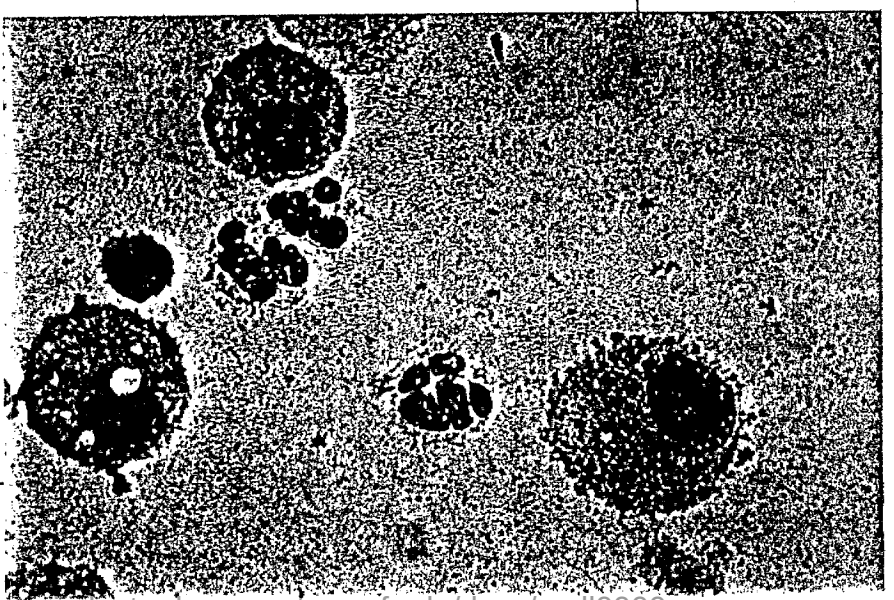

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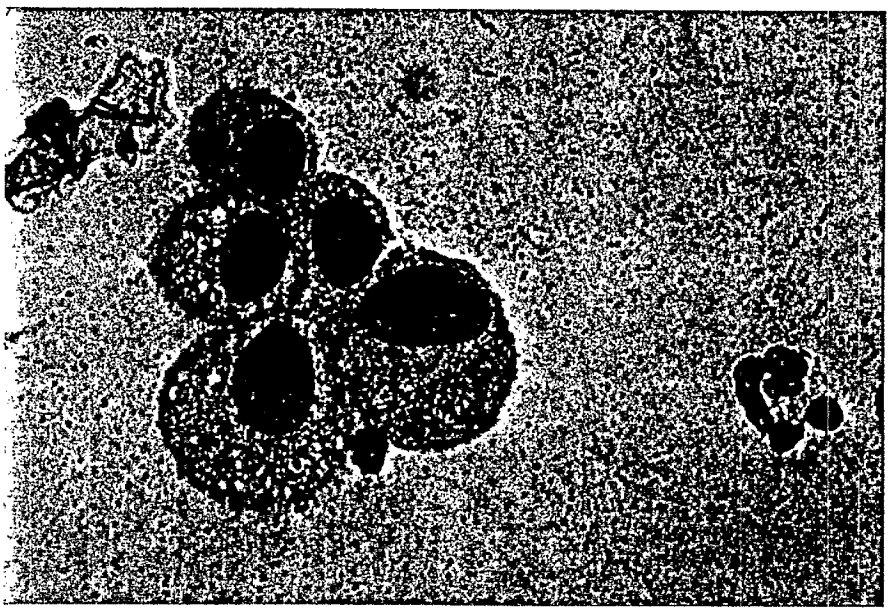

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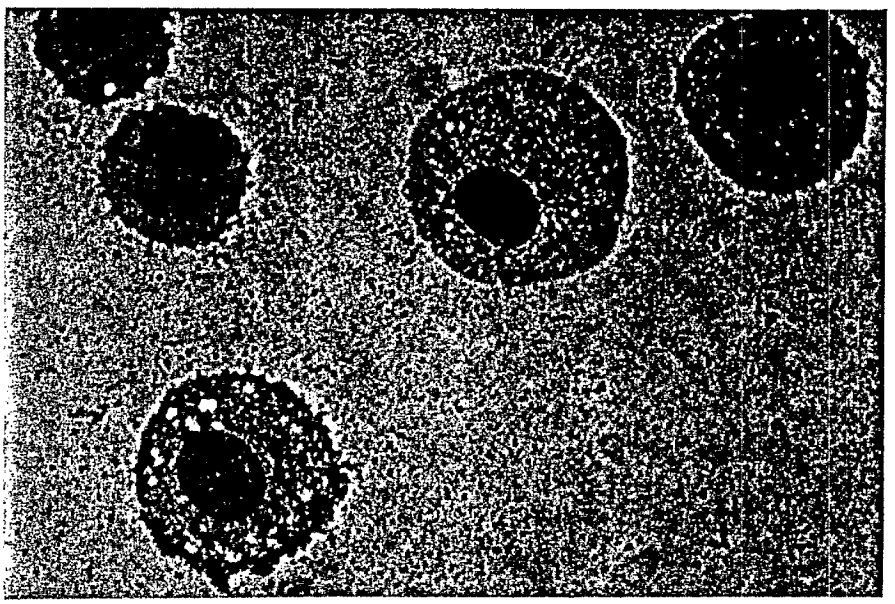

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BC FIGURE 7

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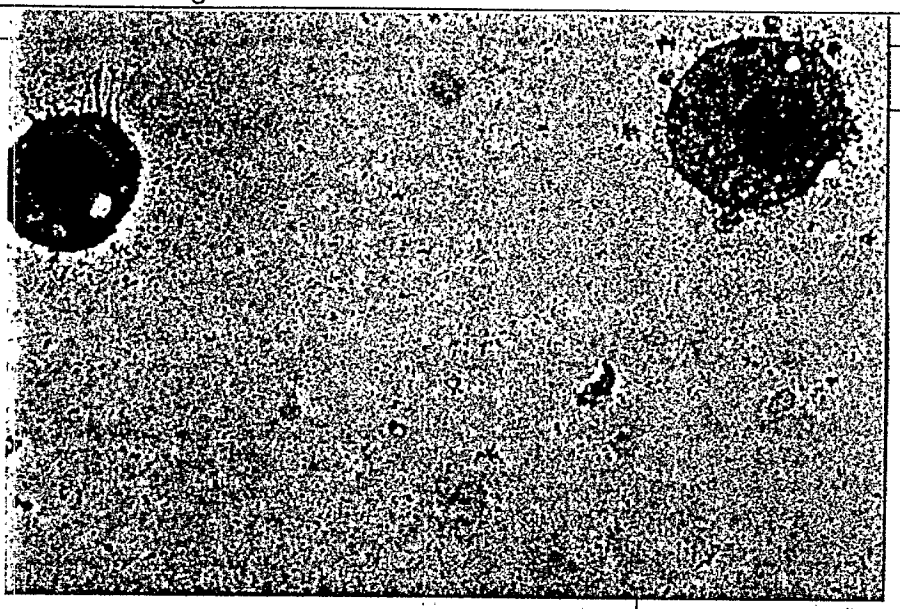
A
BC FIGURE 8



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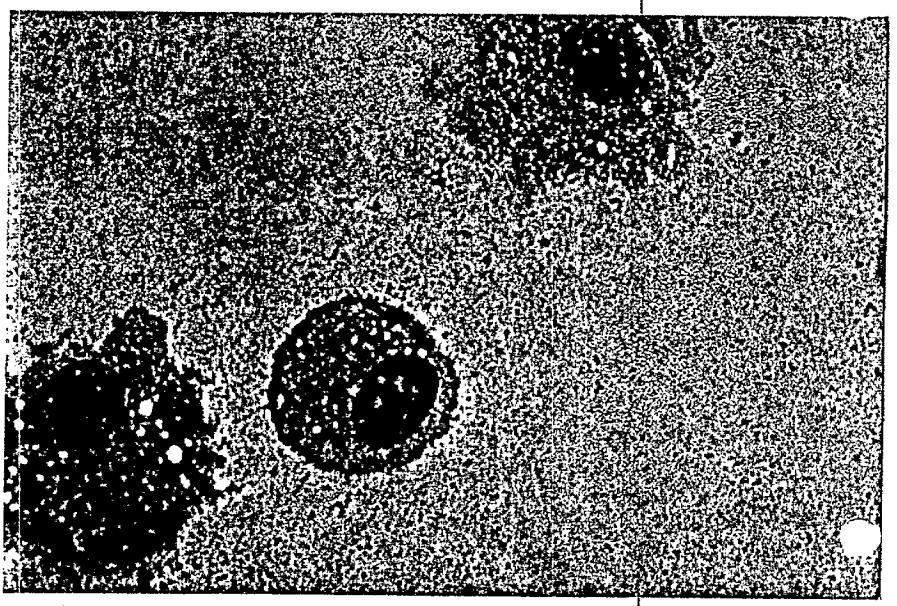

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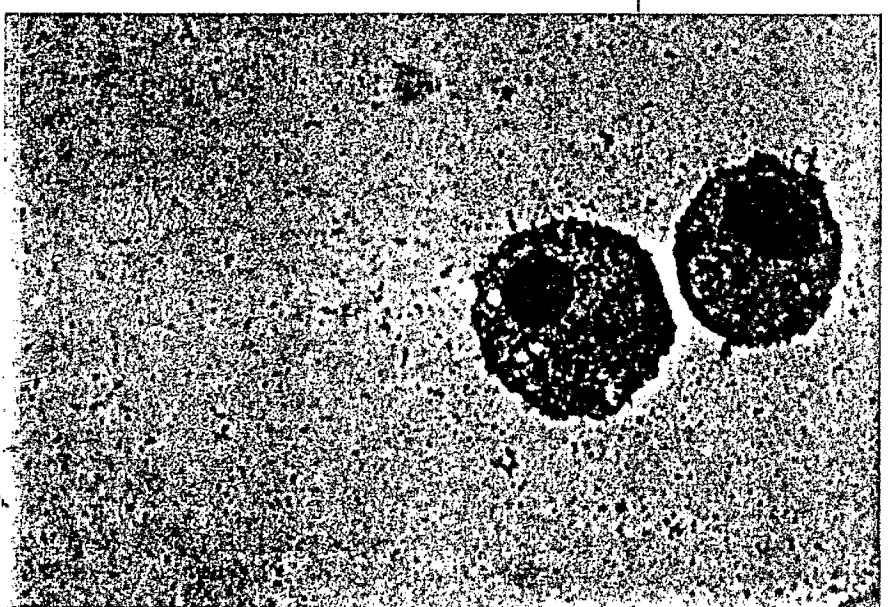

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

BC FIGURE 9

SDS-PAGE PROTEIN PATTERNS OF VARIOUS POOLS OF FLC, SILVER STAIN

Remarks:

SLOT	GROUP	POOL	PROTEIN (ug)
1	standards	-	-
2	1.2.2	4	0.91
3	standards	-	-
4	1.3.2	7	1.14
5	0.2.3	4	1.38
6	0.3.3	5	1.48
7	0.3.3	6	0.94
8	0.3.3	7	1.07
9	standards	-	-

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

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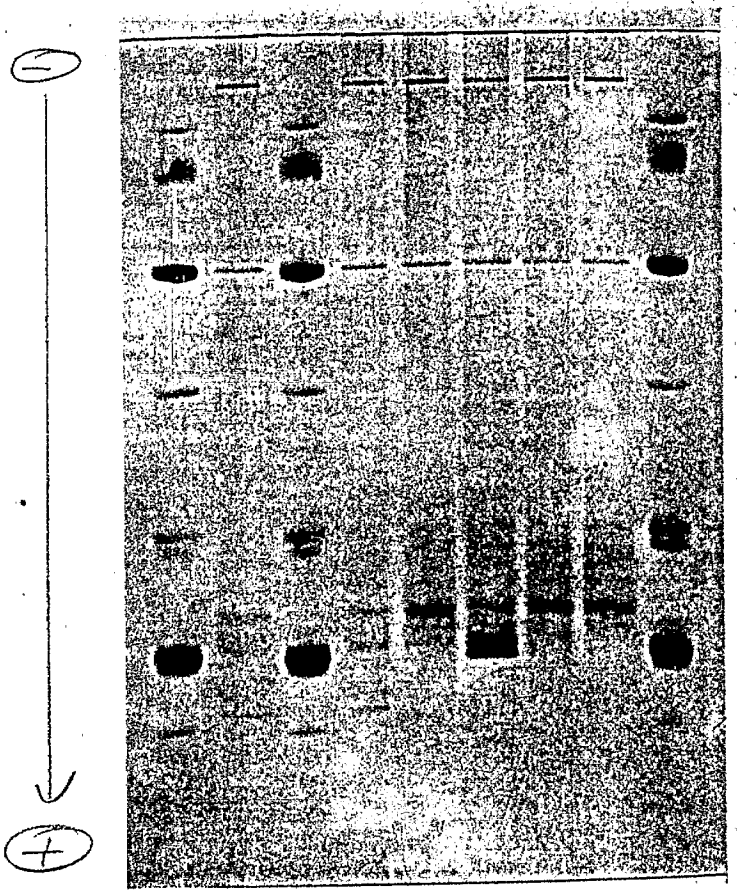

SLOT	GROUP	POOL	PROTEIN (µg)
1	STANDARDS	—	—
2	1.2.2	4	0.91
3	STANDARDS	—	—
4	1.3.2	7	1.14
5	0.2.3	4	1.38
6	0.3.3	5	1.48
7	0.3.3	6	0.94
8	0.3.3	7	1.07
9	STANDARDS	—	—

BC FIGURE 9

45. KW 83

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Source: https://www.industrydocuments.ucsf.edu/docs/...

BC FIGURE 10

SDS-PAGE PROTEIN PATTERNS OF VARIOUS POOLS OF FLC, COOMASSIE BLUE STAIN

Remarks: ——————————————————————

SLOT	GROUP	POOL	PROTEIN (ug)
1	-	-	-
2	1.1.2	2	4.86
3	1.2.2	4	4.24
4	1.3.2	7	5.31
5	0.2.3	4	3.21
6	0.3.3	7	2.49
7	1.2.3	4	5.26
8	1.3.3	7	3.51
9	2.2.3	4	5.09
10	2.3.3	7	3.21
11	standards	-	-

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

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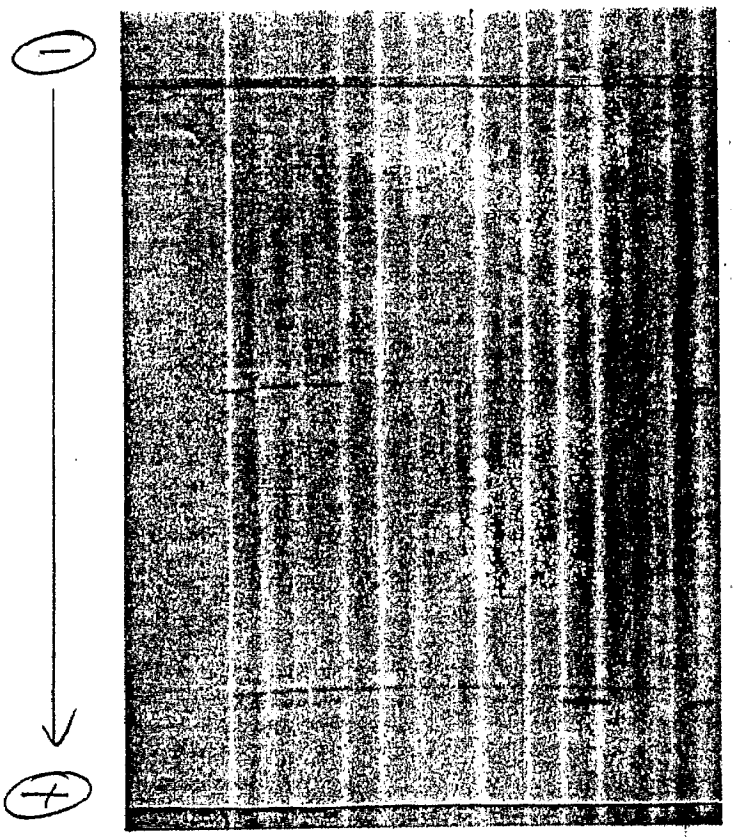

SLOT	GROUP	POOL	PROTEIN (µg)
1	—	—	—
2	1.1.2	2	4.86
3	1.2.2	4	4.24
4	1.3.2	7	5.31
5	0.2.3	4	3.21
6	0.3.3	7	2.49
7	1.2.3	4	5.26
8	1.3.3	7	3.51
9	2.2.3	4	5.09
10	2.3.3	7	3.21
11	~~Mwkr~~ Standards	—	—

BC FIGURE 1C

45. KW 83

Fortsetzung auf Blatt:

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B19 WS M HO3448

BC FIGURE 11

FATTY ACID METHYL ESTERS FROM FLC, INDIVIDUAL POOLS

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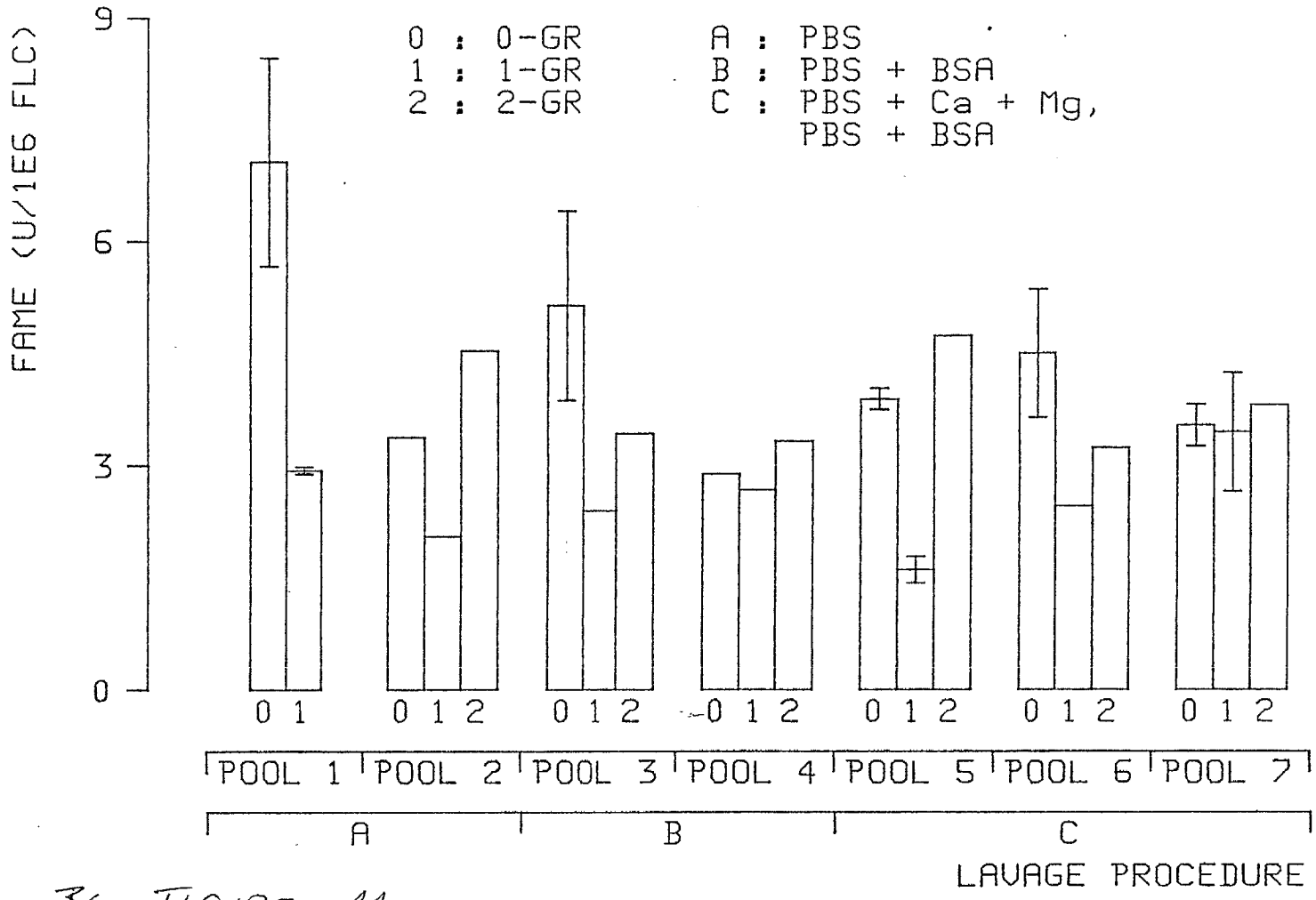
Remarks: for details see BC TABLE 11

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 11

FATTY ACID METHYL ESTERS FROM FLC, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 11

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Work Table 11 unrelated Report/Exp

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

BC FIGURE 12

FATTY ACID METHYL ESTERS FROM FLC, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 11

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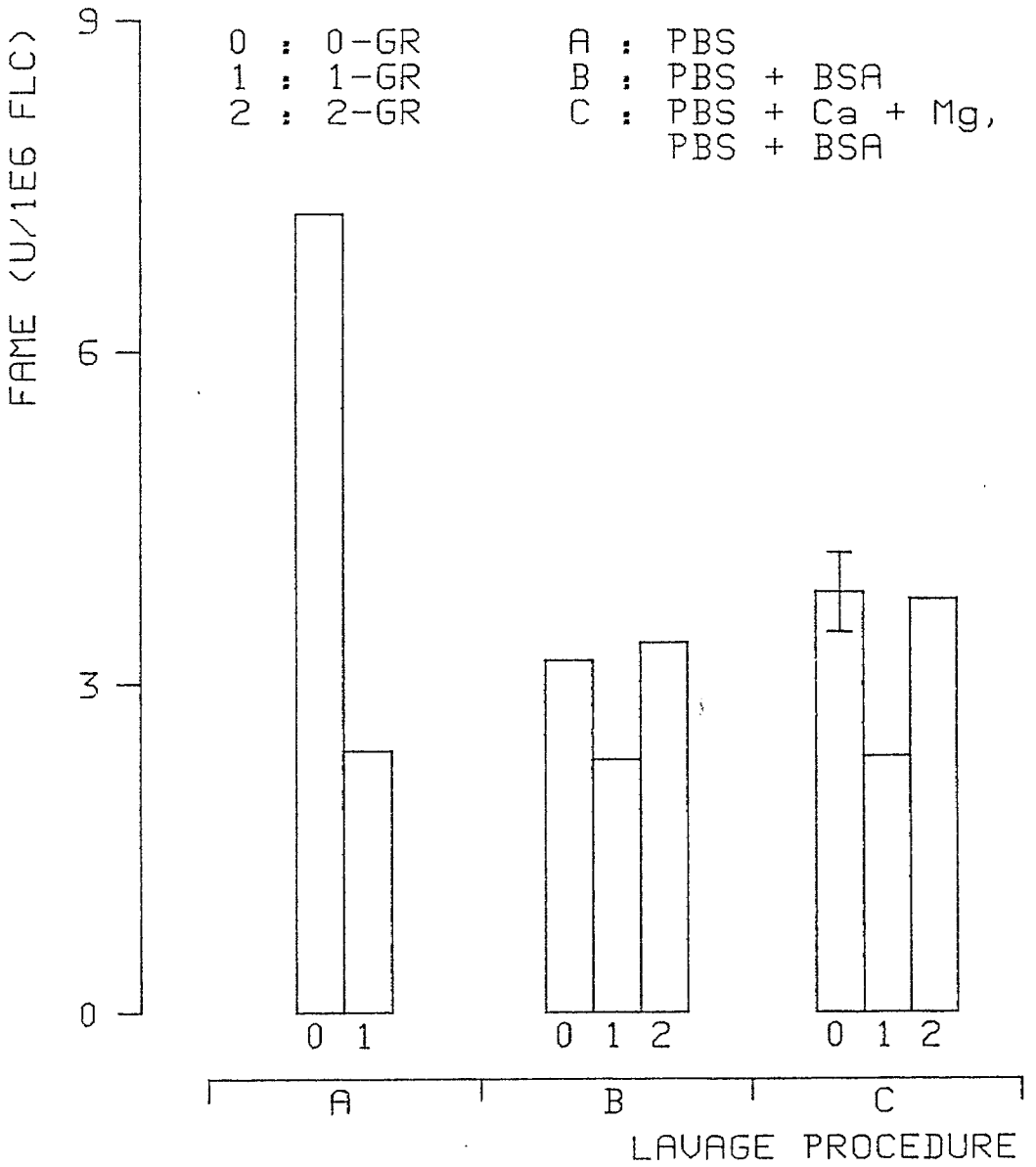

BC FIGURE 12

FATTY ACID METHYL ESTERS FROM FLC, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 11

A 0500/3057, H03449, TH, V138 F155 '94

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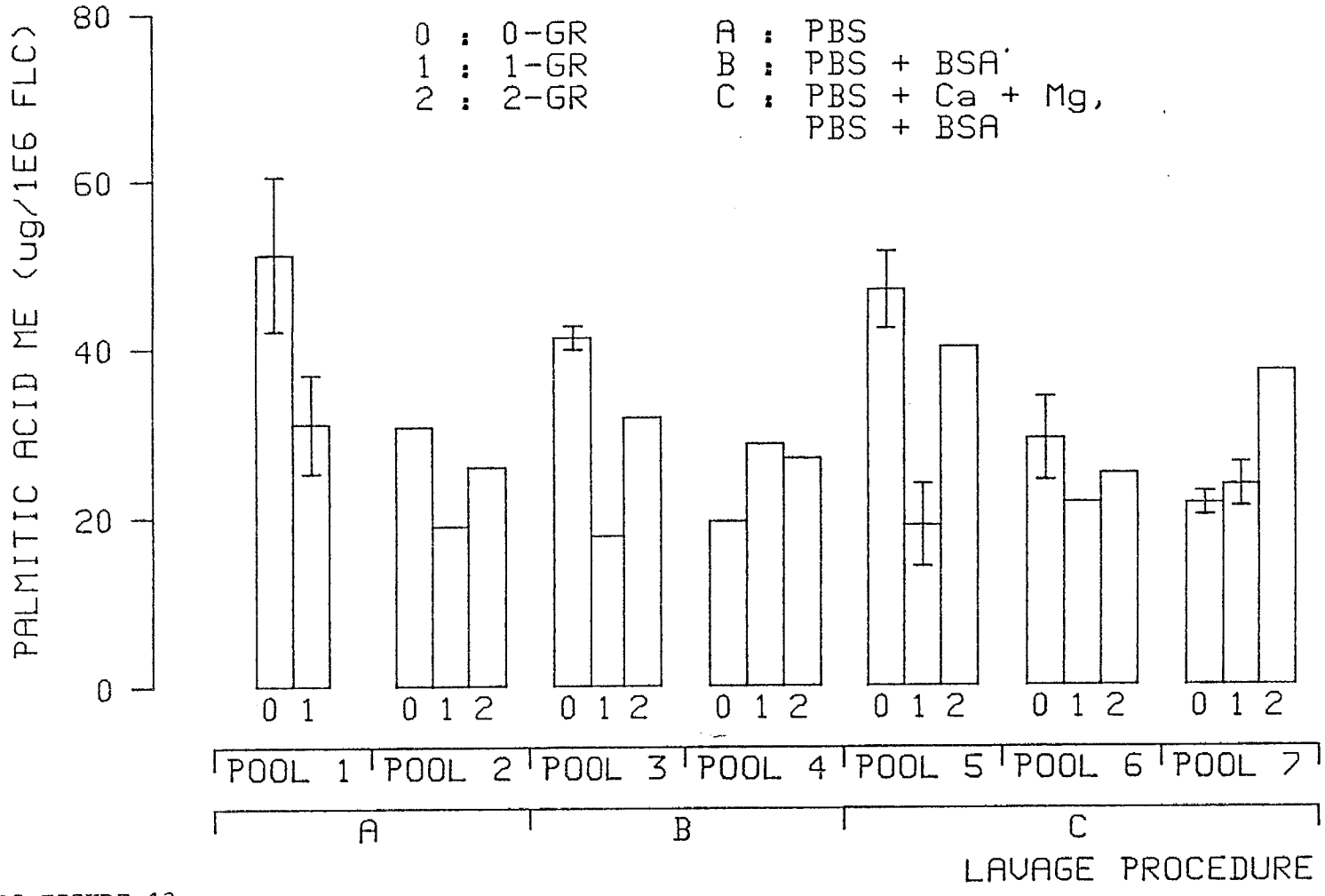

BC FIGURE 13

PALMITIC ACID METHYL ESTER (16 : 0) FROM FLC, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 13

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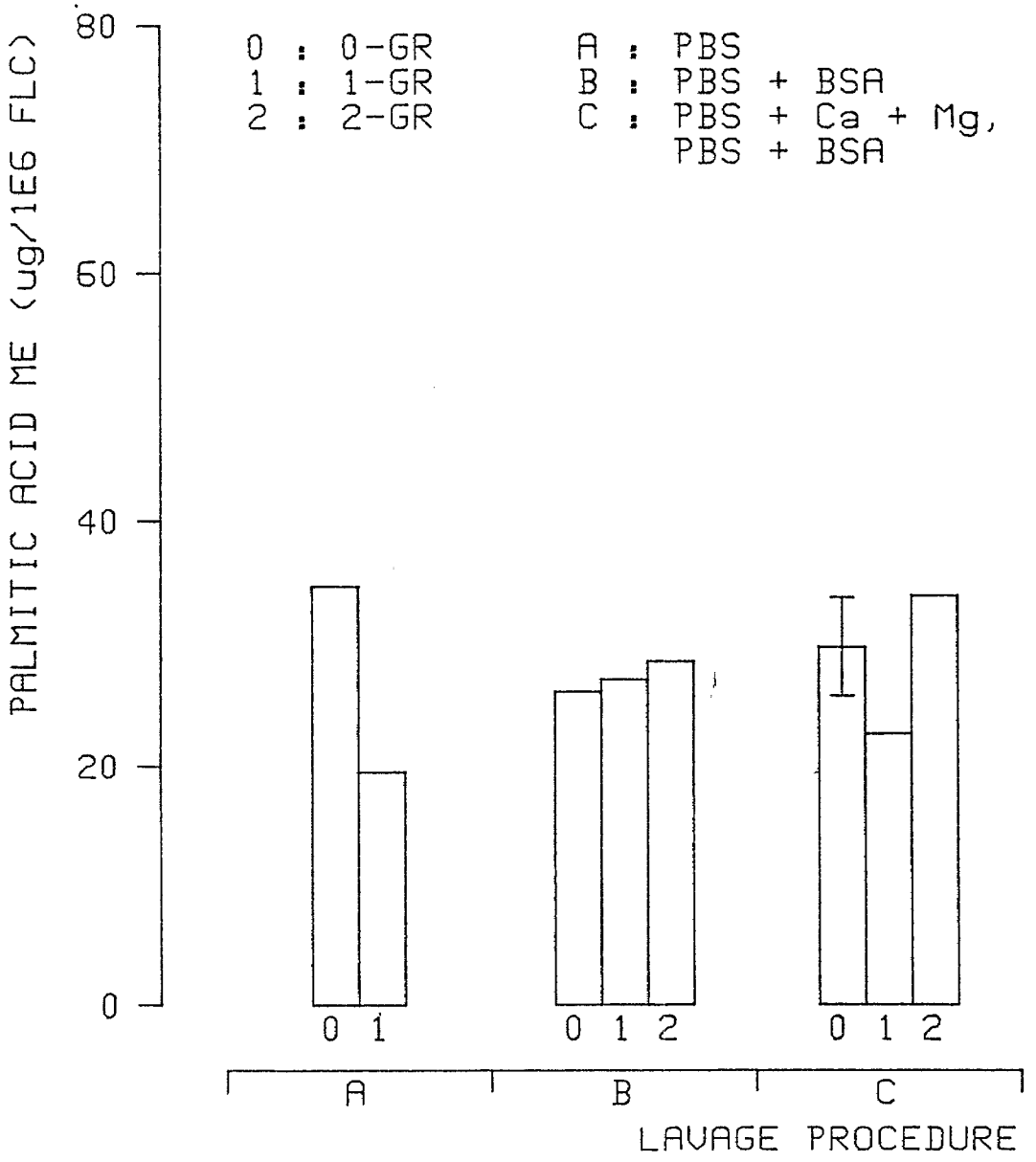
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 14

PALMITIC ACID METHYL ESTER (16 : 0) FROM FLC, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 13

A 0500/3057, HO3453, TH, U98 F155 US

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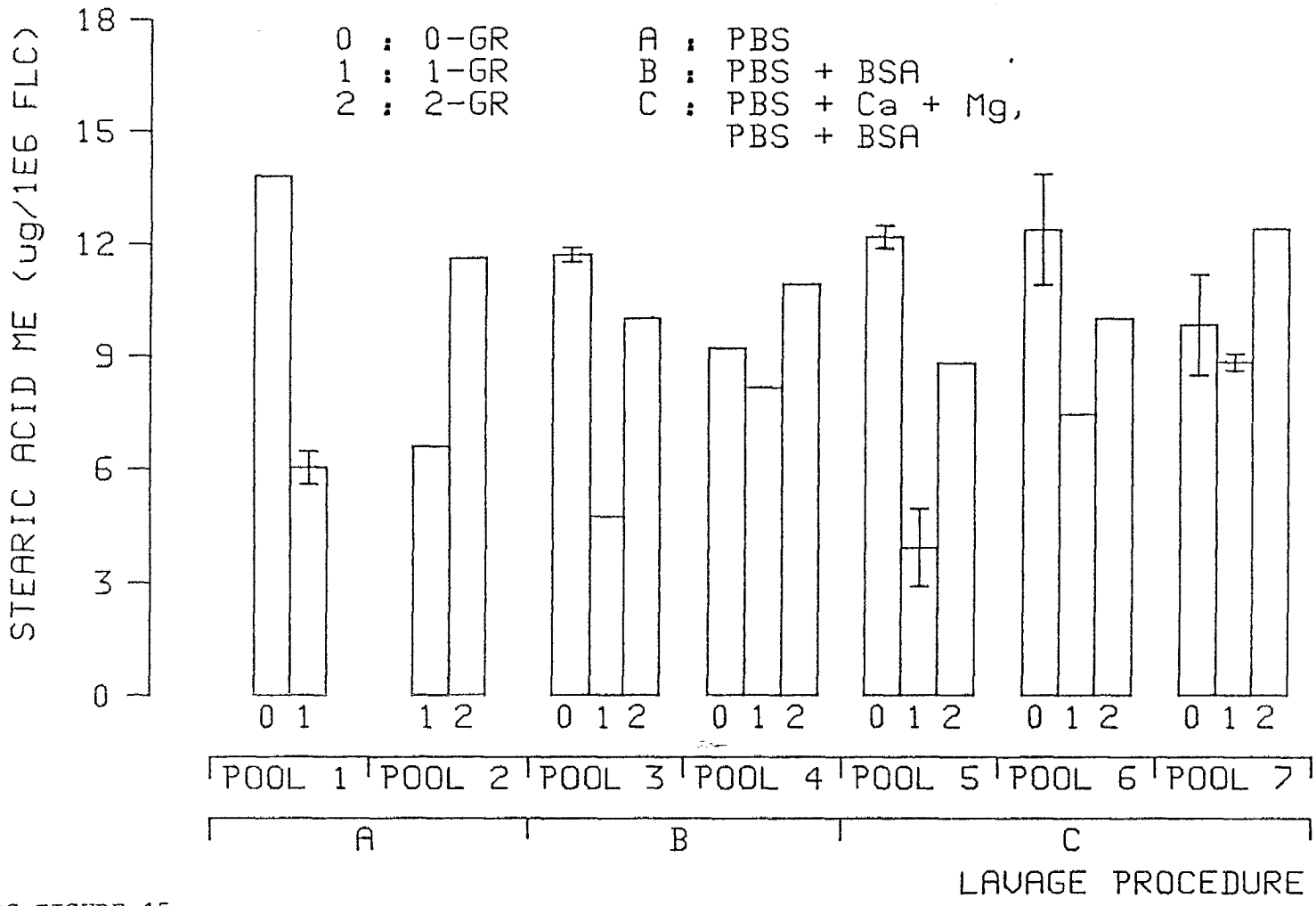

BC FIGURE 15

STEARIC ACID METHYL ESTER (18 : 0) FROM FLC, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 15

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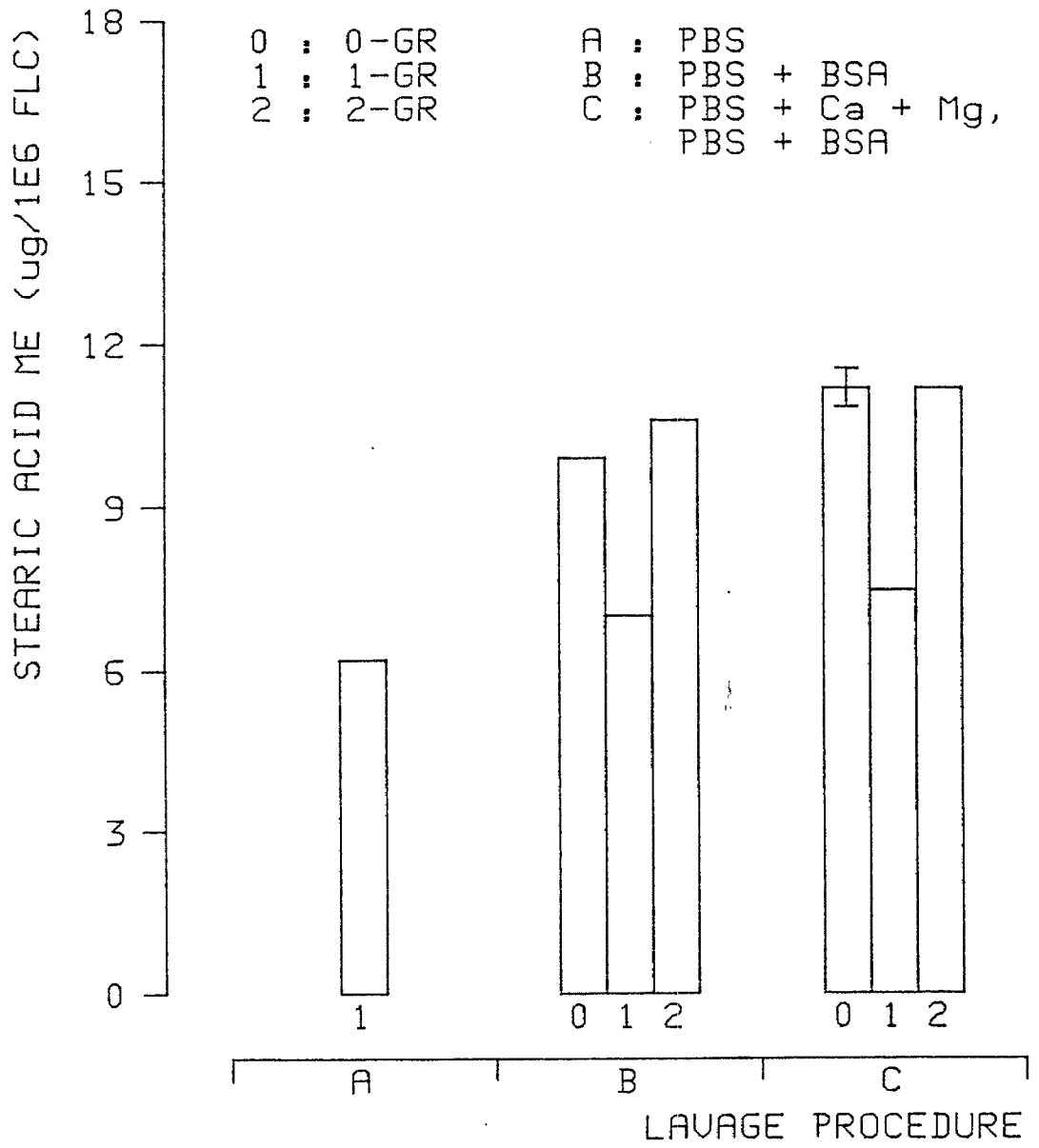

BC FIGURE 16

STEARIC ACID METHYL ESTER (18 : 0) FROM FLC, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 15

A 0500/3057, HO3451, TH, V123 F155 U96

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

BC FIGURE 17

RATIO OF PALMITIC ACID METHYL ESTER (16 : 0) VERSUS STEARIC ACID METHYL ESTER (18 : 0), INDIVIDUAL POOLS

Remarks: for details see BC TABLE 16

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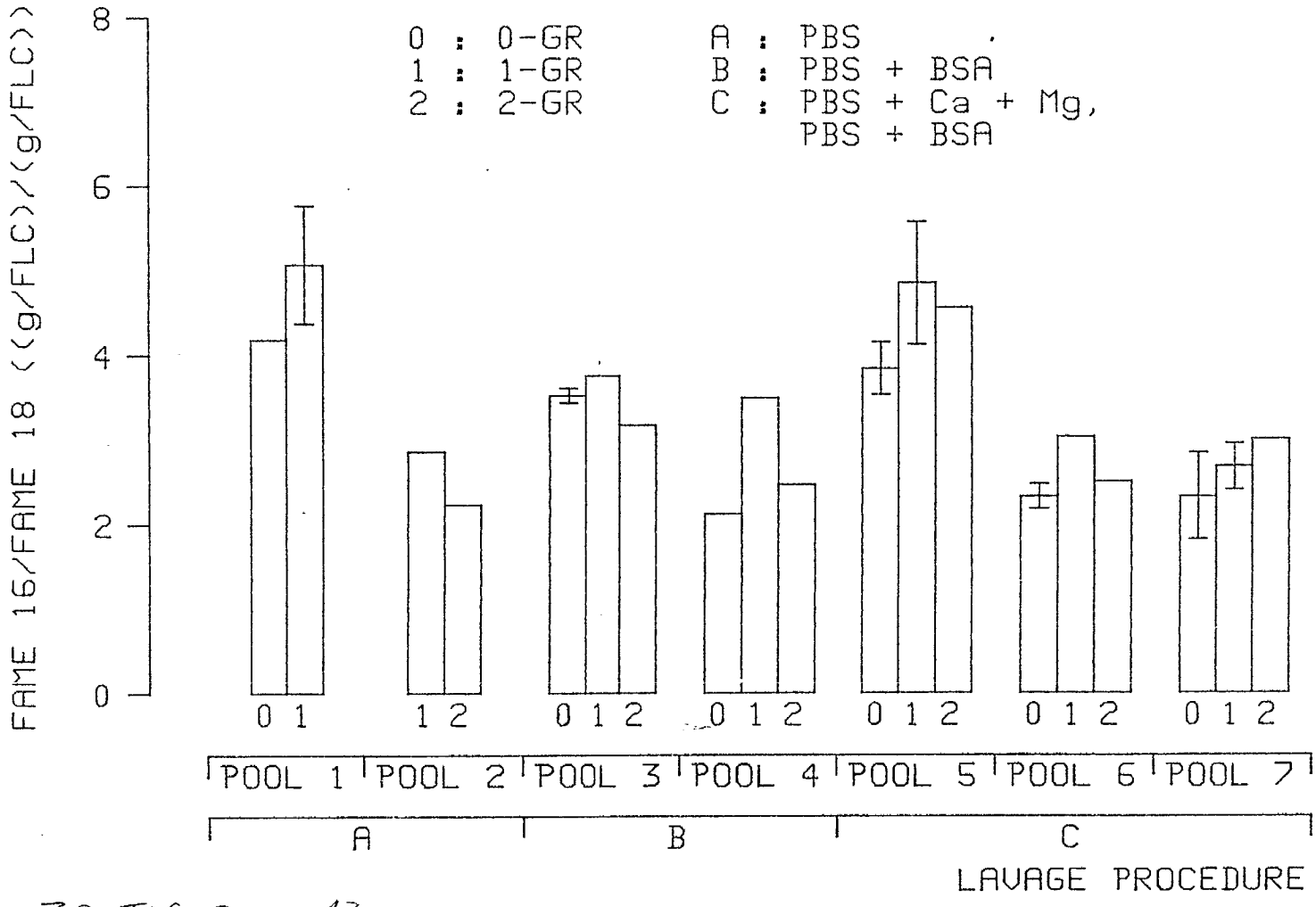
A 0500/3057, H03454, TH, V140 F155 V120



BC FIGURE 17

RATIO OF PALMITIC ACID METHYL ESTER (16:0) VERSUS STEARIC ACID METHYL ESTER (18:0), INDIVIDUAL POOLS

Remarks: for details see BC TABLE 16

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

BC FIGURE 18

RATIO OF PALMITIC ACID METHYL ESTER (16 : 0) VERSUS STEARIC ACID METHYL ESTER (18 : 0), WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 16

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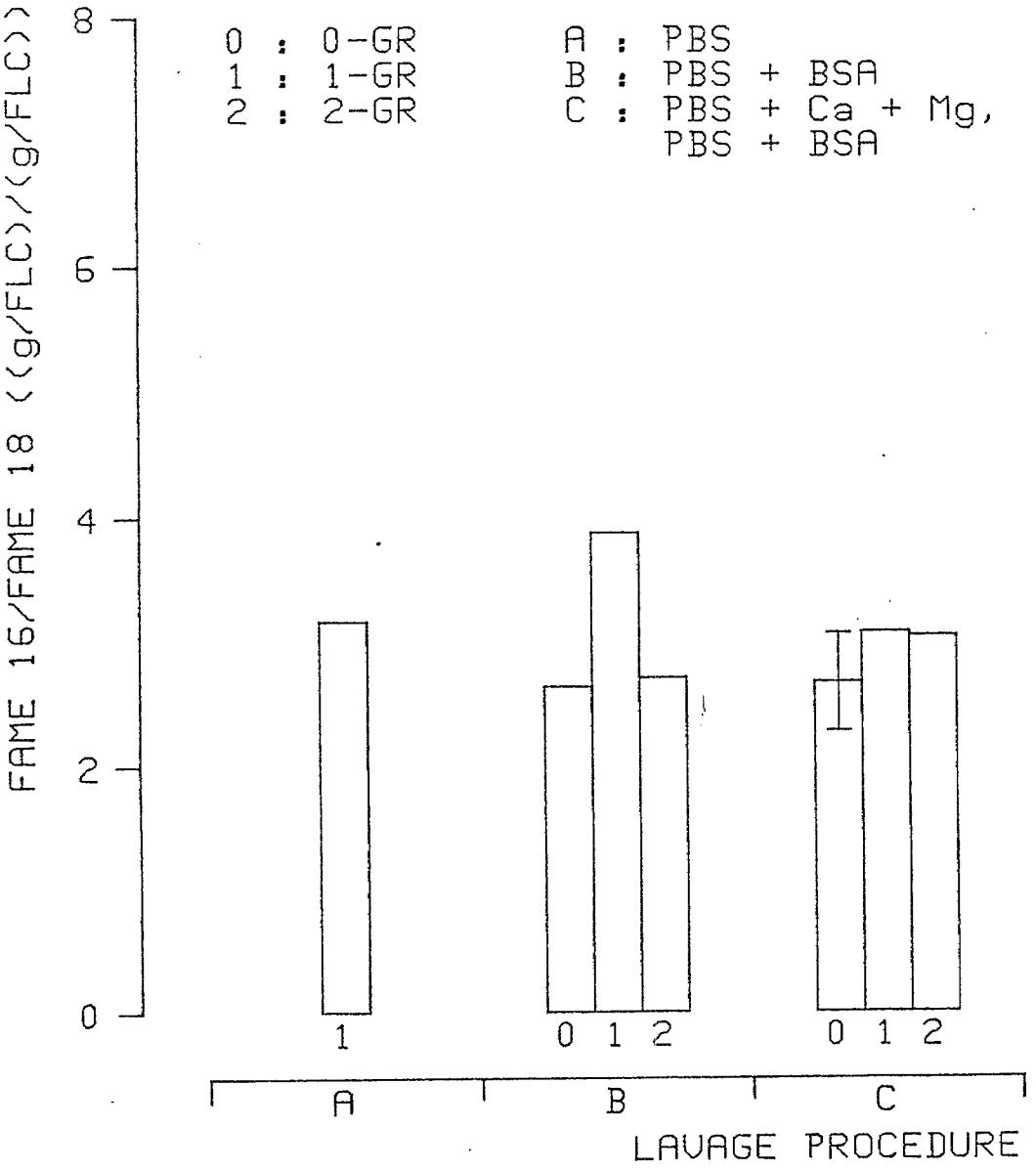
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 18

RATIO OF PALMITIC ACID METHYLESTER (16:0) VERSUS STEARIC ACID METHYL ESTER (18:0), WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 16

A 0500/30527, H03455, TH, V141 F155 ''21

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BC FIGURE 19

MEAN RATIO OF PALMITIC ACID METHYL ESTER (16 : 0) VERSUS STEARIC ACID METHYL ESTER (18 : 0), EARLY AND LATE POOLS

Remarks: for details see BC TABLE 17

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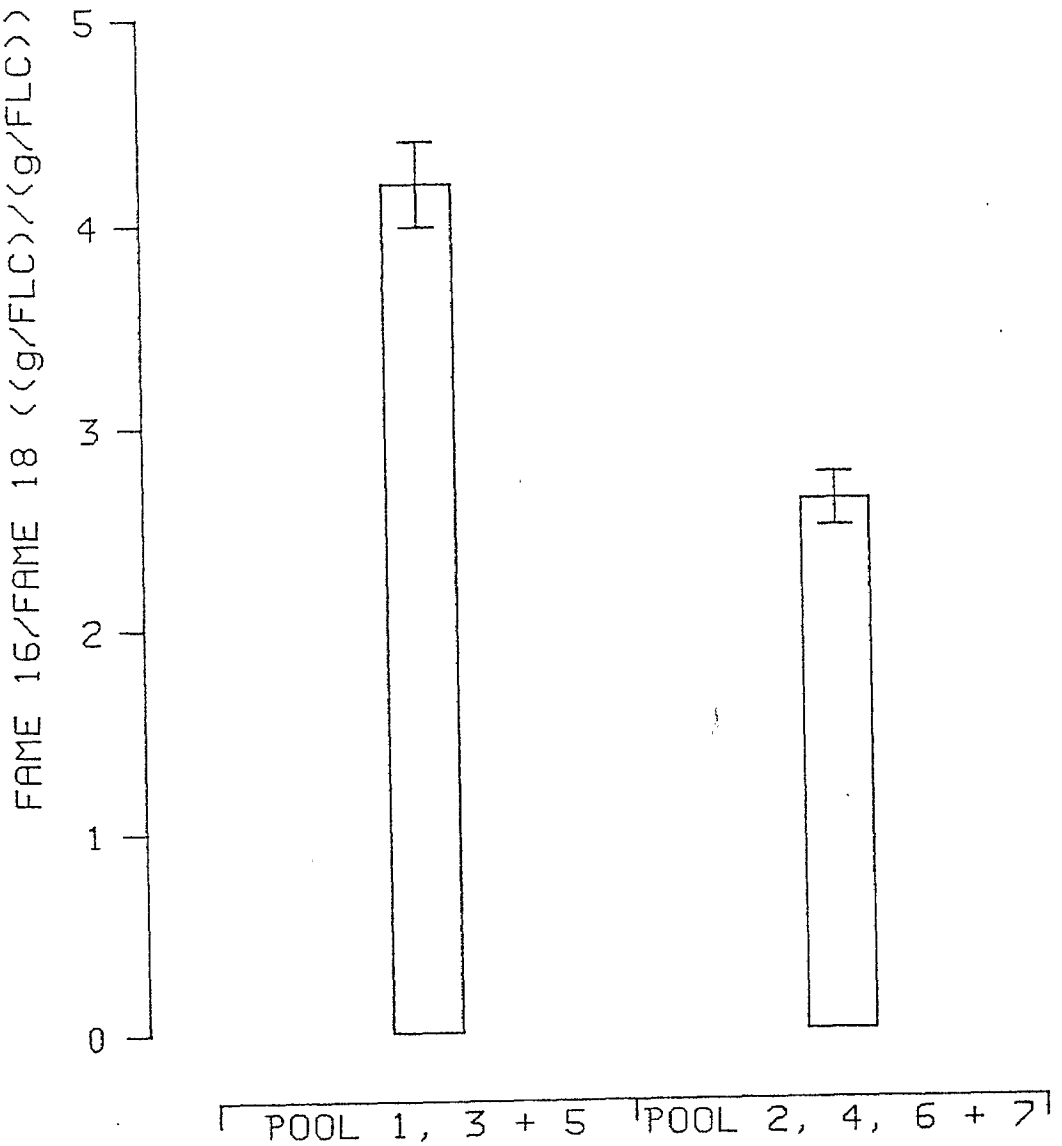

BC FIGURE 19

MEAN RATIO OF PALMITIC ACID METHYL ESTER (16:0) VERSUS STEARIC ACID METHYLESTER (18:0), EARLY AND LATE POOLS

Remarks: for details see BC TABLE 17

A 0500/30057, H03471, TH, V139 F155 ' 22

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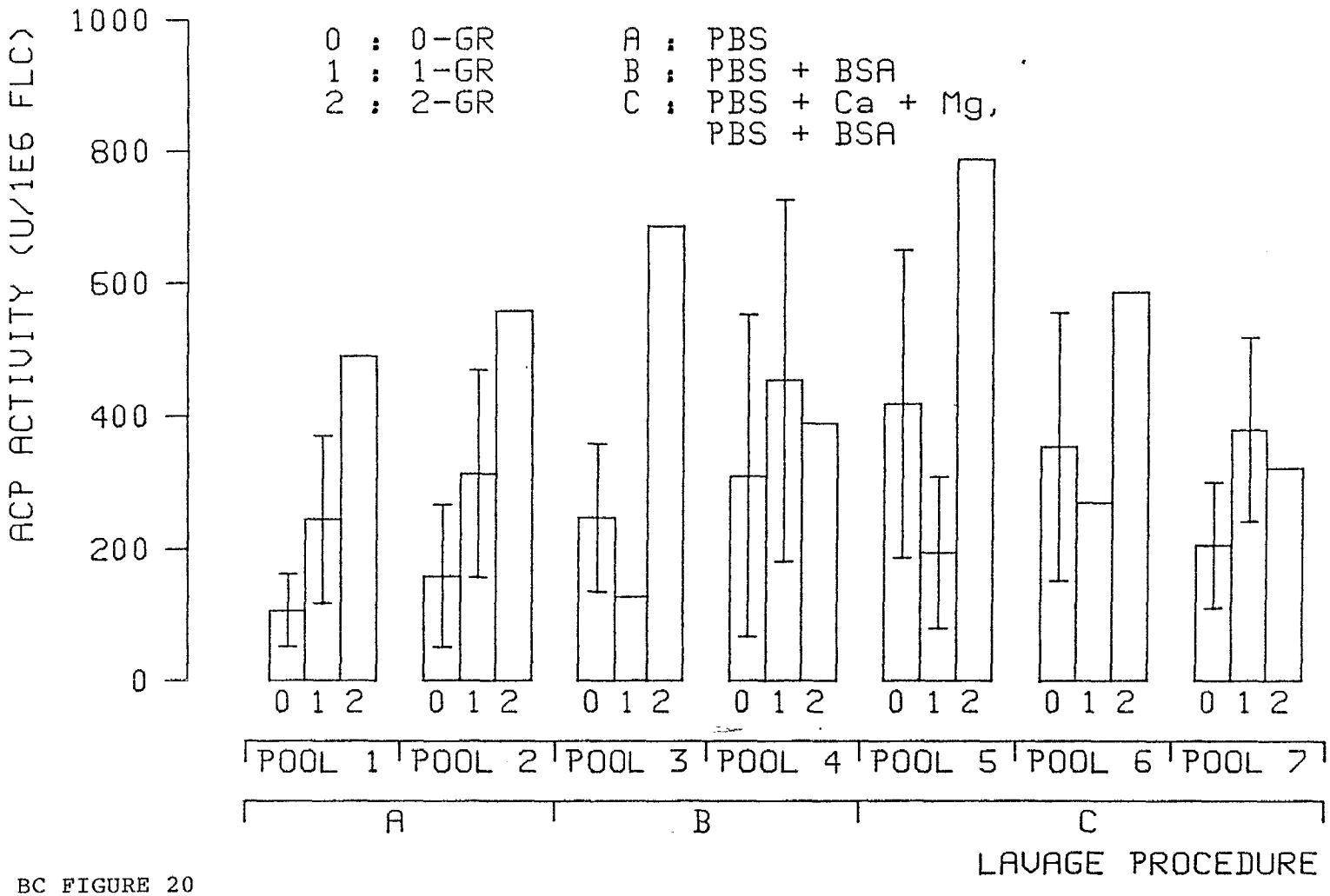
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 20

ACID PHOSPHATASE ACTIVITY, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 21

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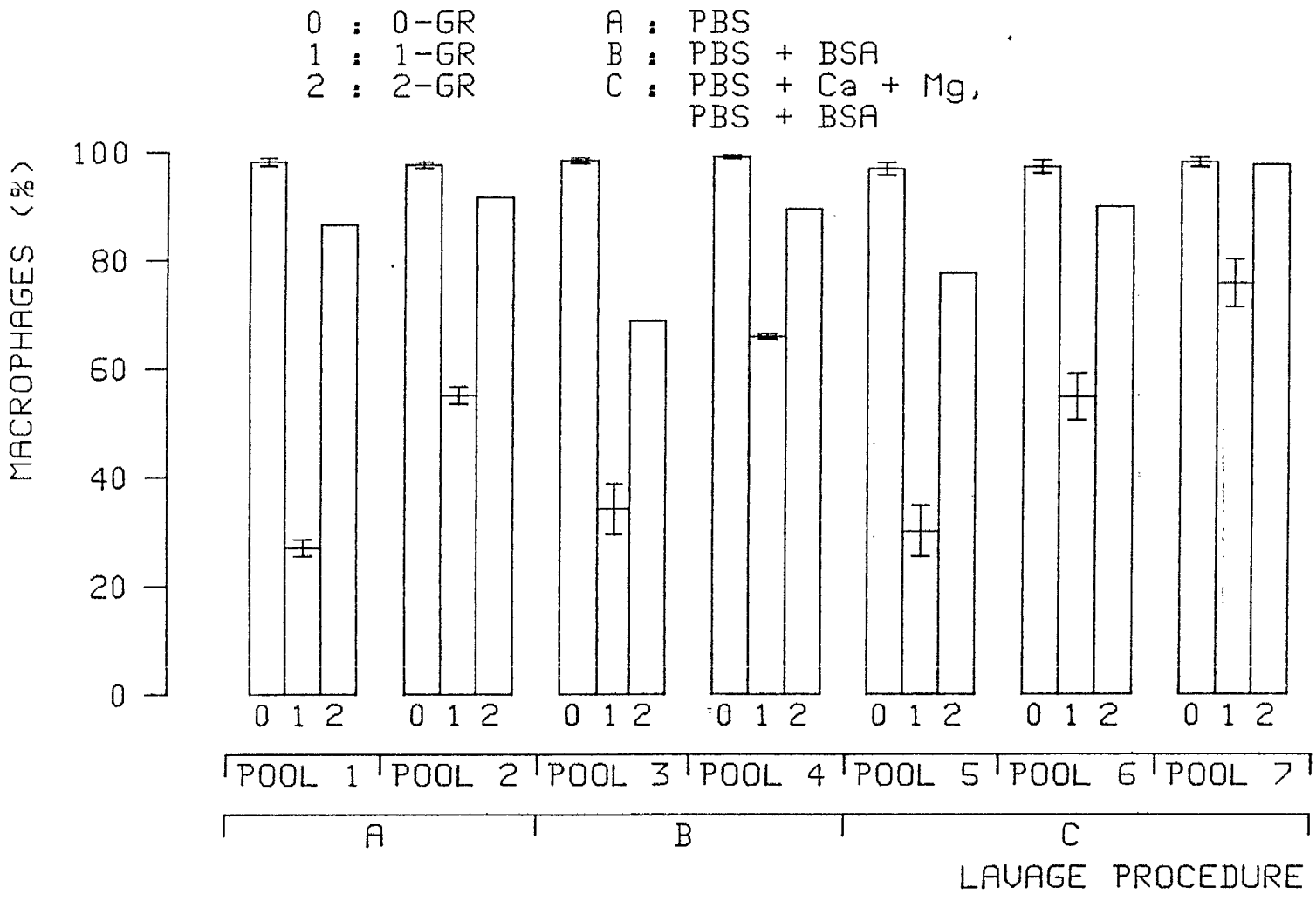
20290274 92

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

2284



BC FIGURE 21

RELATIVE NUMBER OF MACROPHAGES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

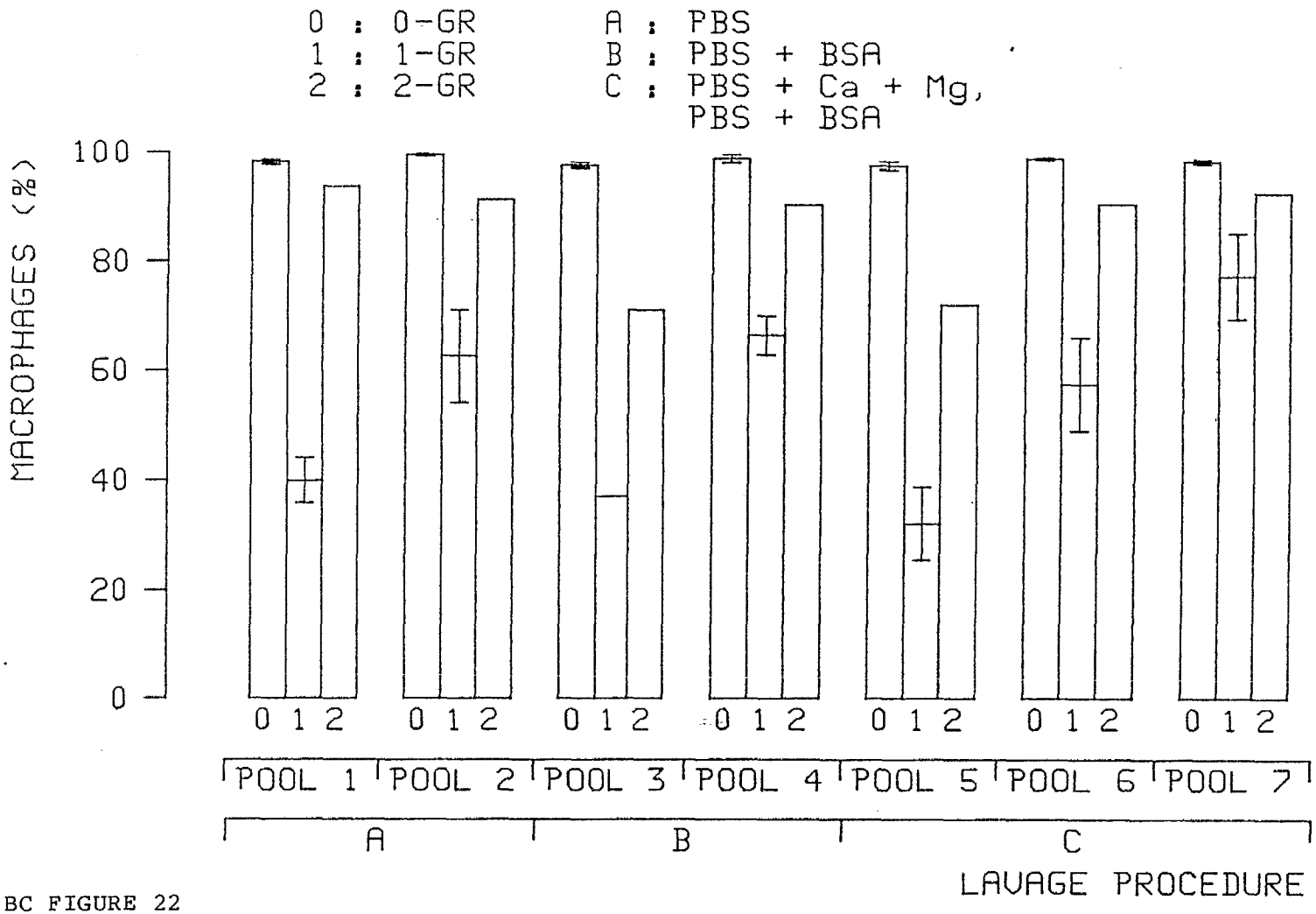
Remarks: for details see BC TABLE 24

2029027493

INBIFO Institut für biologische Forschung · Köln

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

2284



BC FIGURE 22

RELATIVE NUMBER OF MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 26

20290277494

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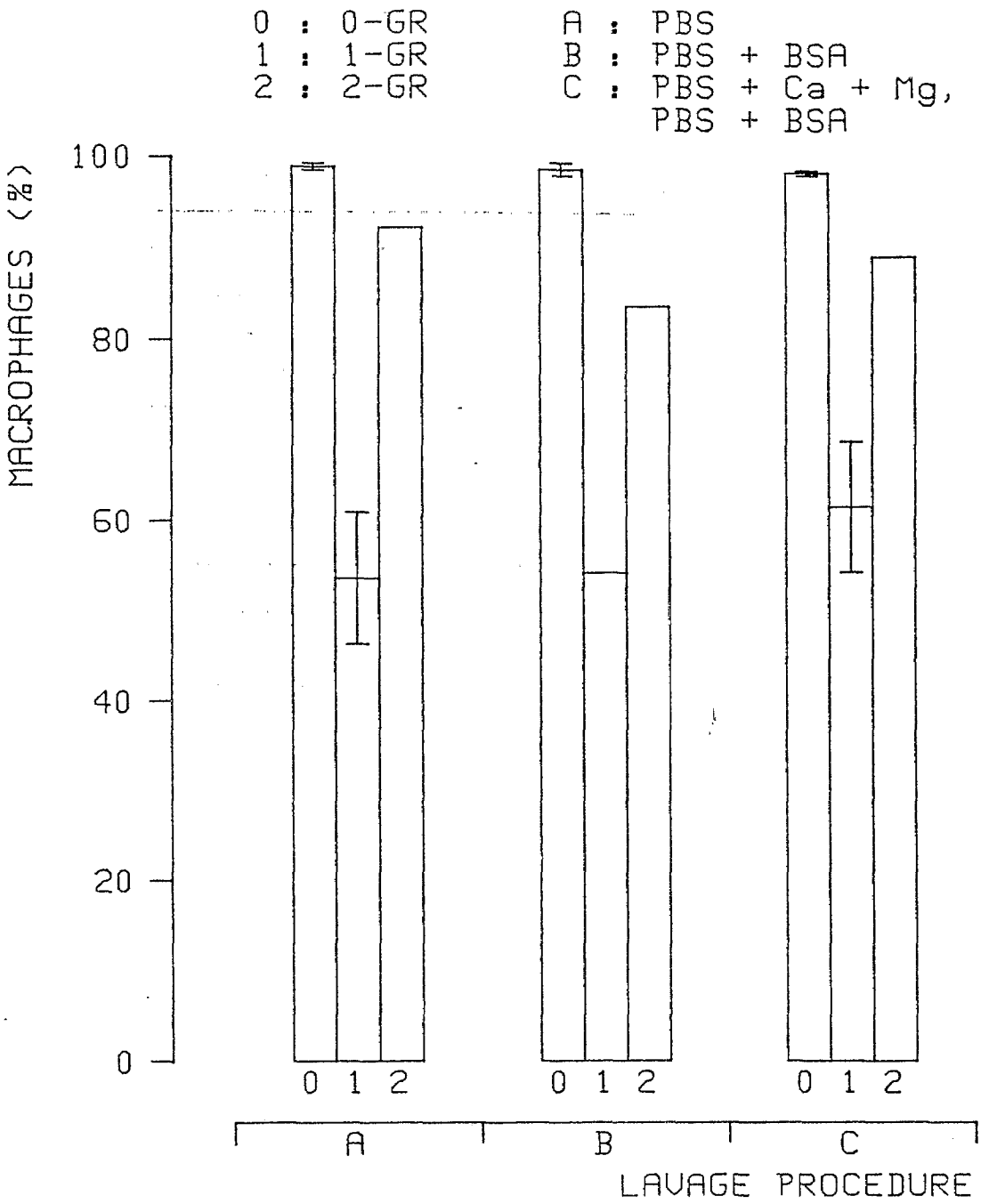
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B21 WS M HO3215



BC FIGURE 23

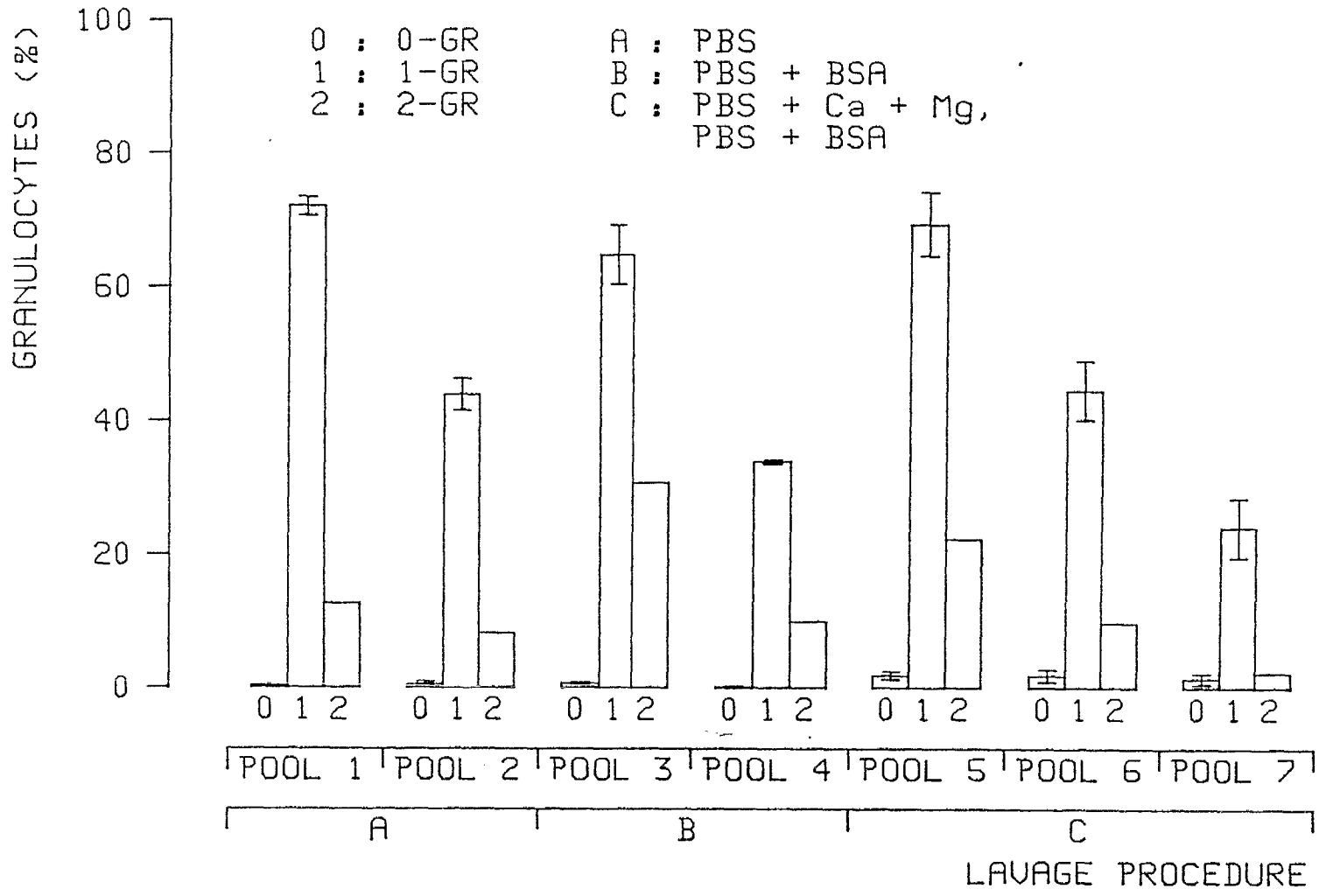
RELATIVE NUMBER OF MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 26

A 0500/3057, HO3215, TH, V85 F155 V82, R

2029027495

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 24

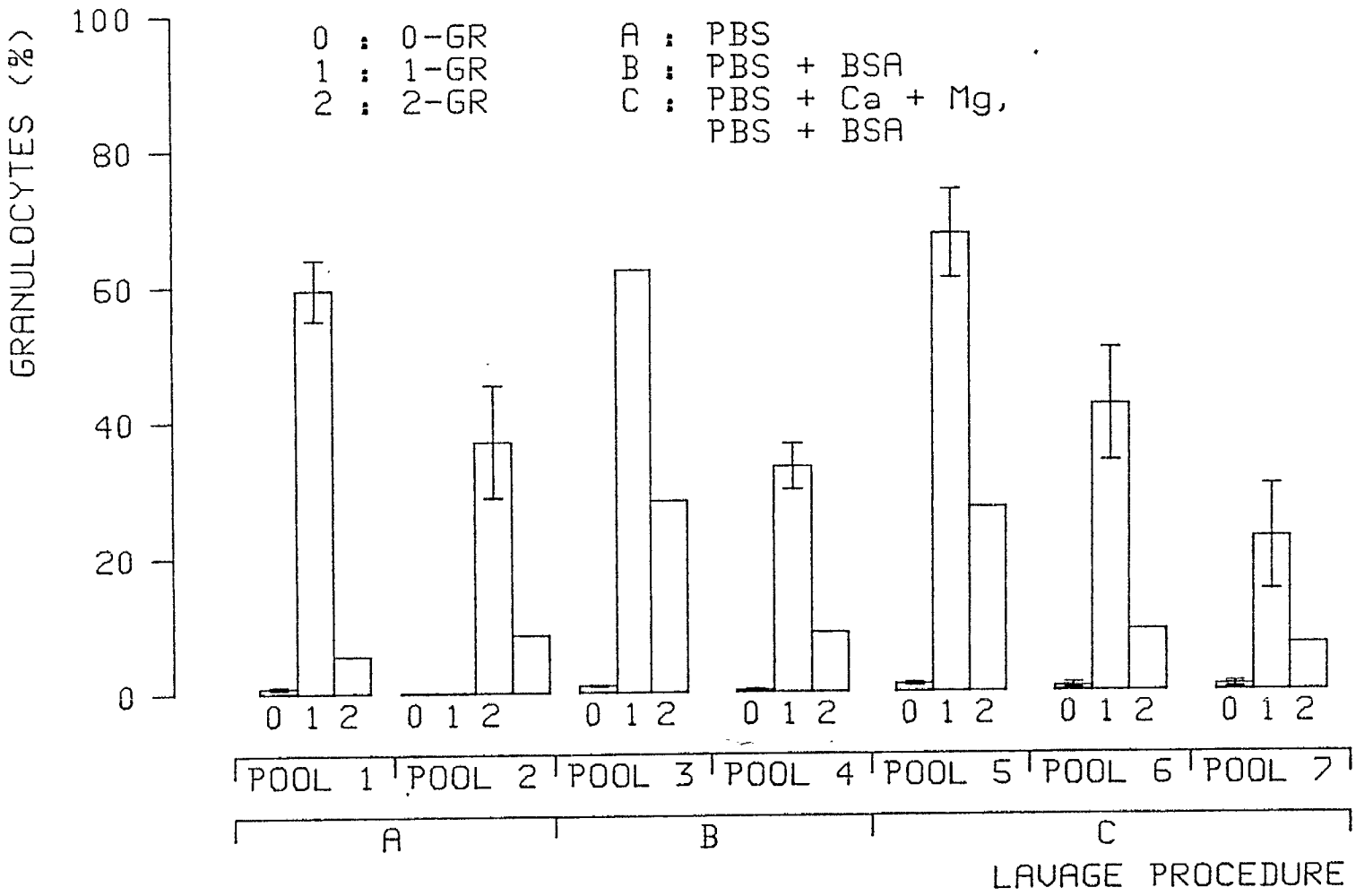
RELATIVE NUMBER OF GRANULOCYTES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 28

20290277496

INBIFO Institut für biologische Forschung · Köln

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 25

RELATIVE NUMBER OF GRANULOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 30

2029027497

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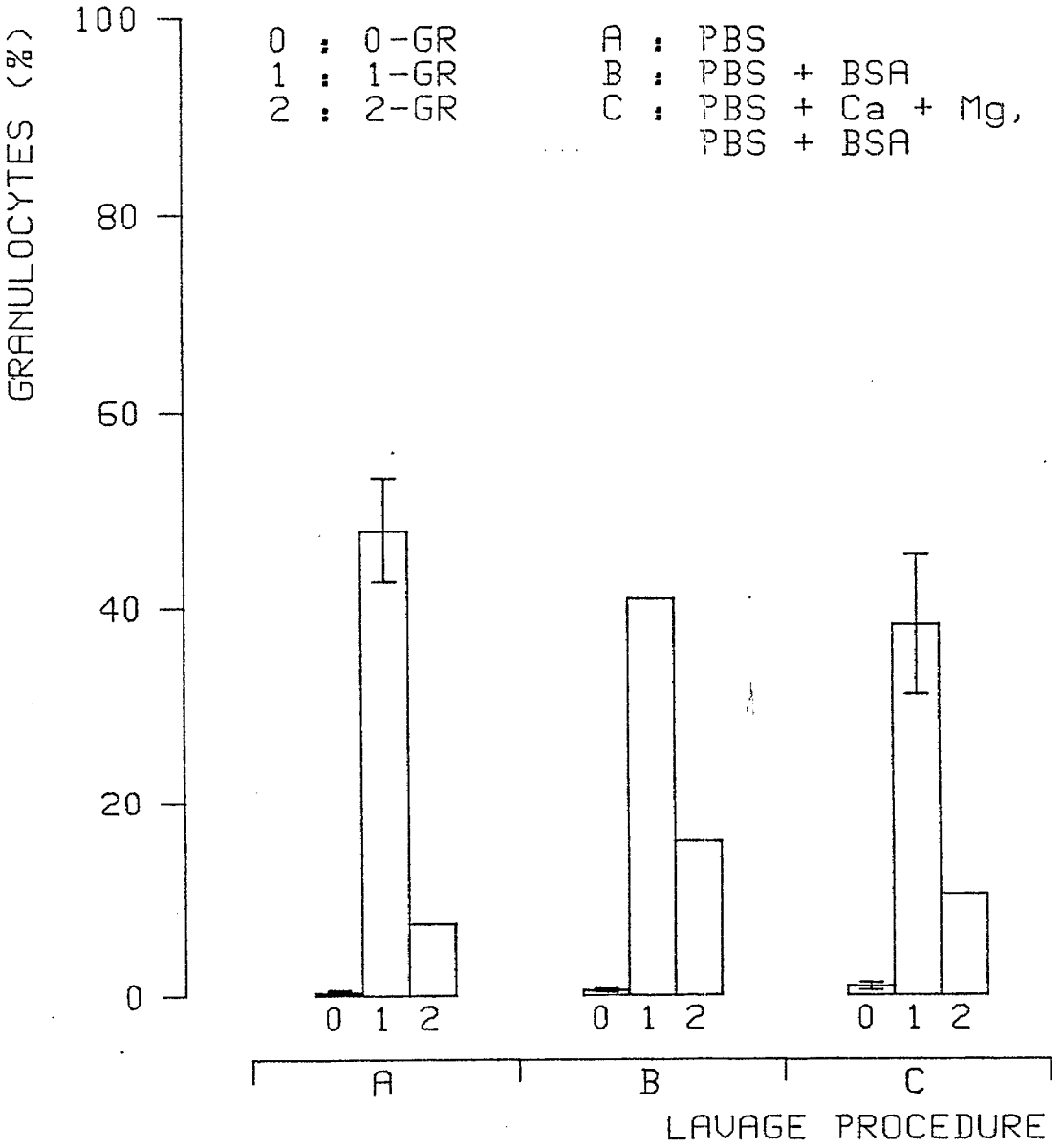
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

Werte!



BC FIGURE 26

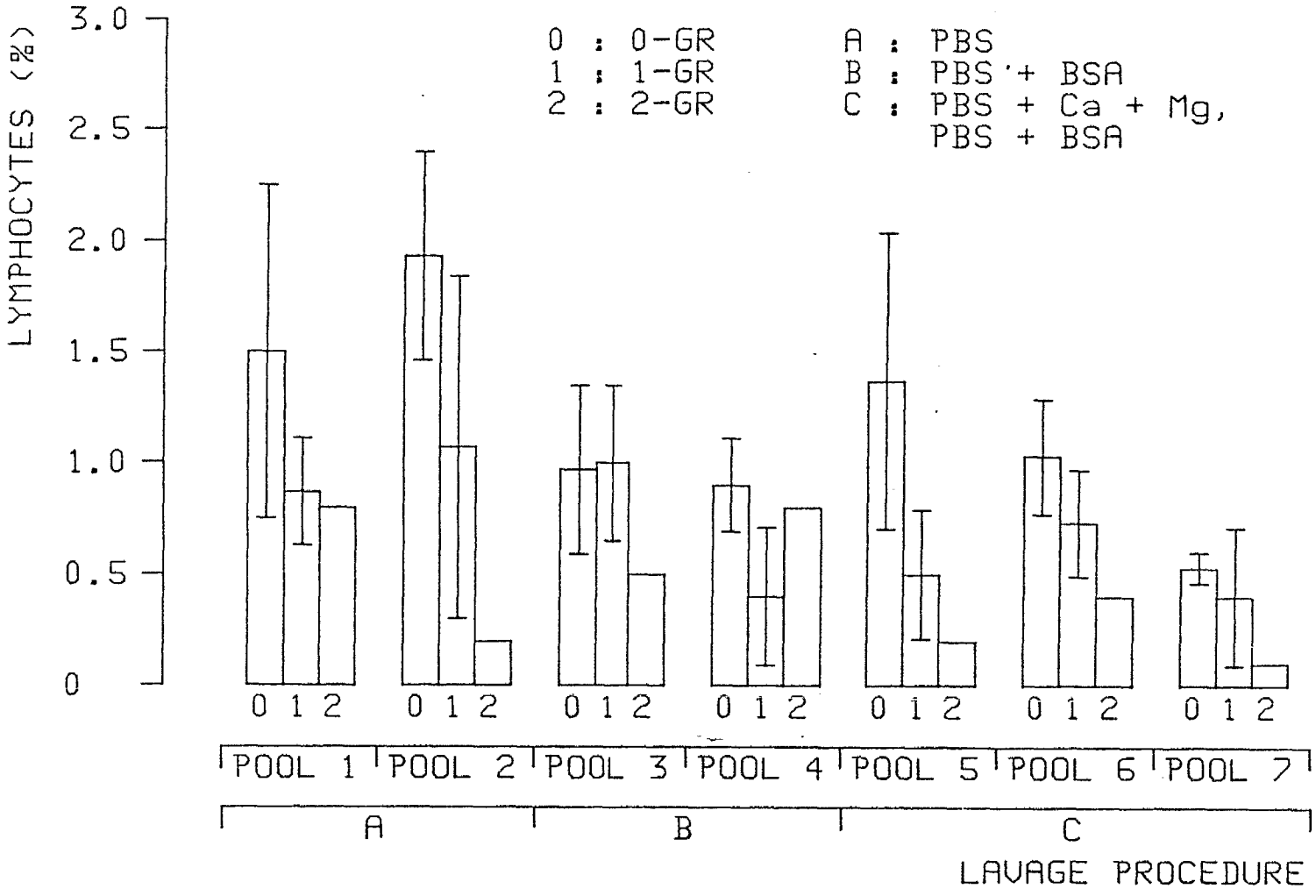
RELATIVE NUMBER OF GRANULOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 30

A 0500/3057, HO3219, TH, V76 F155 V7[illegible] R

2029027498

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 27

RELATIVE NUMBER OF LYMPHOCYTES, LAVAGE MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 32

20290274 99

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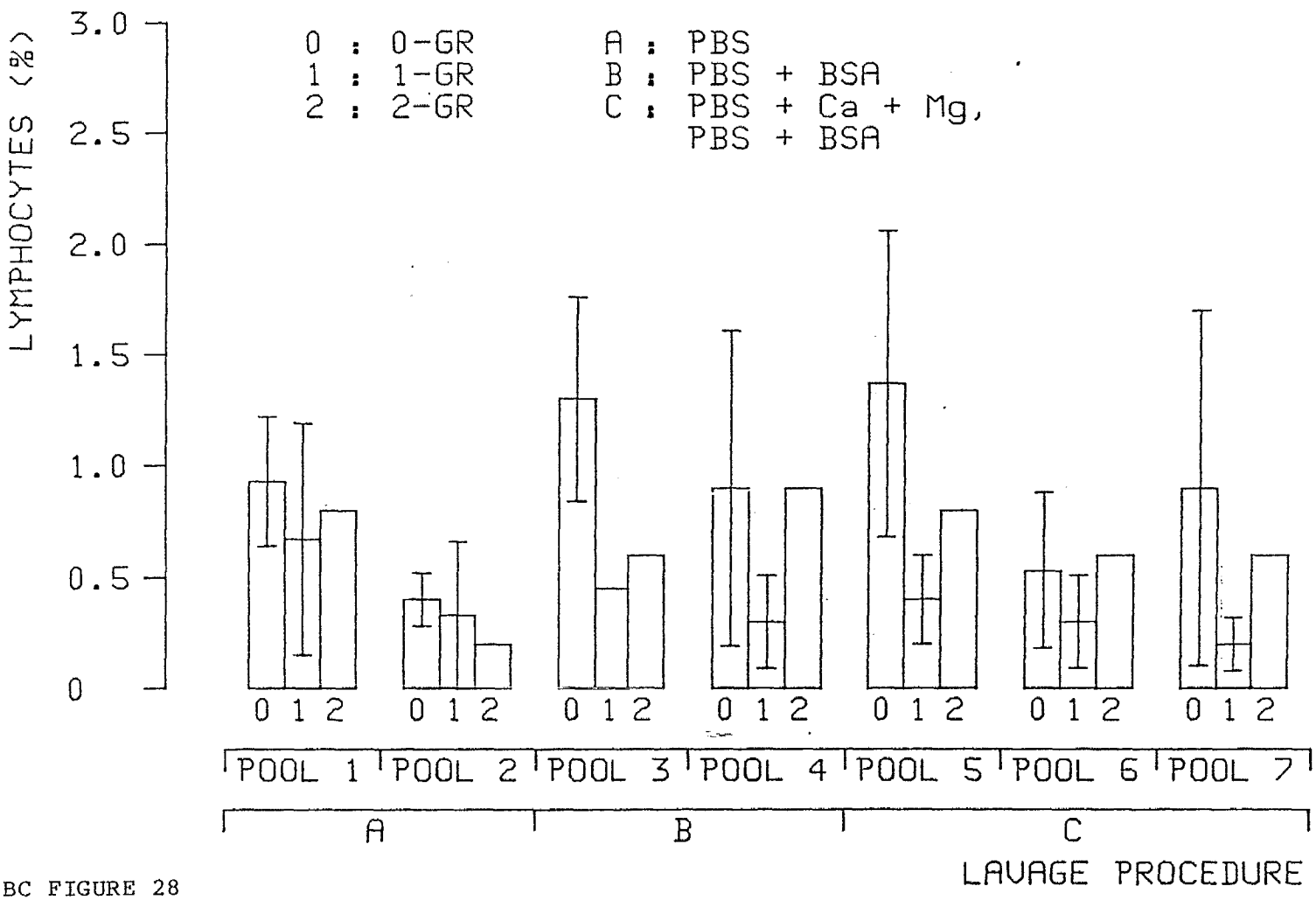
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 28

RELATIVE NUMBER OF LYMPHOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 34

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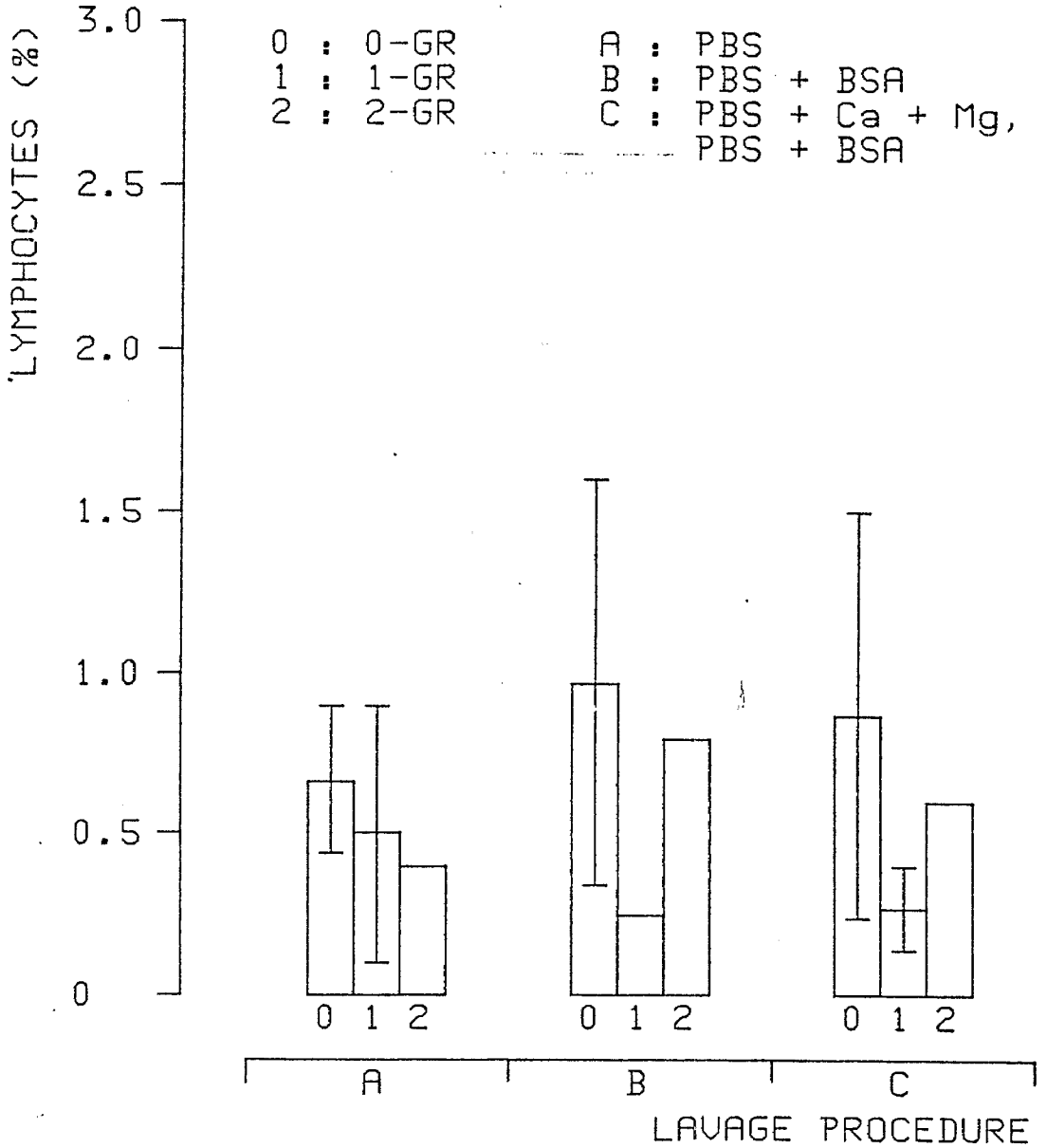
20290275OO

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 29

RELATIVE NUMBER OF LYMPHOCYTES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 34

A 0500/3057, HO3211, TH, U79 F155 U7[illegible] R

2029027501

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B22 WS M HO3456

BC FIGURE 30

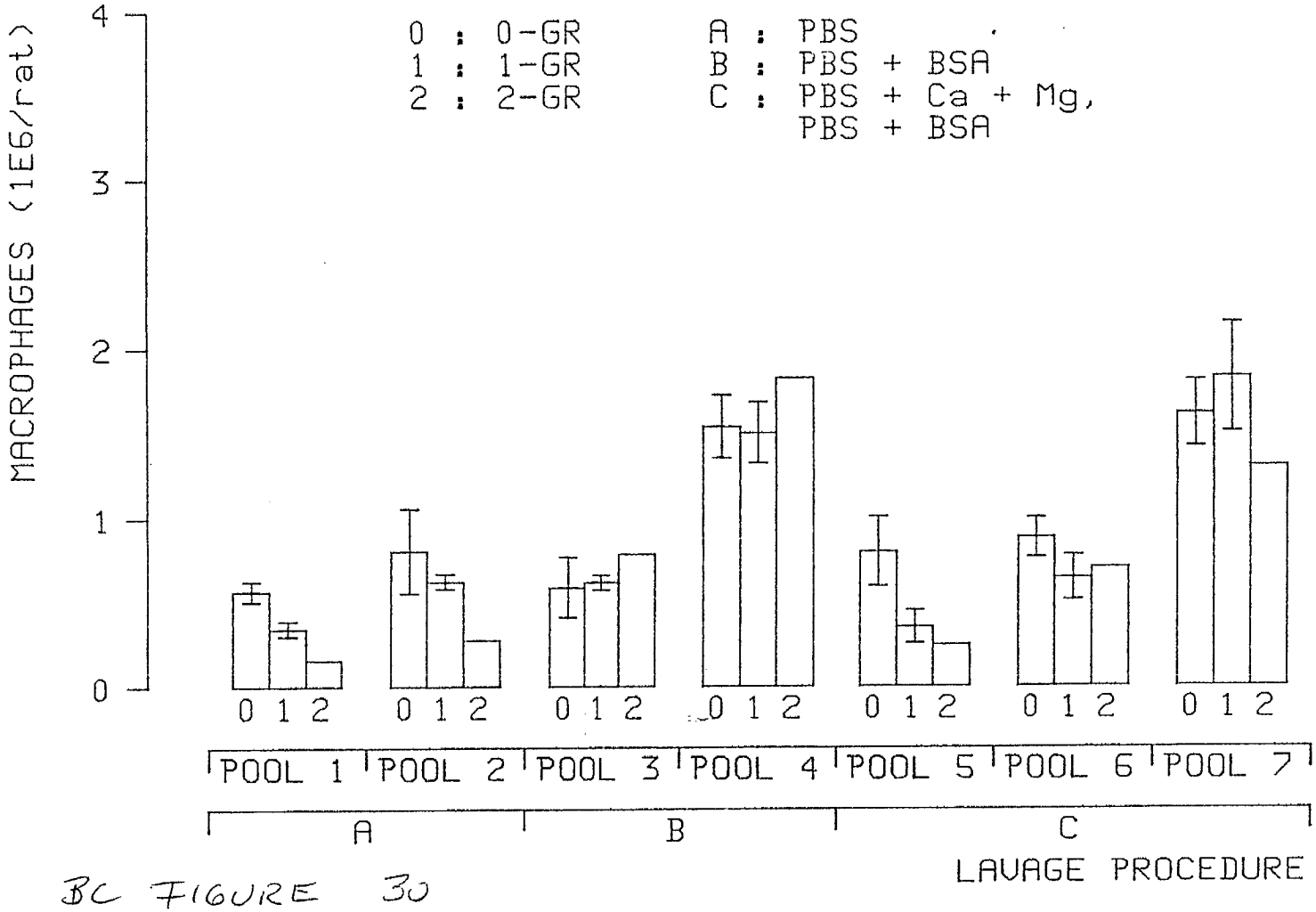
NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 36

20290275O2

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03456, TH, V136 F155 V101 R



BC FIGURE 30

NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 36

20290275O3

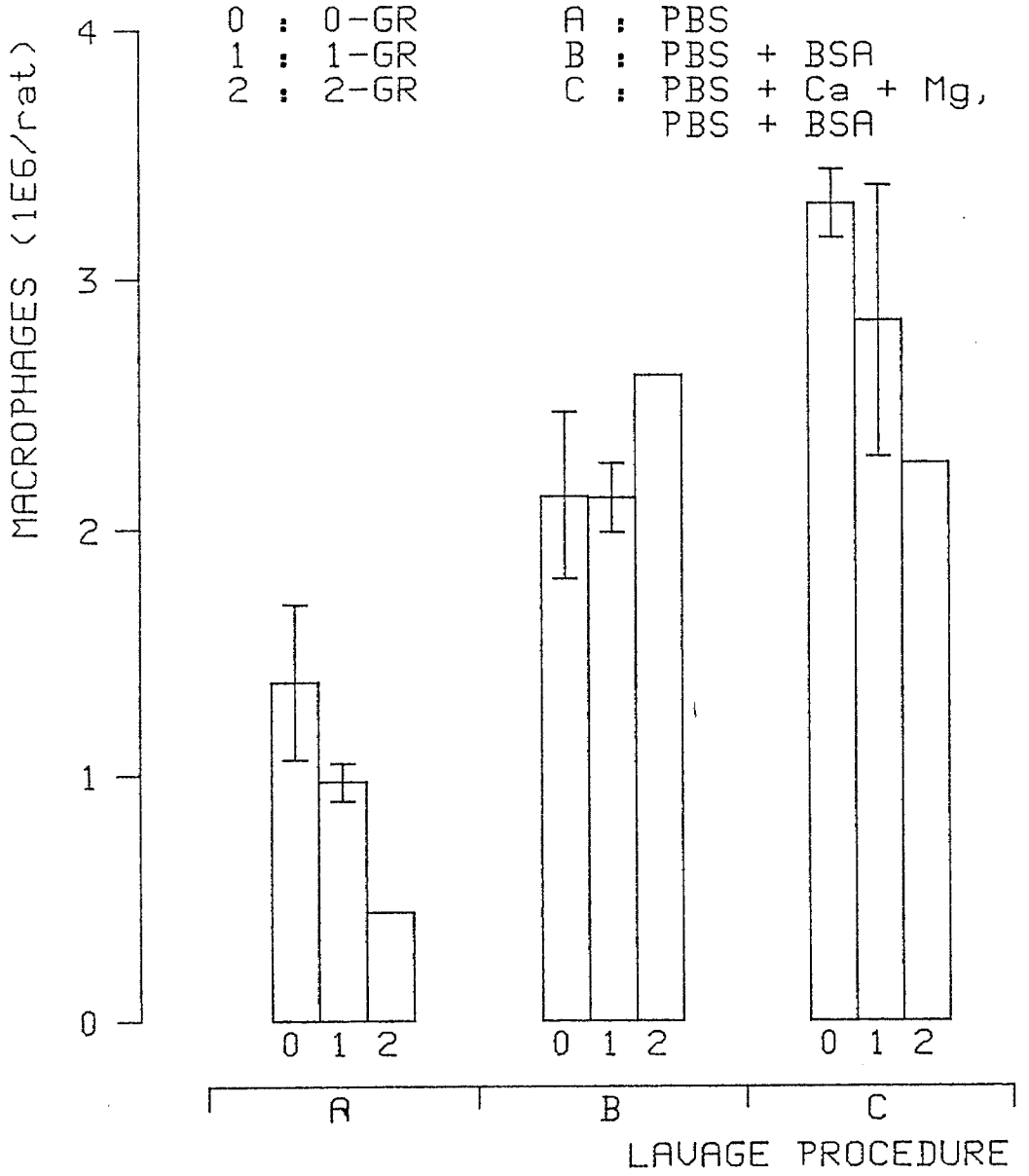
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

BC FIGURE 31

NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER, SUM OF POOLS
Remarks: for details see BC TABLE 36

2029027504

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 31

NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER, SUM OF POOLS

Remarks : for details see BC TABLE 36

A 0500/3057, H03457, TH, V130 F155 ! J2 R

2029027505

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

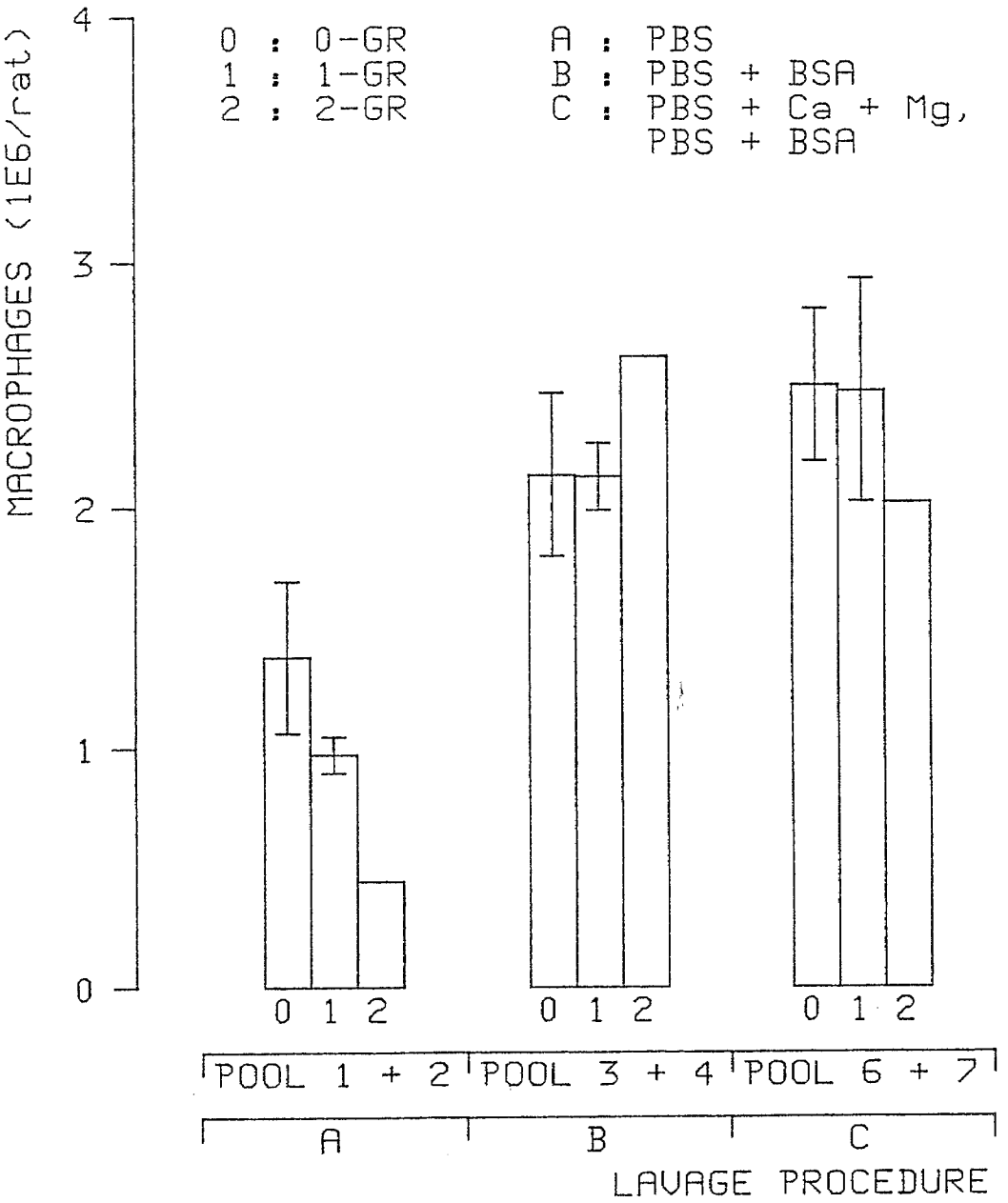
BC FIGURE 31 (continued)

NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER, SUM OF POOLS

Remarks: for details see BC TABLE 36

2029027506

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 31 continued

NUMBER OF MACROPHAGES PER RAT, RESUSPENSION MEDIUM, HEMOCYTOMETER, SUM OF POOLS

Remarks: for details see BC TABLE 36

A 0500/3057, H03458, TH, V131 F155 ' 93 R

20290275O7

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B23 WS M HO3231

BC FIGURE 32

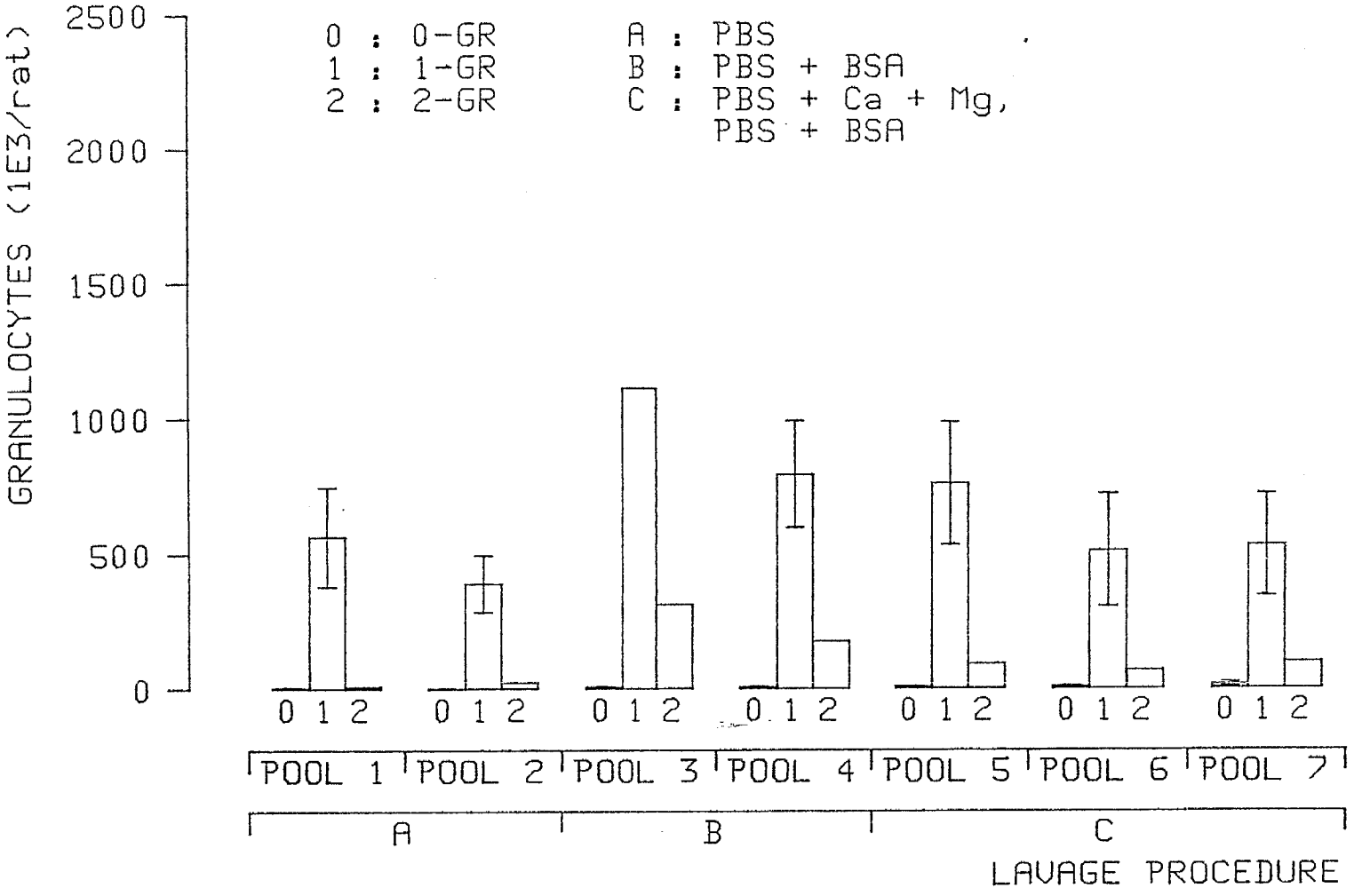
NUMBER OF GRANULOCYTES PER RAT, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 38

2029027508

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03231, TH, V132 F155 V73 R



BC FIGURE 32

NUMBER OF GRANULOCYTES PER RAT, RESUSPENSION MEDIUM, CYTO CENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE =>

20290275O9

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

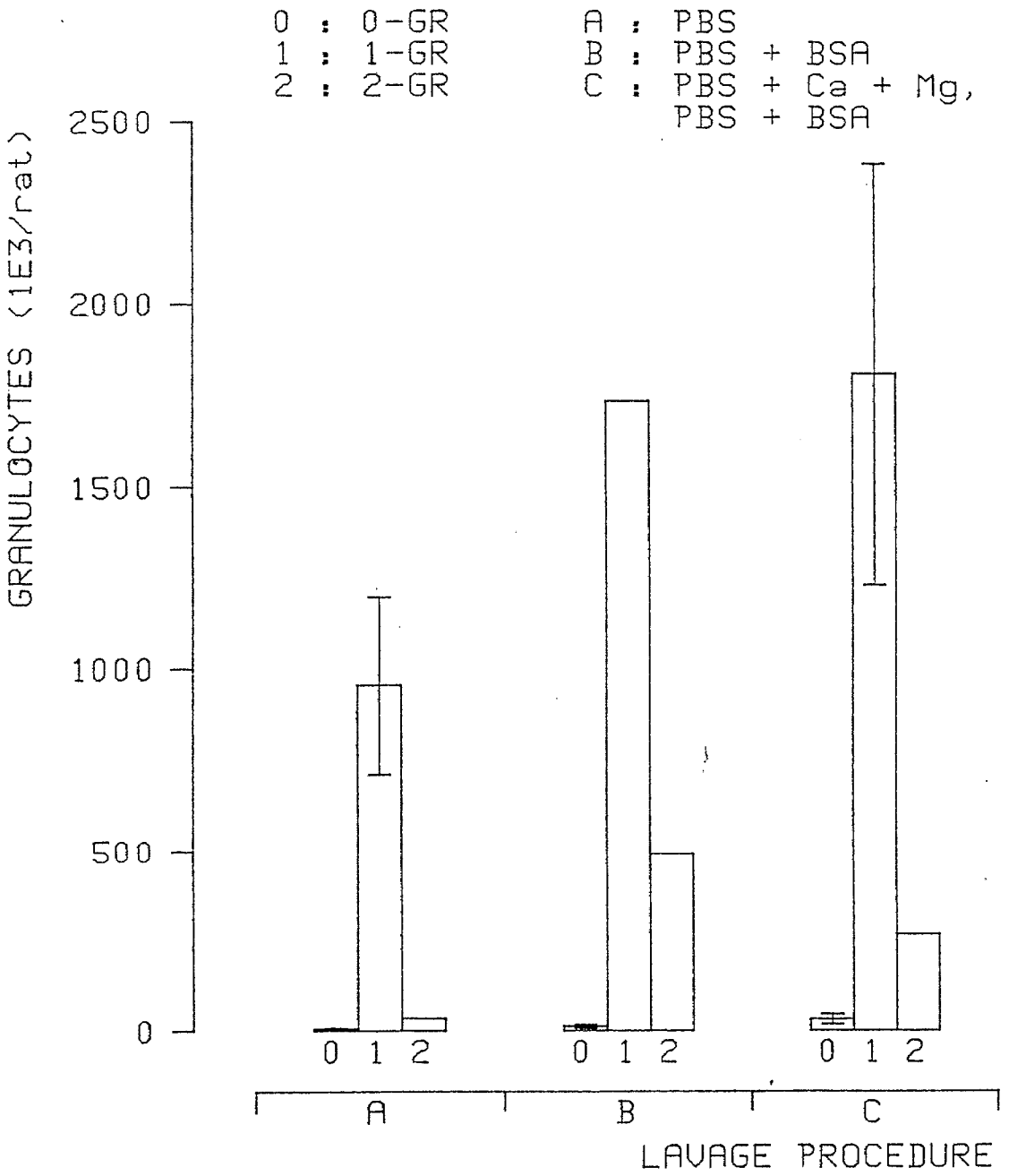
BC FIGURE 33

NUMBER OF GRANULOCYTES PER RAT, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, SUM OF POOLS

Remarks: for details see BC TABLE 38

2029027510

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 33

NUMBER OF GRANULOCYTES PER RAT, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, SUM OF POOLS

Remarks: for details see BC TABLE 38

A 0500/3057, H03228, TH, V129 F155 2 R

2029027511

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

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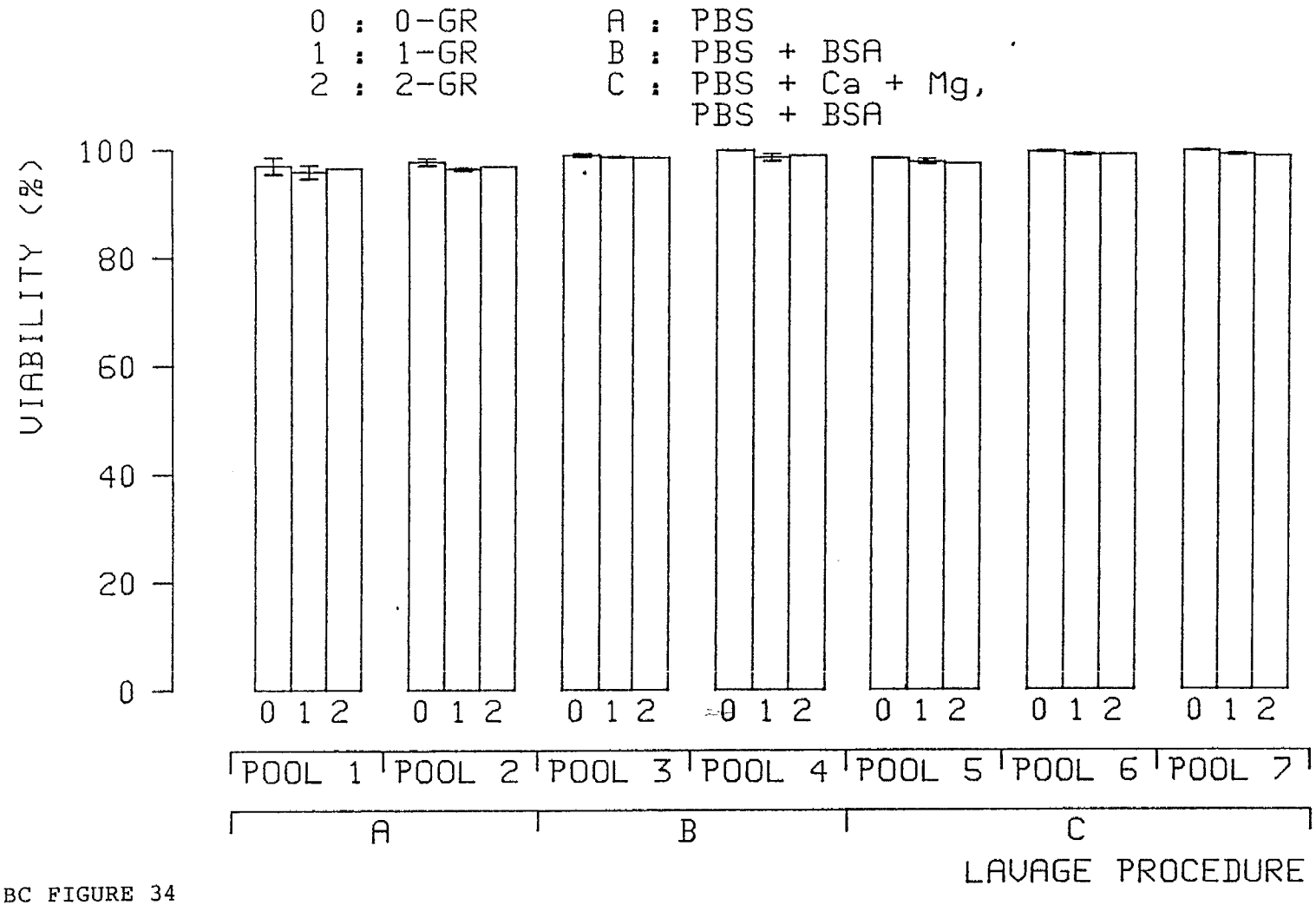


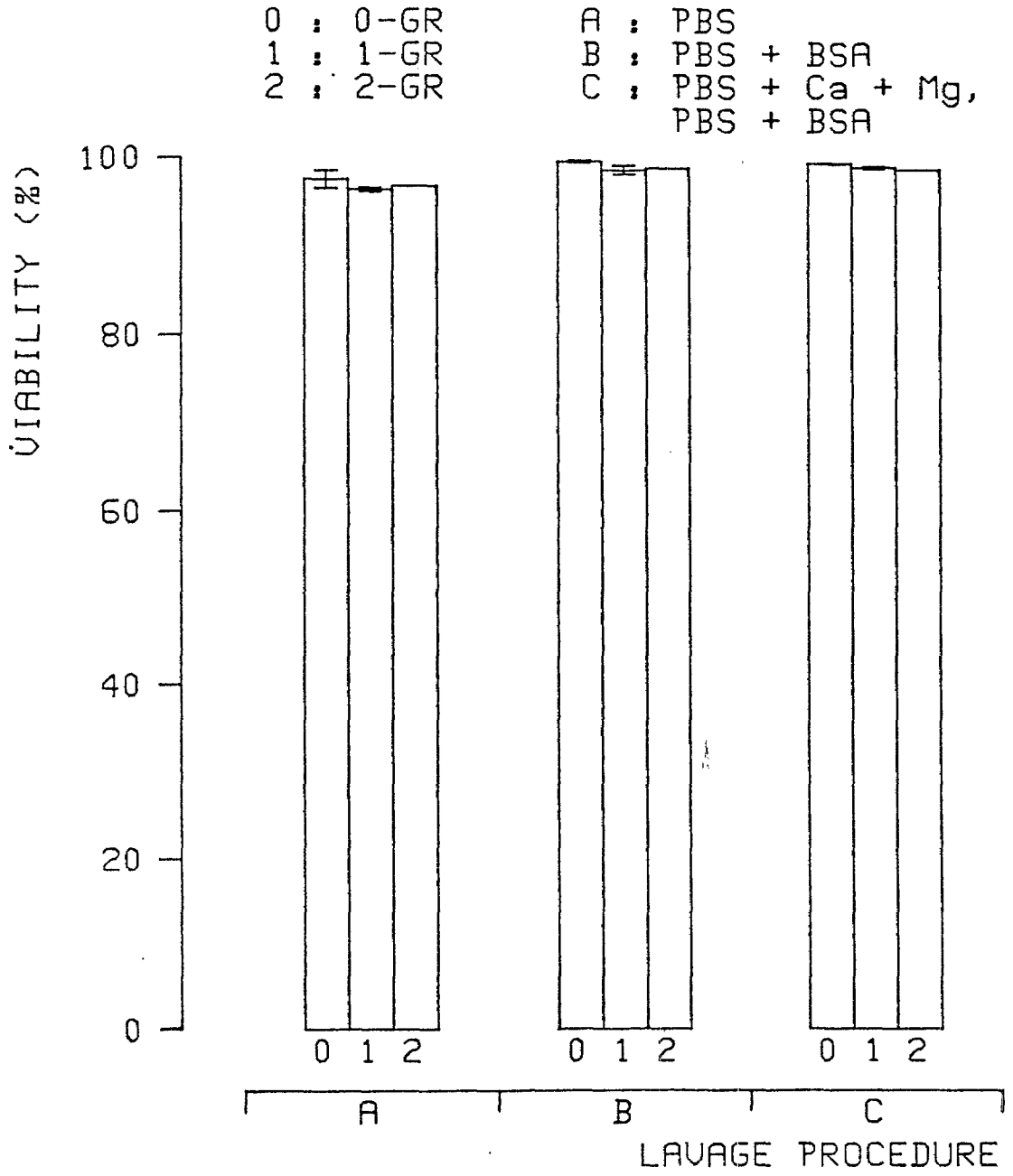
BC FIGURE 34

VIABILITY OF MACROPHAGES, RESUSPENSION MEDIUM, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 40

20290275I2

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 35

VIABILITY OF MACROPHAGES, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 40

A 0500/3057, HO3187, TH, V82 F155 V , R

2029027513

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

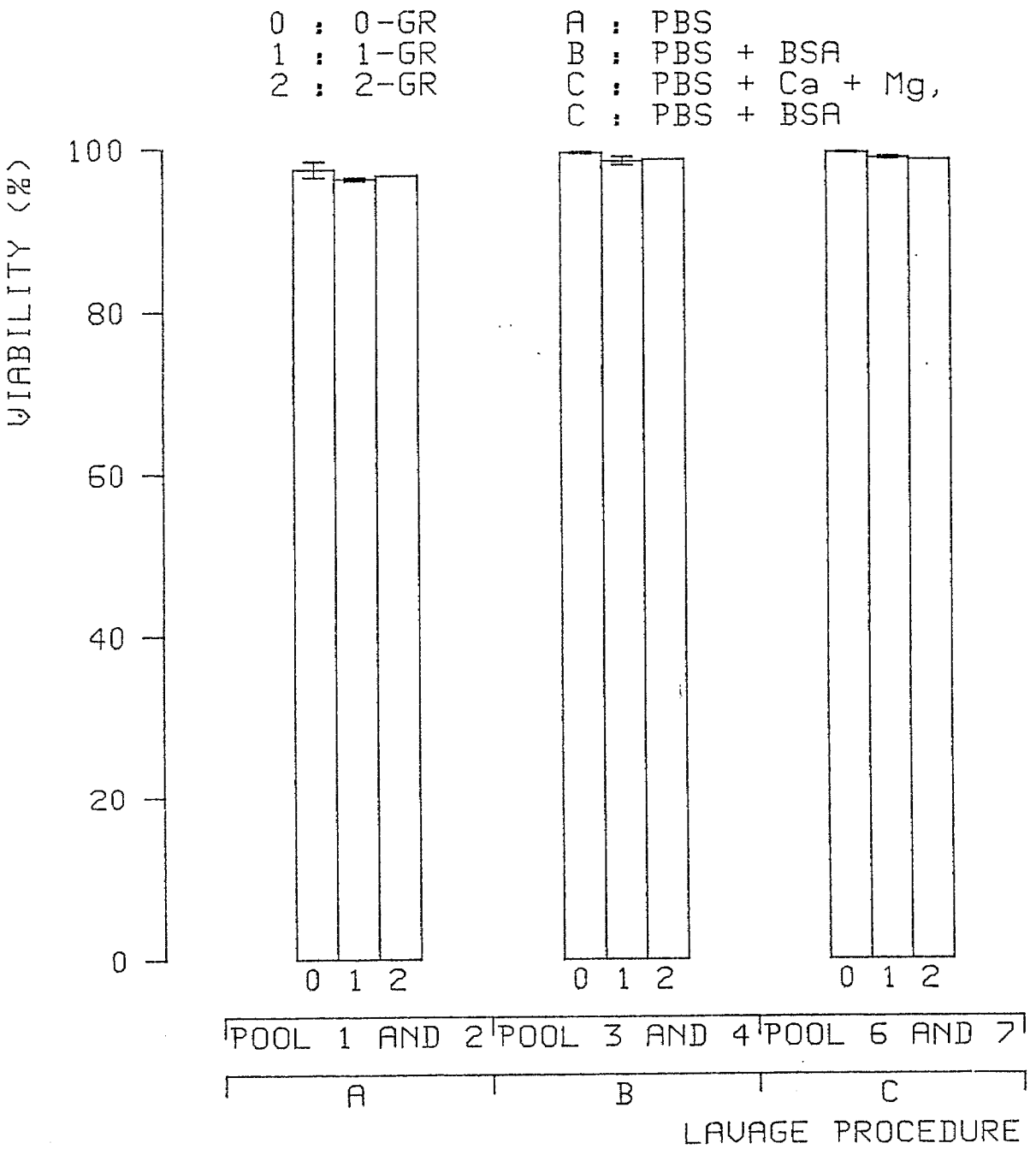
BC FIGURE 35 (continued)

VIABILITY OF MACROPHAGES, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 40

2029027514

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 35 continued

VIABILITY OF MACROPHAGES, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 40

A 0500/3057, H03472, TH, V124 F155 18 R

2029027515

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

BC FIGURE 36

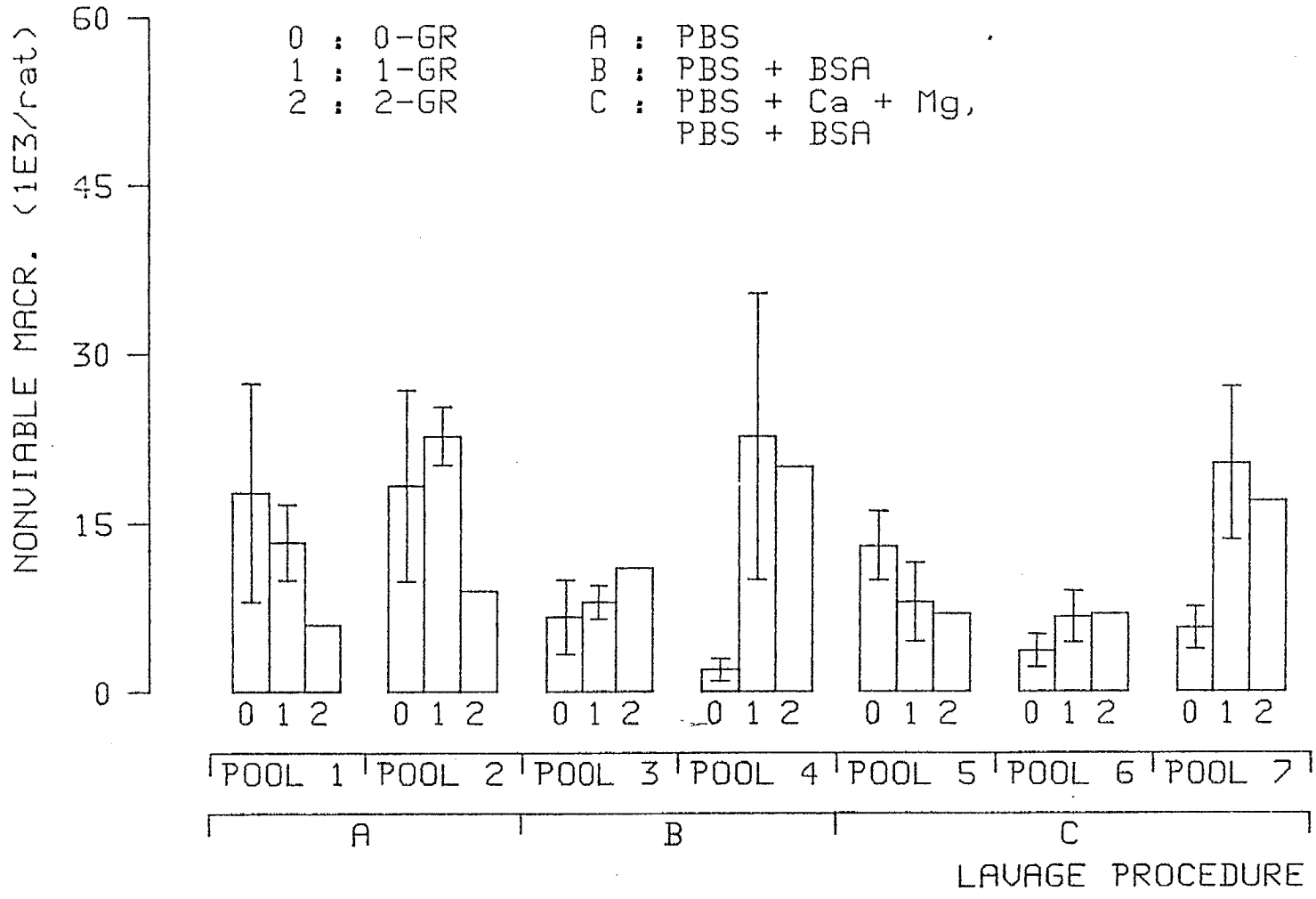
NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 42

20290275l6

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03444, TH, V133 F155 V89 R


0 : 0-GR
1 : 1-GR
2 : 2-GR
A : PBS
B : PBS + BSA
C : PBS + Ca + Mg, PBS + BSA
NONVIABLE MACR. (1E3/rat)
60
45
30
15
0
0 1 2 0 1 2 0 1 2 0 1 2 0 1 2 0 1 2 0 1 2
POOL 1 POOL 2 POOL 3 POOL 4 POOL 5 POOL 6 POOL 7
A
B
C
LAVAGE PROCEDURE

BC FIGURE 36

NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM, INDIVIDUAL POOLS

Remarks : for details see BC TABLE 42

2029027517

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B24 WS M HO3445

BC FIGURE 37

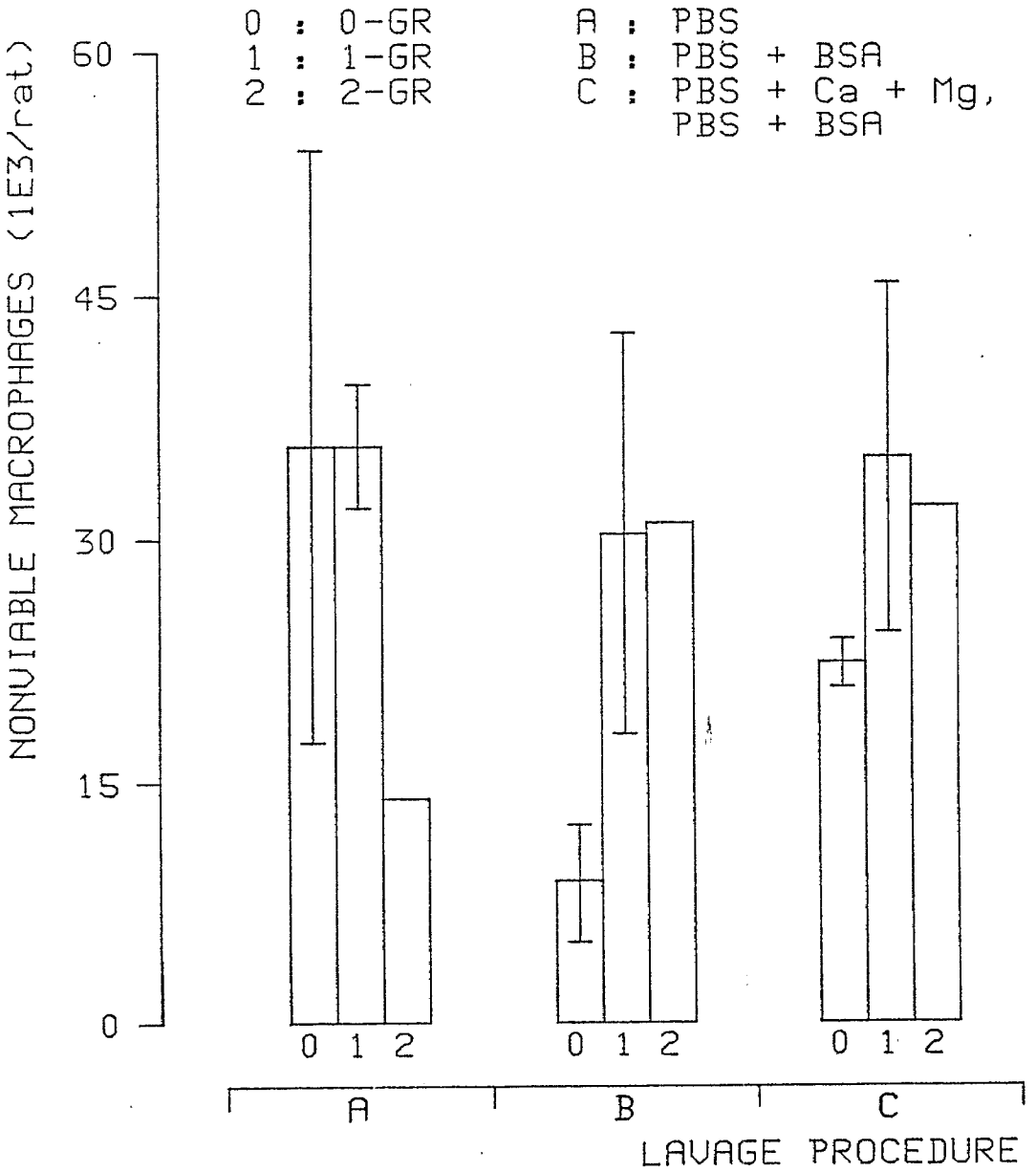
NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM, SUM OF POOLS

Remarks: for details see BC TABLE 42

2029027518

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

Wrke!



BC FIGURE 37

NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM, SUM OF POOLS

Remarks : for details see BC TABLE 42

A 0500/3057, H03445, TH, V134 F155 ' 0 R

2029027519

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

BC FIGURE 37 (continued)

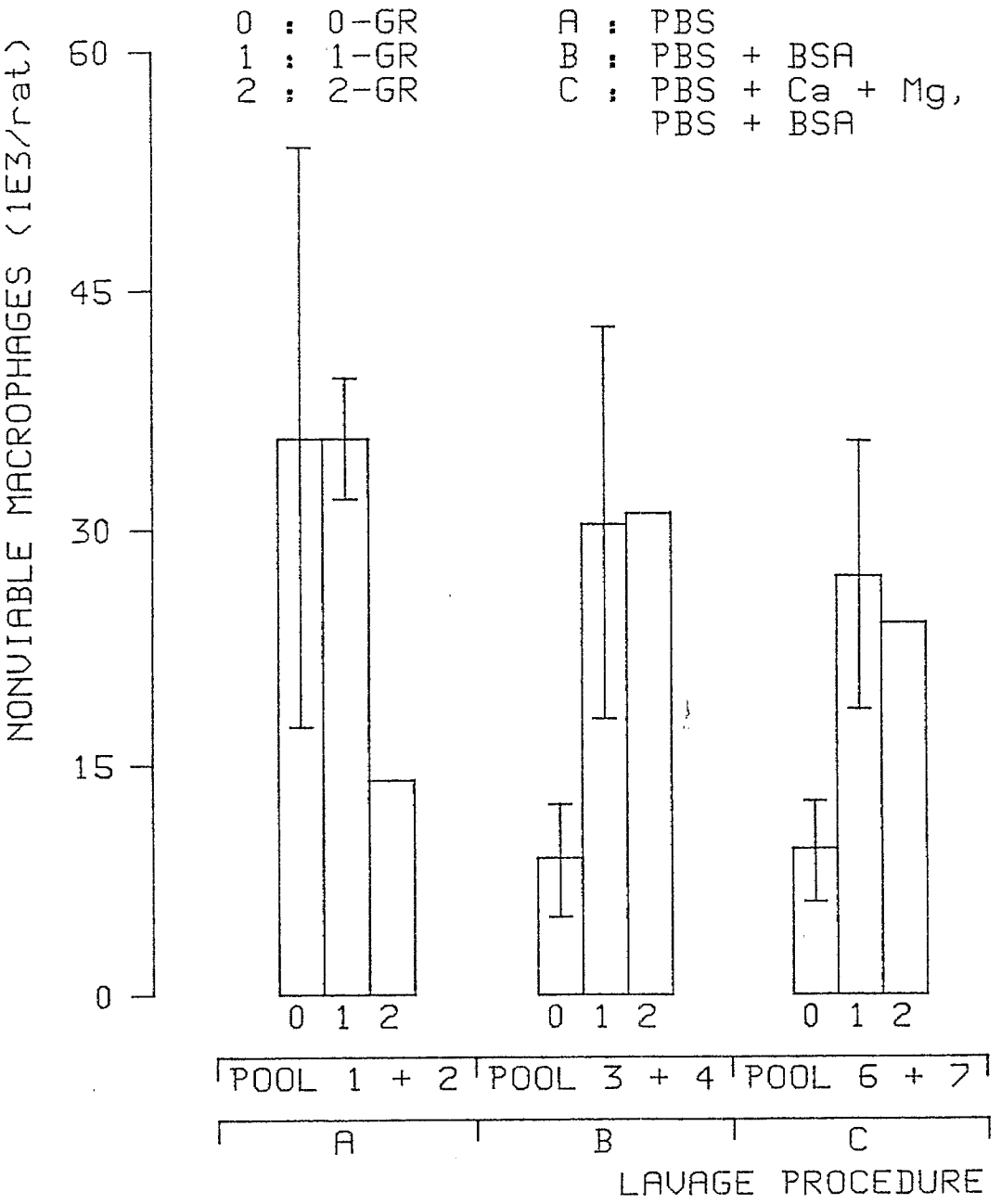
NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM, SUM OF POOLS

Remarks: for details see BC TABLE 42

2029027520

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

Werk!



BC FIGURE 37 continued

NUMBER OF NONVIABLE MACROPHAGES PER RAT, RESUSPENSION MEDIUM, SUM OF POOLS

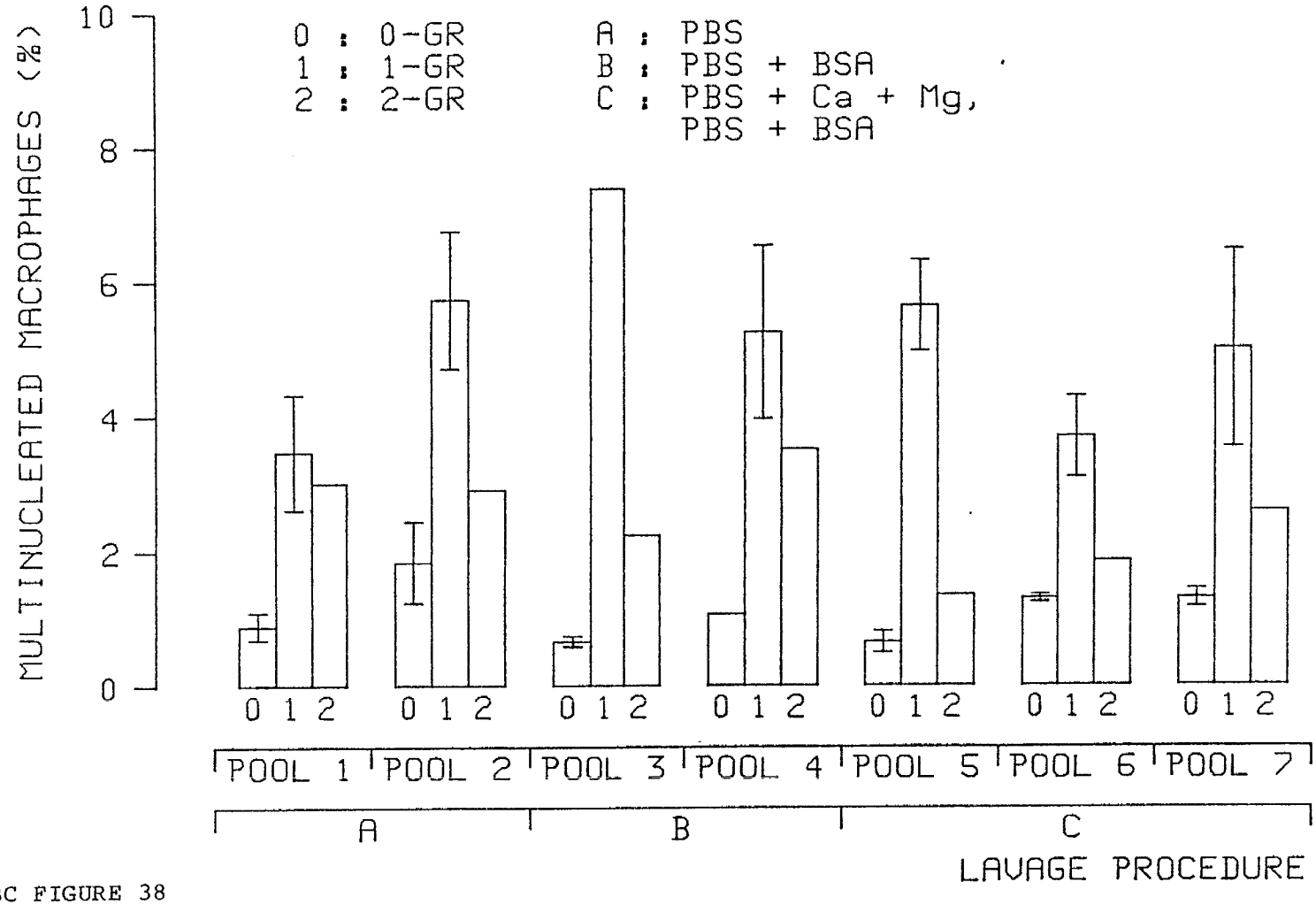
Remarks : for details see BC TABLE 42

A 0500/3057, H03446, TH, V135 F155 1 R

2029027521

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

2284


MULTINUCLEATED MACROPHAGES (%)
10
8
6
4
2
0
0 : 0-GR
1 : 1-GR
2 : 2-GR
A : PBS
B : PBS + BSA
C : PBS + Ca + Mg, PBS + BSA
0 1 2 0 1 2 0 1 2 0 1 2 0 1 2 0 1 2 0 1 2
POOL 1 POOL 2 POOL 3 POOL 4 POOL 5 POOL 6 POOL 7
A
B
C
LAVAGE PROCEDURE

BC FIGURE 38

RELATIVE NUMBER OF MULTINUCLEATED MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 44

20290275Z2

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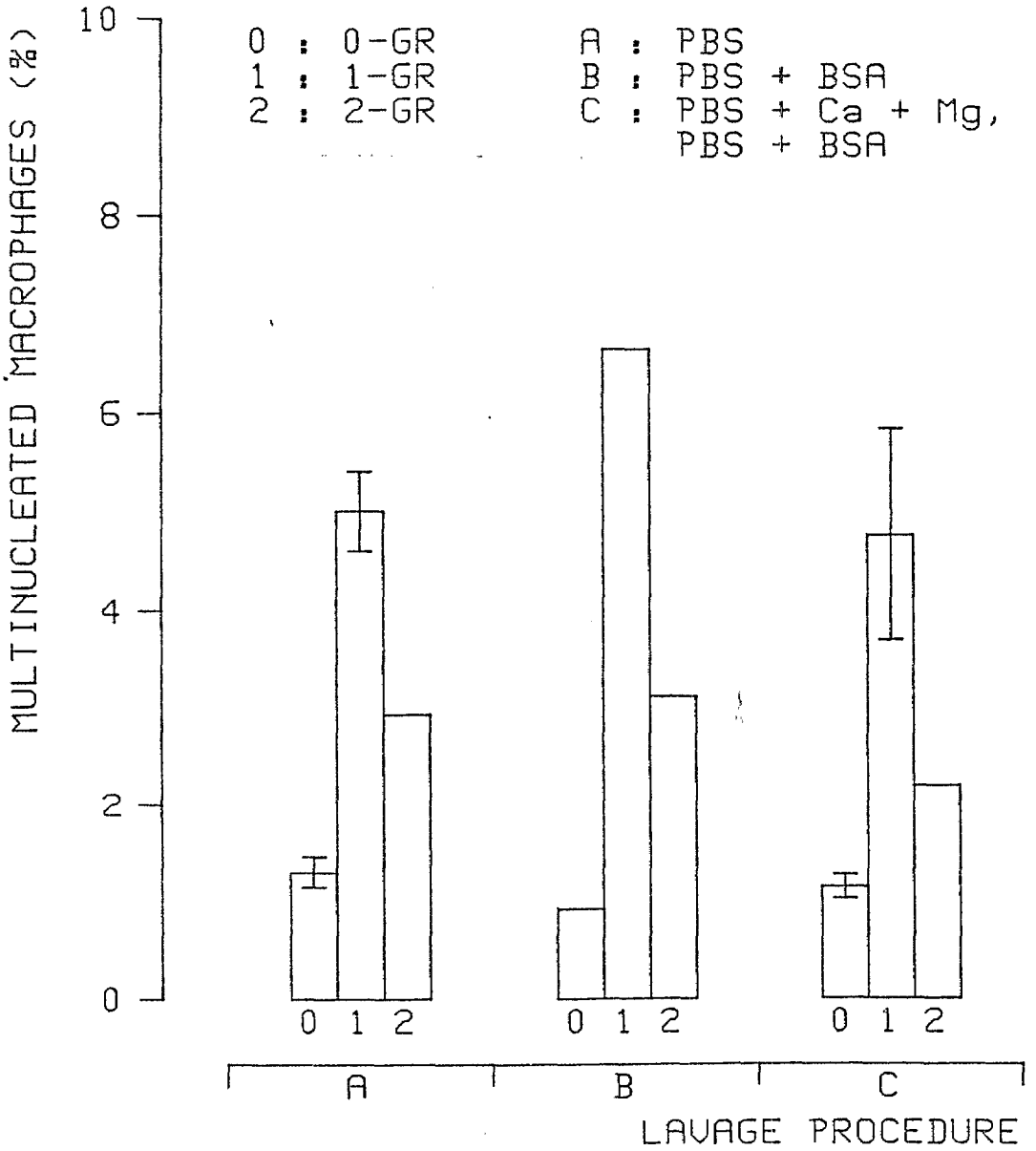
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 39

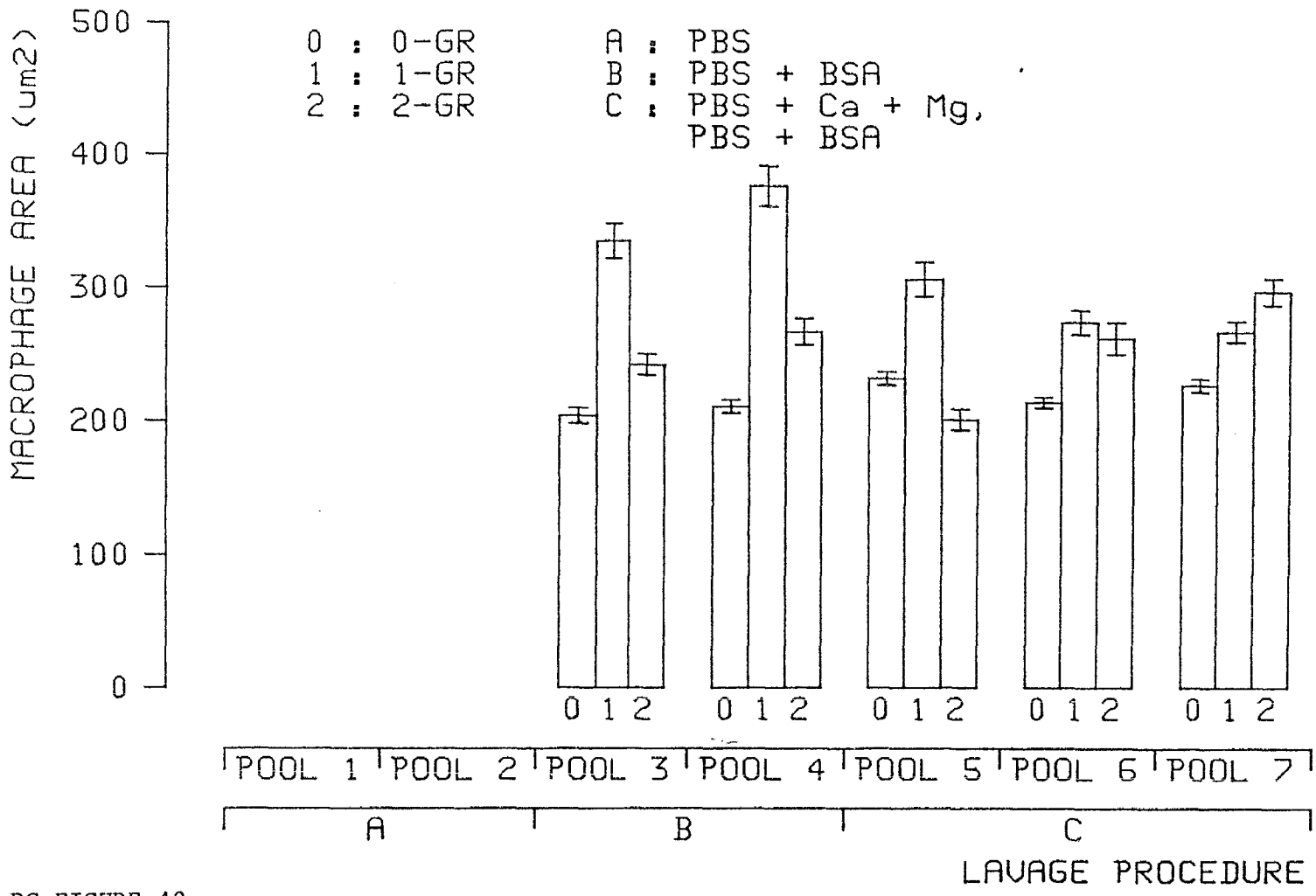
RELATIVE NUMBER OF MULTINUCLEATED MACROPHAGES, RESUSPENSION MEDIUM, CYTOCENTRIFUGE PREPARATION, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 44

A 0500/3057, HO3198, TH, U88 F155 U79, R

2029027523

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 40

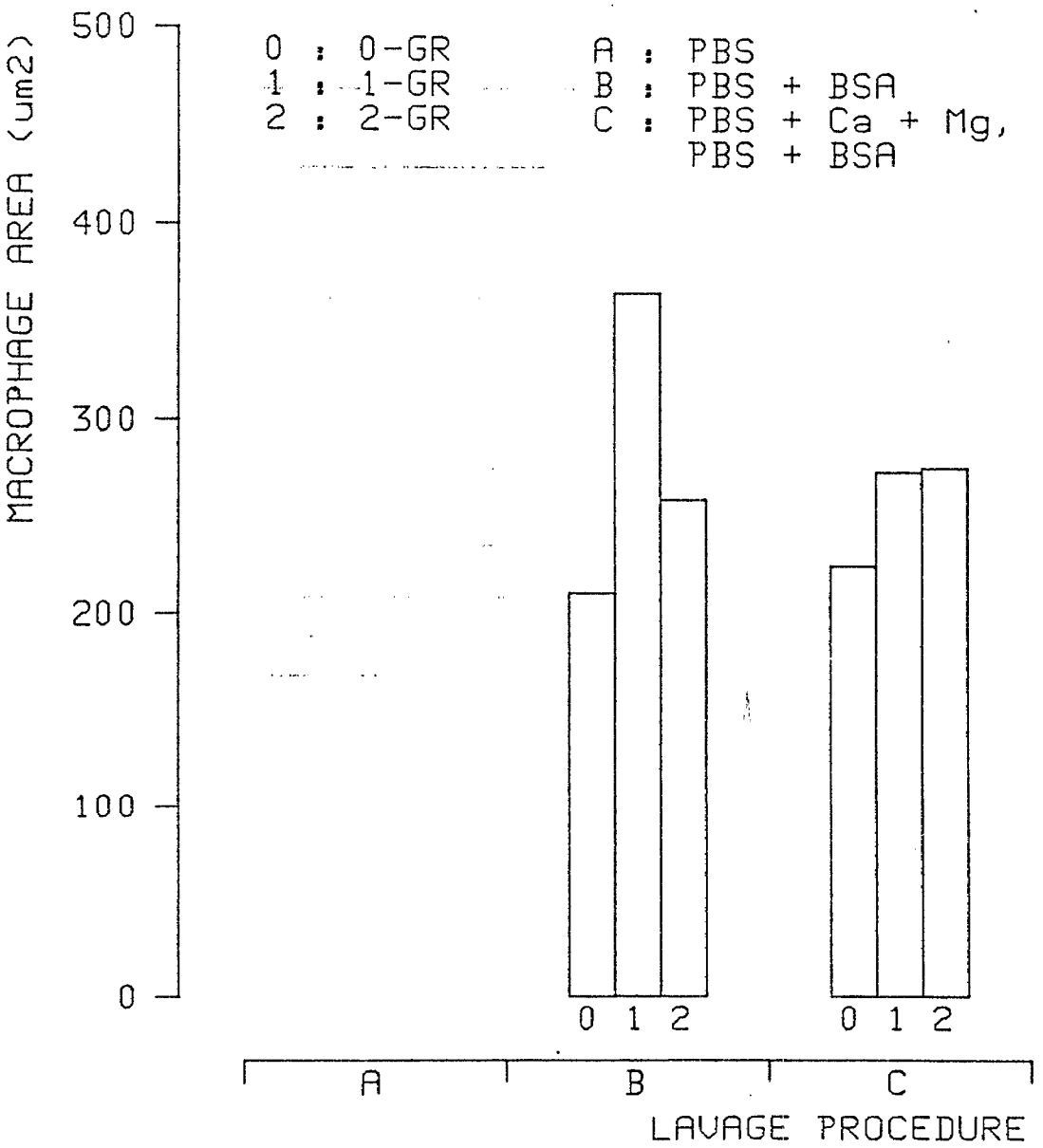
MEAN MACROPHAGE AREA, RESUSPENSION MEDIUM, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 47

20290275Z4

INBIFO Institut für biologische Forschung · Köln

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 41

MACROPHAGE AREA, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

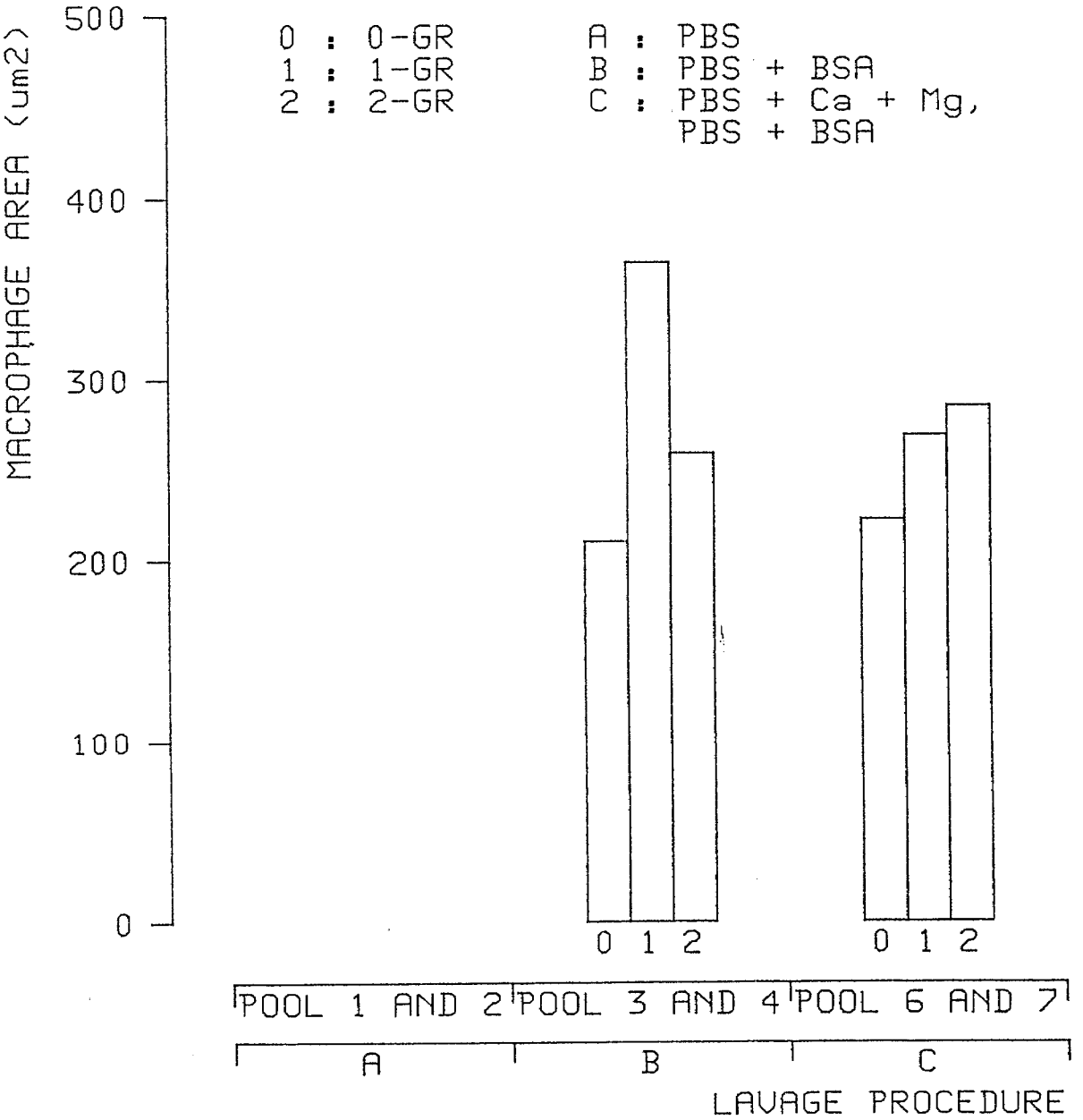
Remarks: for details see BC TABLE 47

A 0500/3057, HO3460, TH, V10S F15S 1'90, R

2029027525

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B25 WS M HO3461



BC FIGURE 41 (continued)

MACROPHAGE AREA, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

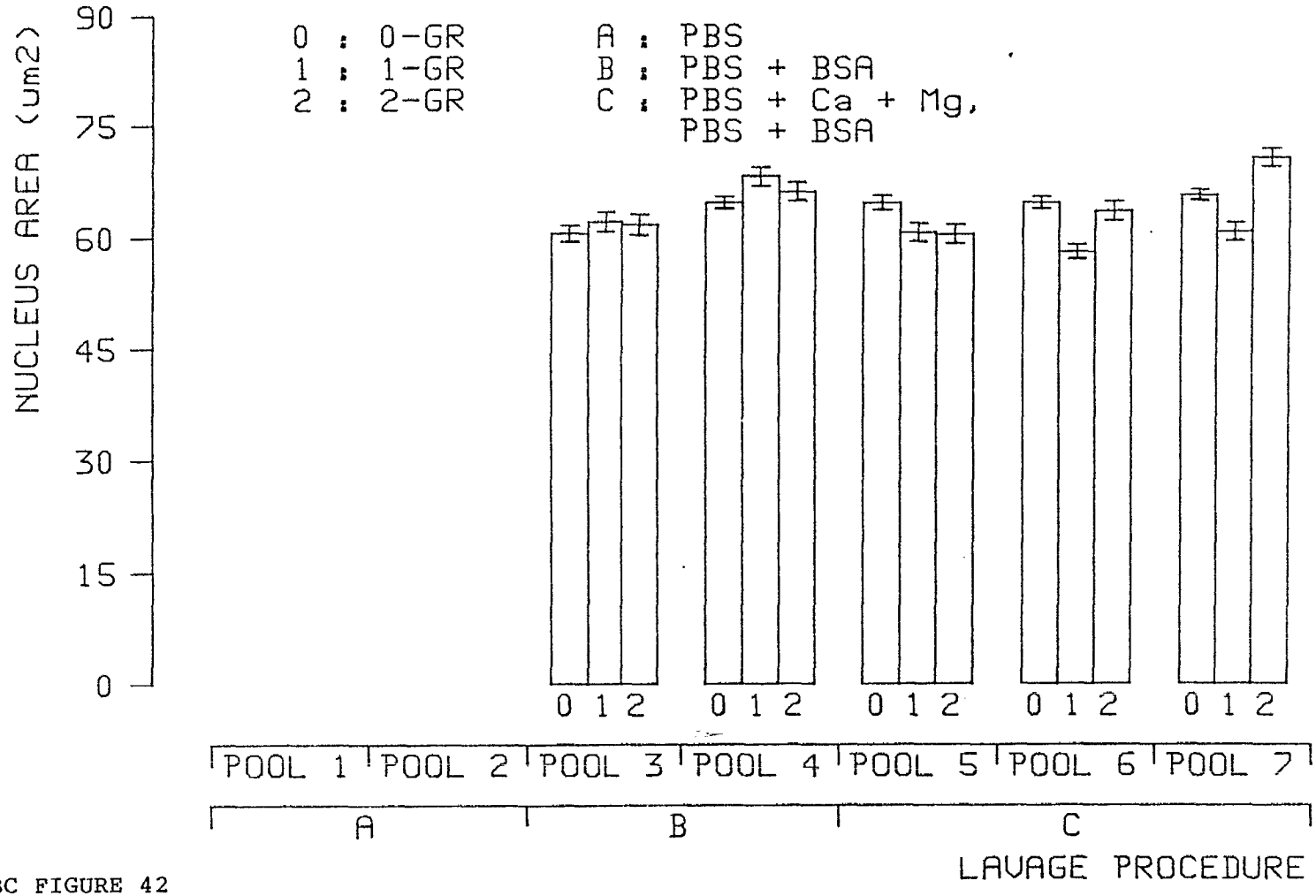
Remarks: for details see BC TABLE 47

A 0500/3057, HO3461, TH, U125 F155 / J6 R

2029027526

2284

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 42

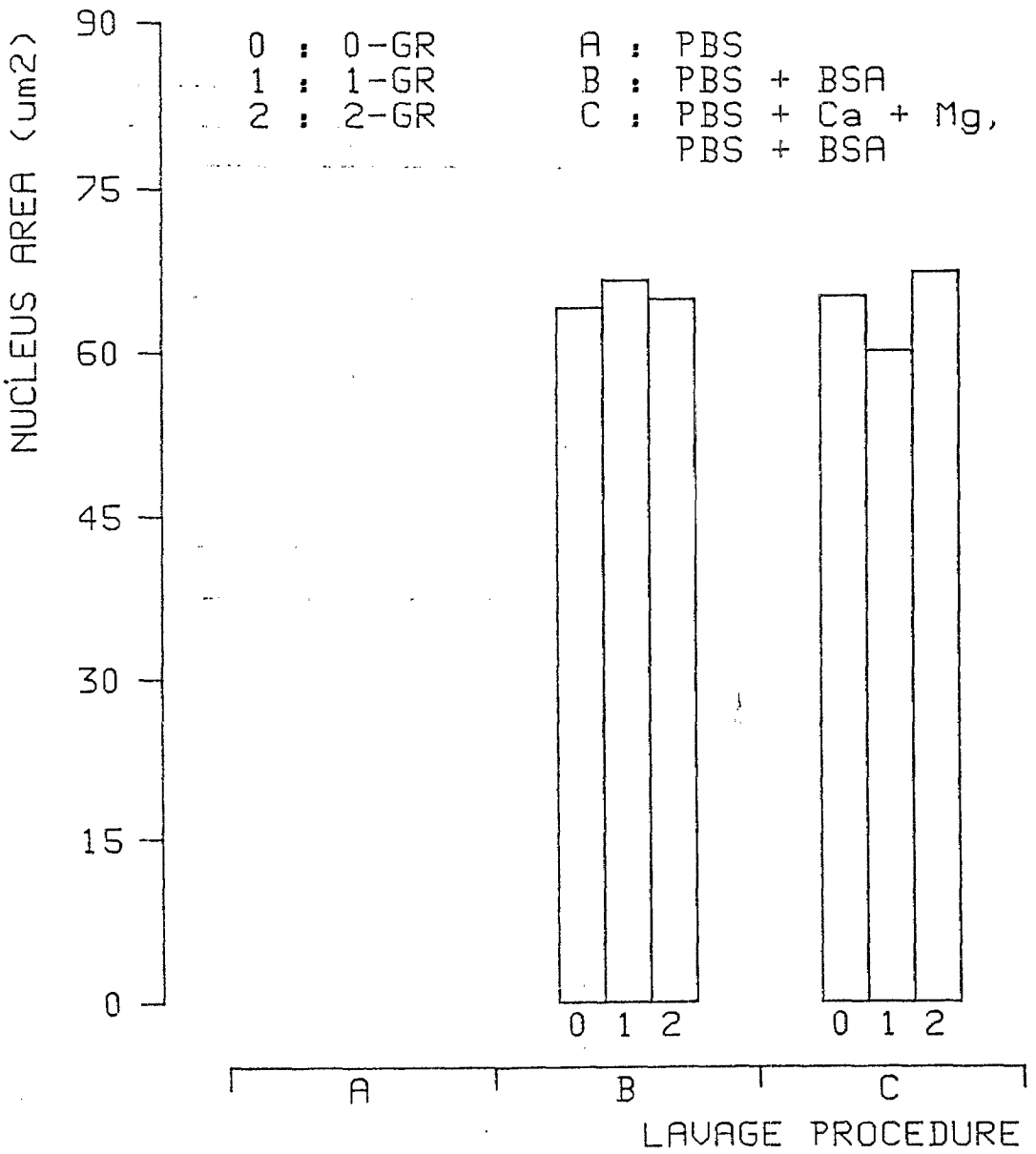
MEAN NUCLEUS AREA, RESUSPENSION MEDIUM, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 49

INBIFO Institut für biologische Forschung · Köln

20290275Z7

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 43

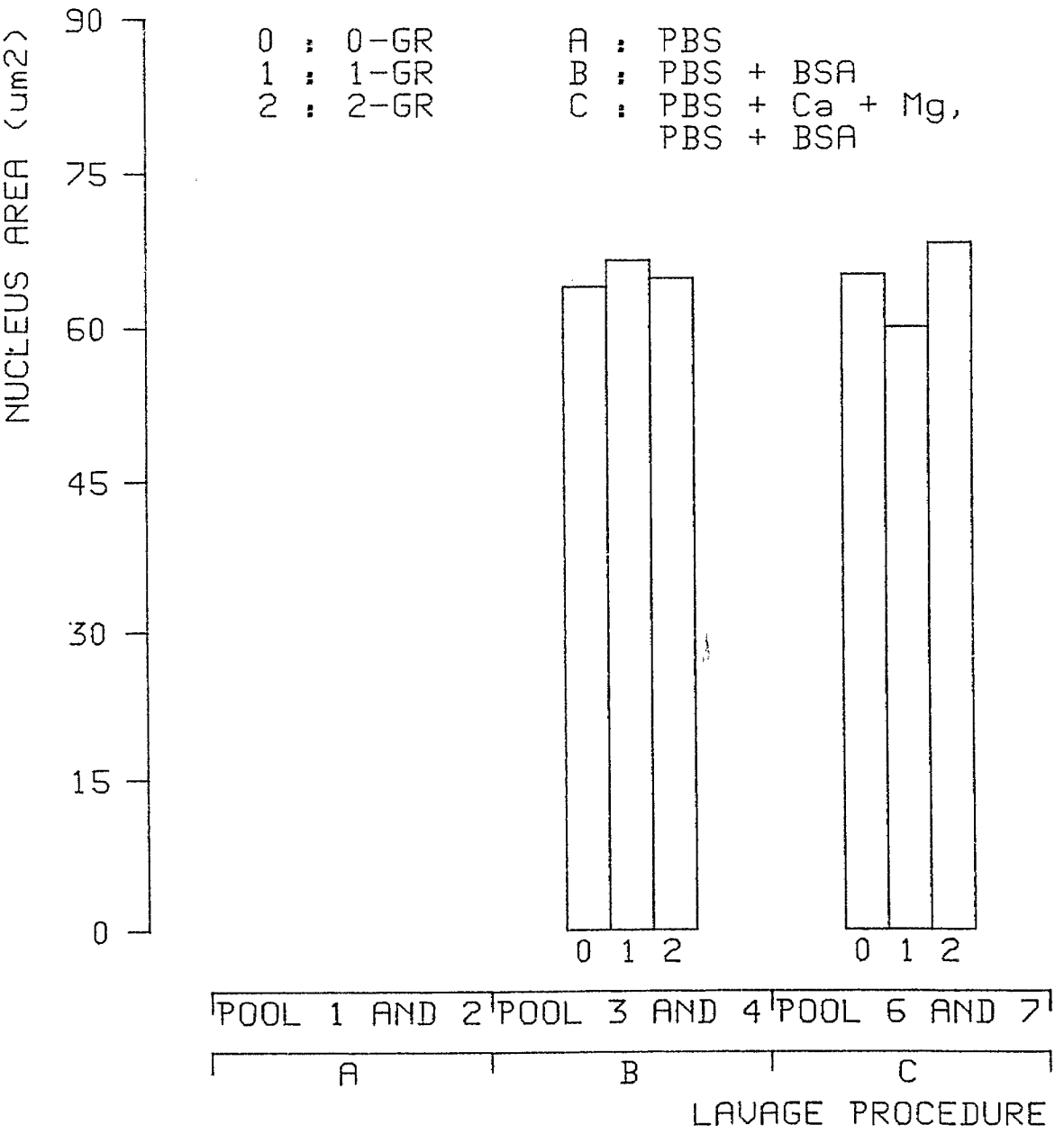
NUCLEUS AREA, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 49

A 0500/3052, HO3463, TH, V108 F155 U105, R

2029027528

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 43 (continued)

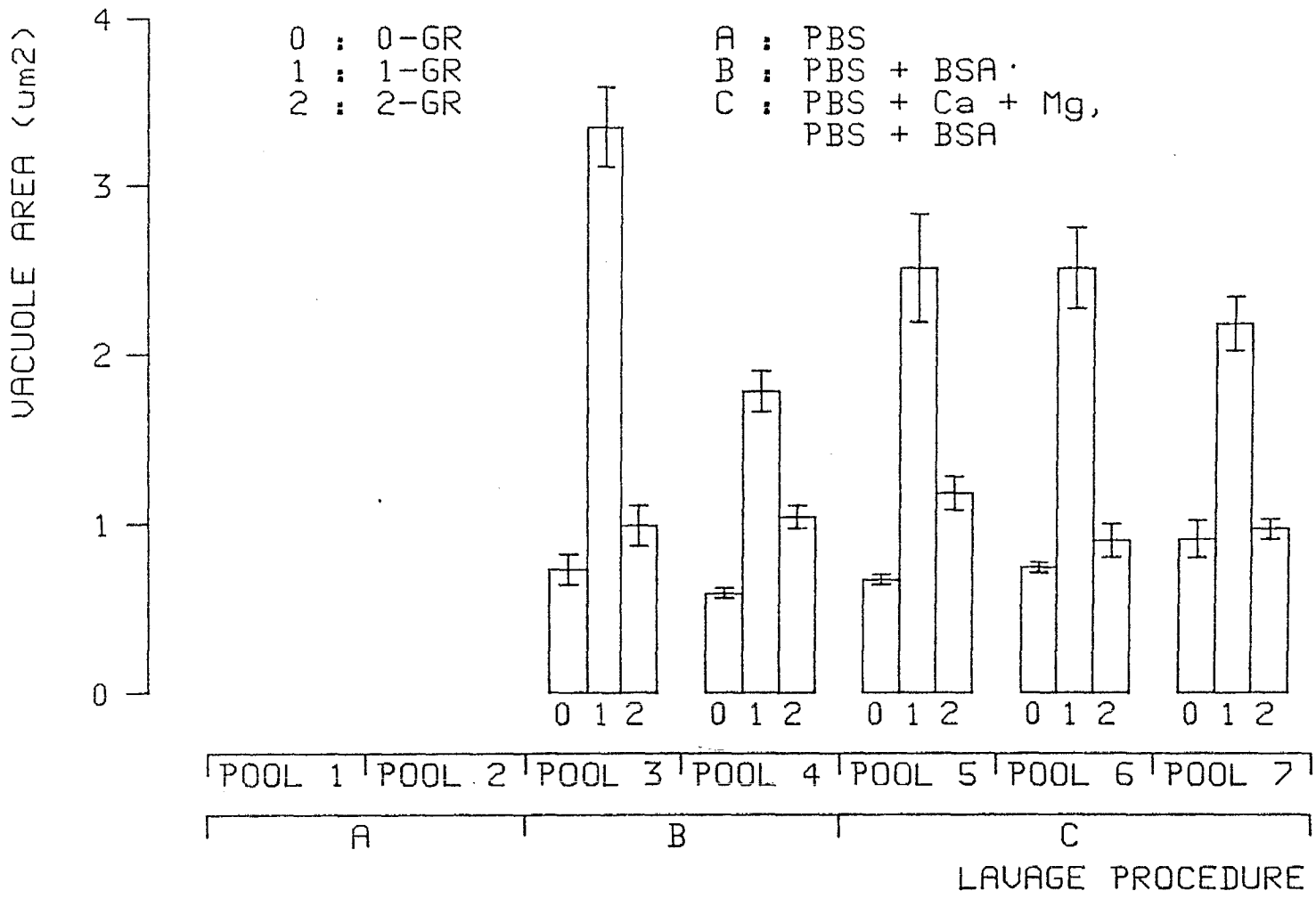
NUCLEUS AREA, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 49

A 0500/3057, HO3464, TH, V126 F155 ' 19 R

2029027529

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 44

MEAN VACUOLE AREA, RESUSPENSION MEDIUM, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 51

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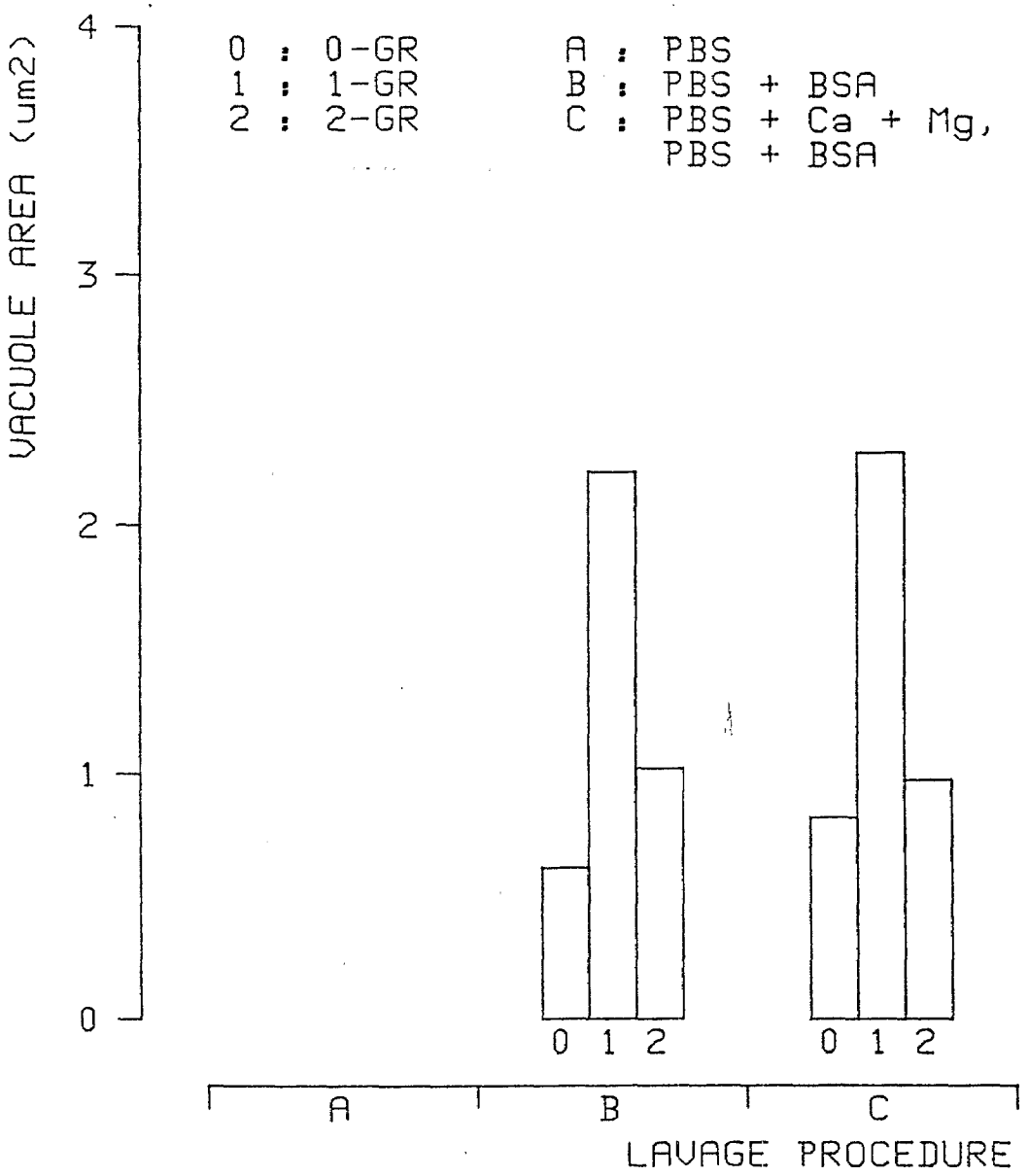
20290275 30

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 45

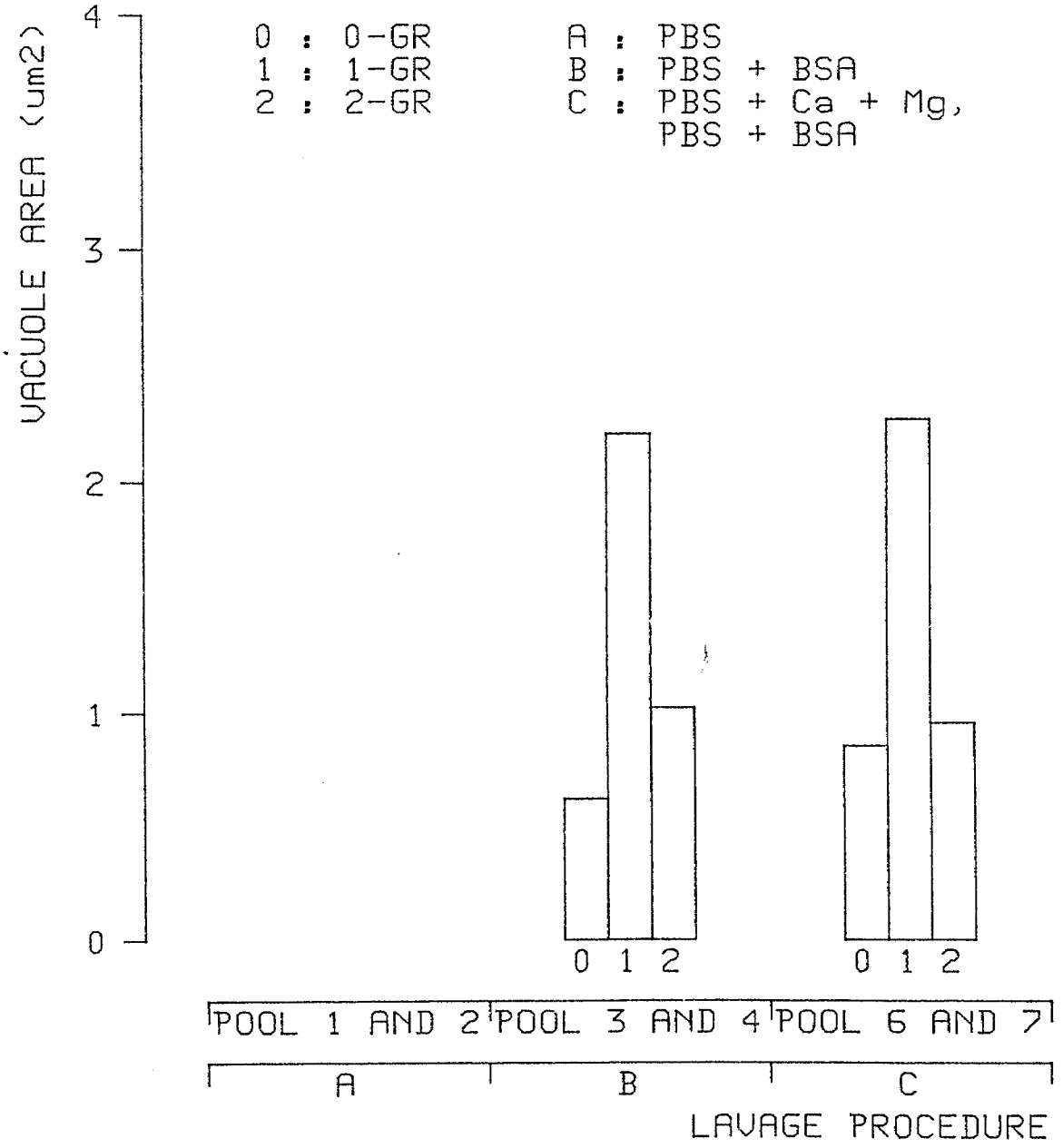
VACUOLE AREA, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 51

A 0500/3057, HO3466, TH, U111 F155 U'08, R

2029027531

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 45 (continued)

VACUOLE AREA, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

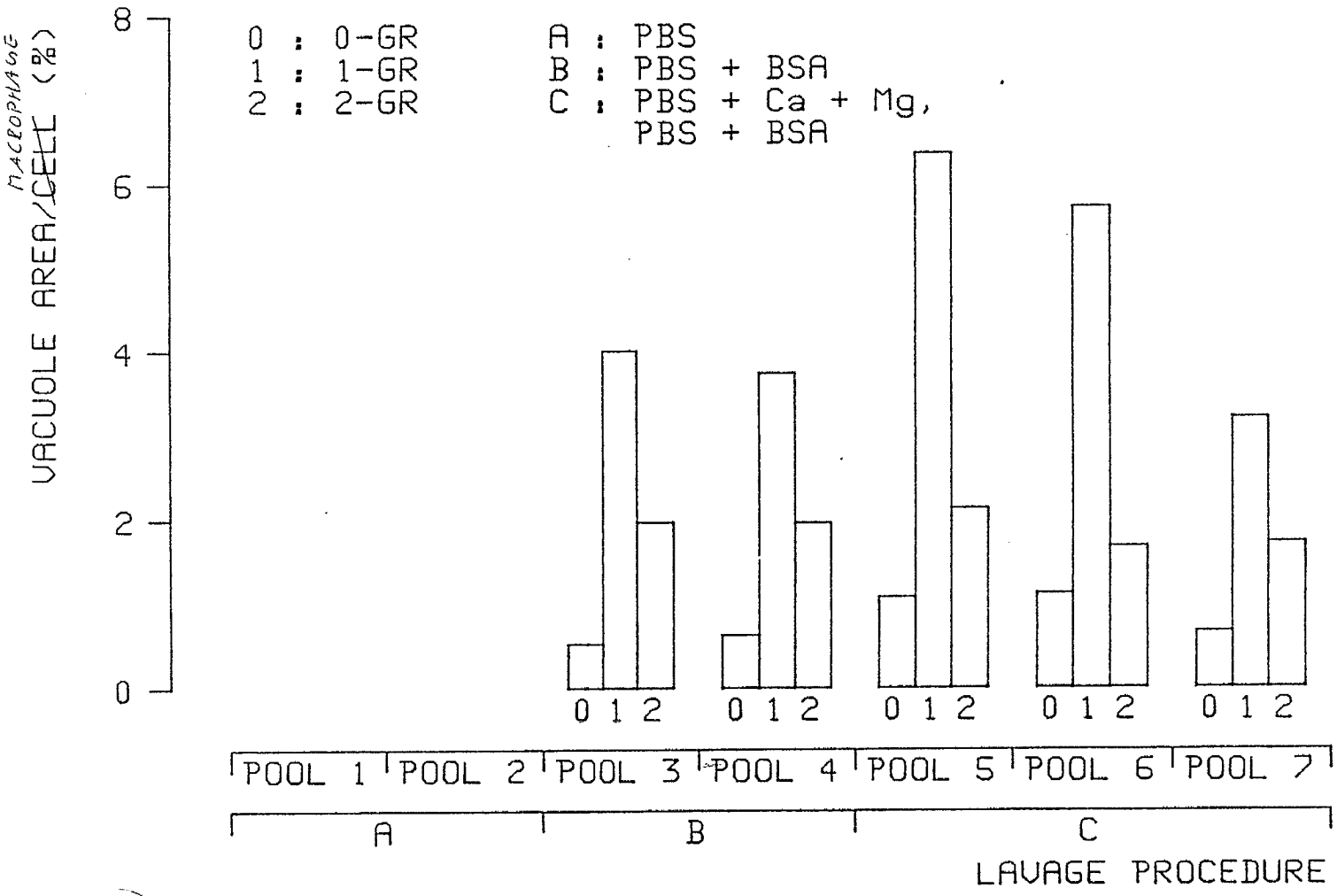
Remarks: for details see BC TABLE 51

A 0500/3057, HO3467, TH, V127 F155 - 12 R

2029027532

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

2284



BC FIGURE 46

RELATIVE MACROPHAGE
VACUOLE AREA PER ~~CELL~~, RESUSPENSION MEDIUM, INDIVIDUAL POOLS

Remarks: for details see BC TABLE 52

2029027533

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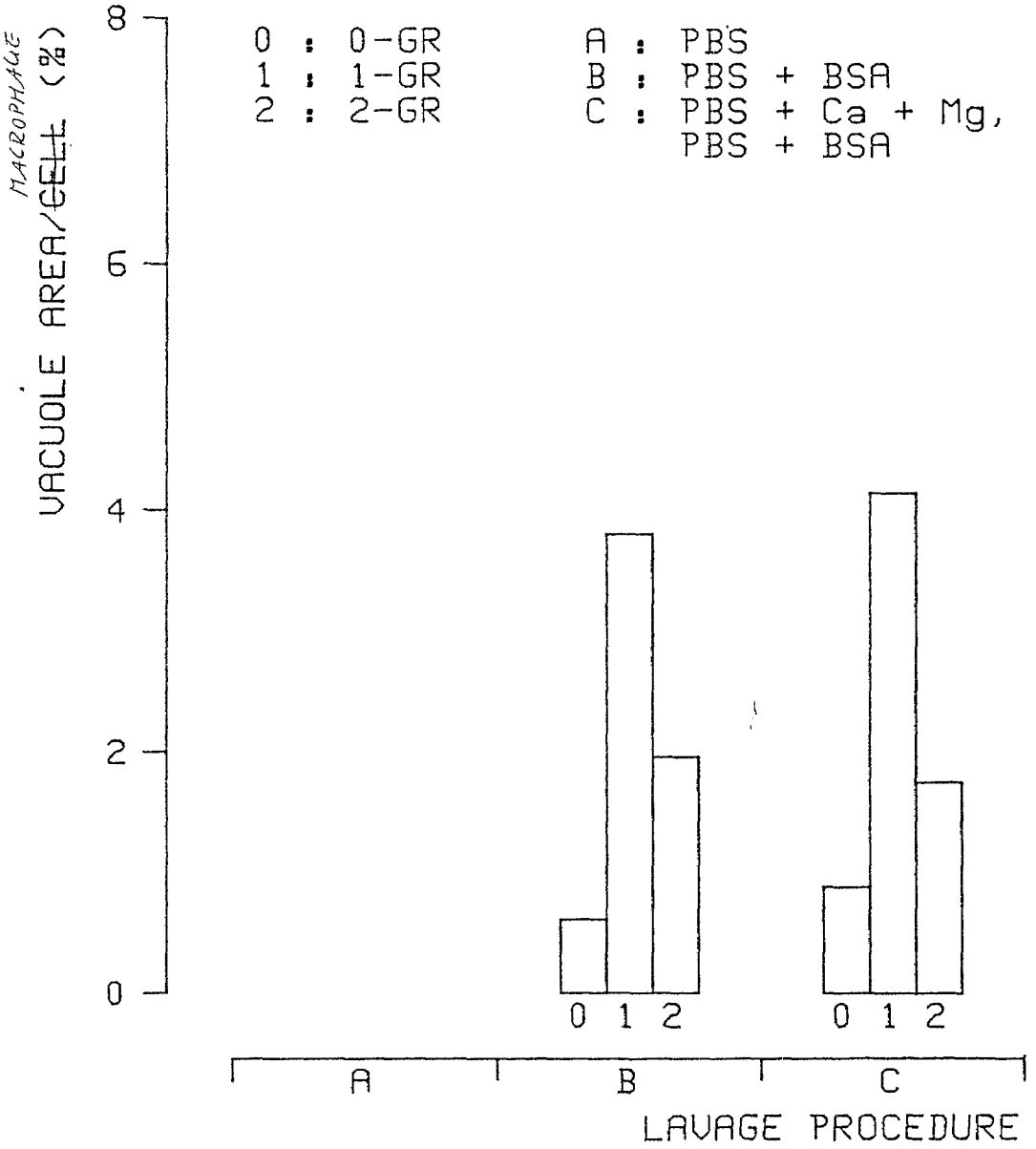
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

SUBREPORT P 0500/3057 GD151 (R) B27 WS M HO3469



BC FIGURE 47

(RELATIVE) MACROPHAGE

VACUOLE AREA PER ~~CELL~~, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

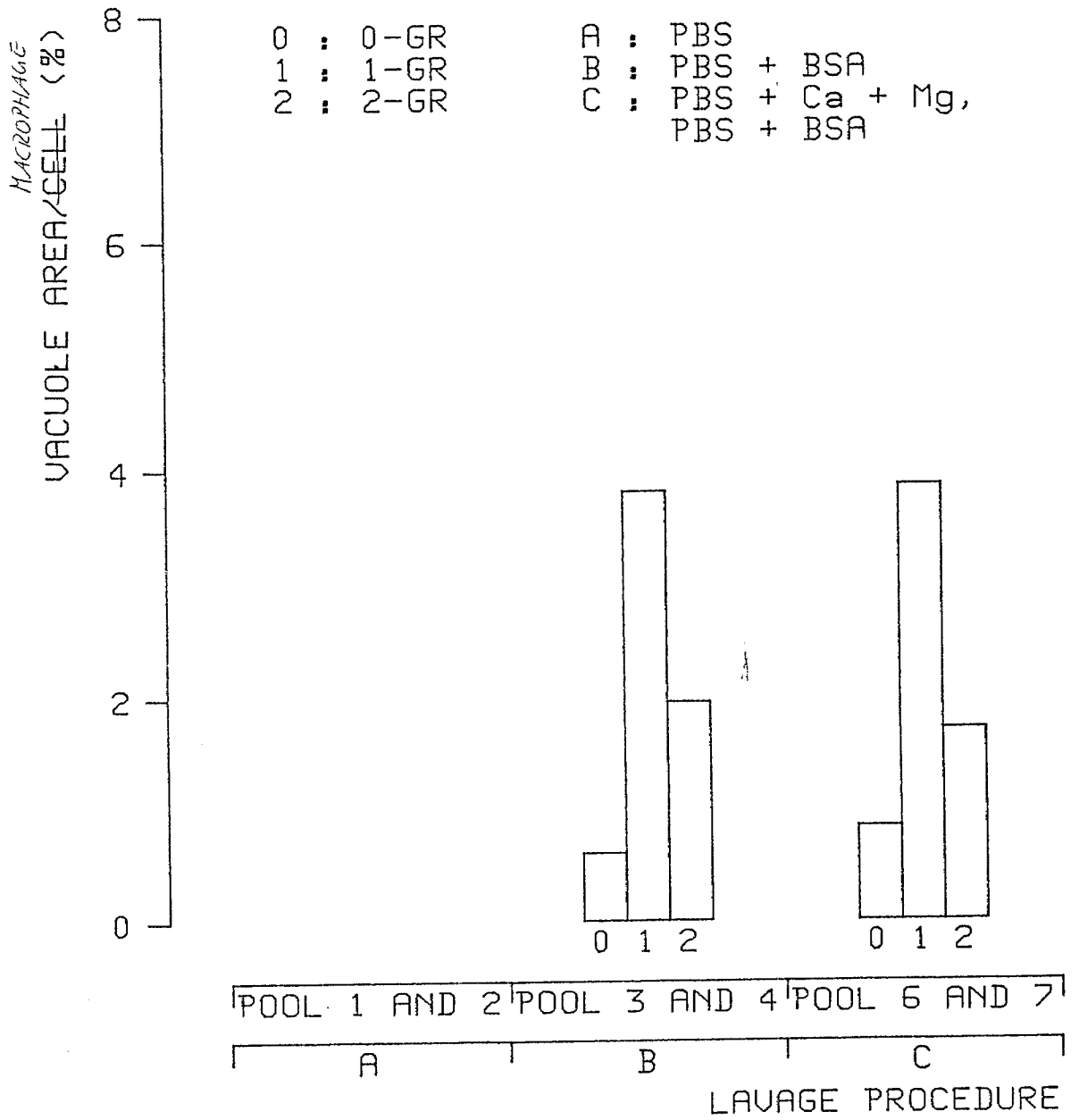
Remarks: for details see BC TABLE 52

A 0500/3057, HO3469, TH, U114 F155 U111, R

2284

2029027534

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE 47 (continued)
RELATIVE MACROPHAGE
VACUOLE AREA PER ~~CELL~~, RESUSPENSION MEDIUM, WEIGHTED MEANS OF POOLS

Remarks: for details see BC TABLE 52

A 0500/3057, HO3470, TH, U128 F155 U115 R

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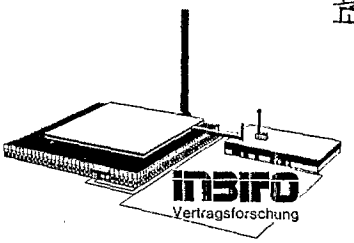
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A/3057



A. Viability

X1 Histogr. / Cytogr.
FWD, AXL

X3 anderes File mit wenig n < 1000

X4 Cytogr. viable, intermediate, nonviable

IC viable cells, aus Fokusauswertung oder FWD vs AXL

B. Phagocytose

X8 wie X1 aus Viab. / L012P.0, L122P.0

X9 GRN T=40 min

X10 Histogr. MP, corr + uncorr, PMN, wie 3092

X11 Cytogramm Grn vs Axl, corr. + uncorr.

FWD 2-GR: Buckel?

X13 RAS, alle 3 Gruppe als Histogramme

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Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

C) ~~DNA~~

X15 wie X1 FWD vs AXL

X16.1 RD RDP mit Reg.
.2 MΦ, PMN
+
X18.1 MΦ, A, P, 60
18.2 PMN, A, P, 60

→ Histogram RDA, nur MΦ aus Cytogr. FWD vs AXL
RDP

X19. DNA RD → FWD vs AXL
alle Gruppen
60, 2x60, 62
} MΦ

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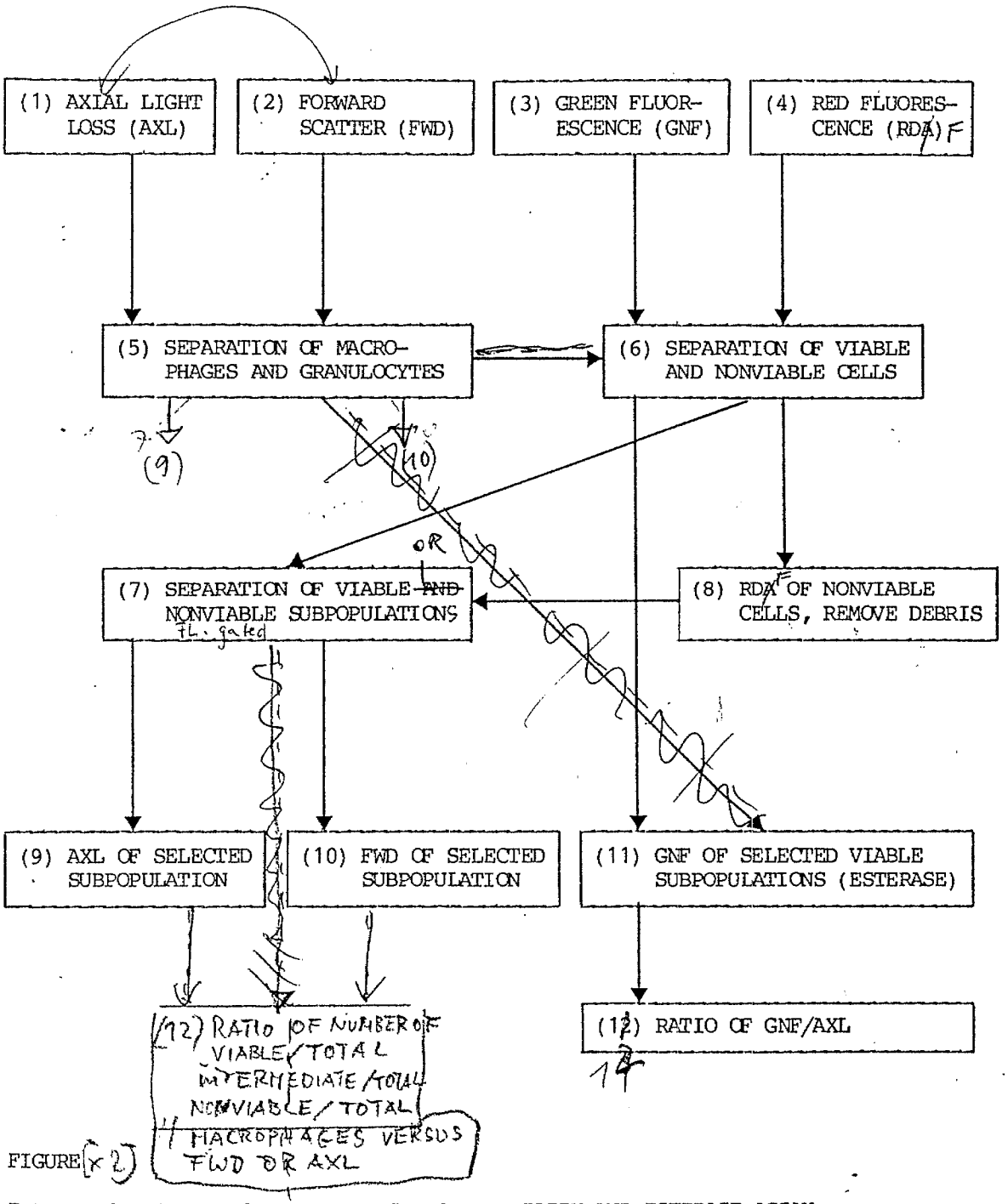
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

Fig [XV]

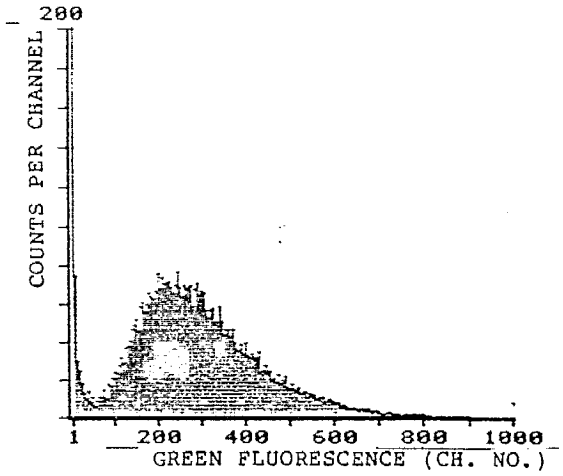
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

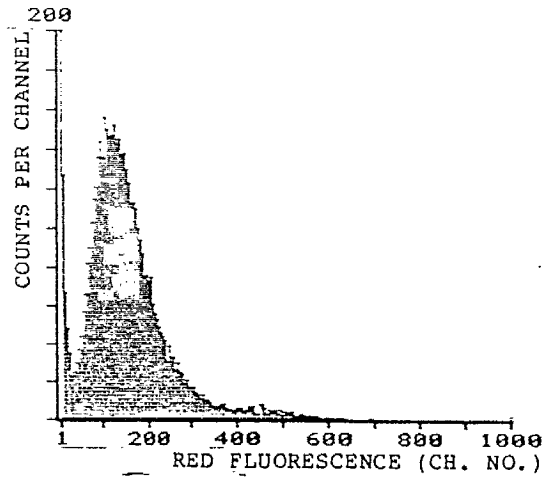

(1) AXIAL LIGHT LOSS (AXL)
(2) FORWARD SCATTER (FWD)
(3) GREEN FLUORESCENCE (GNF)
(4) RED FLUORESCENCE (RDA)
(5) SEPARATION OF MACROPHAGES AND GRANULOCYTES
(6) SEPARATION OF VIABLE AND NONVIABLE CELLS
(9)
(10)
OR
(7) SEPARATION OF VIABLE AND NONVIABLE SUBPOPULATIONS
(8) RDA OF NONVIABLE CELLS, REMOVE DEBRIS
(9) AXL OF SELECTED SUBPOPULATION
(10) FWD OF SELECTED SUBPOPULATION
(11) GNF OF SELECTED VIABLE SUBPOPULATIONS (ESTERASE)
(12) RATIO OF NUMBER OF VIABLE/TOTAL INTERMEDIATE/TOTAL NONVIABLE/TOTAL MACROPHAGES VERSUS FWD OR AXL
(12) RATIO OF GNF/AXL

EVALUATION SCHEME FOR PARAMETERS FROM VIABILITY AND ESTERASE ASSAY

2029027539

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000





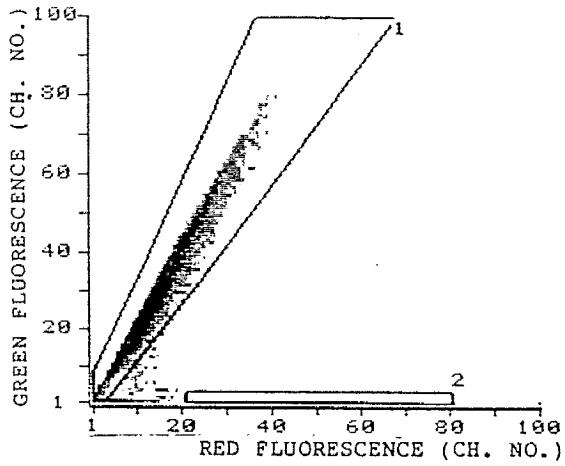


Fig 3

20290275 40

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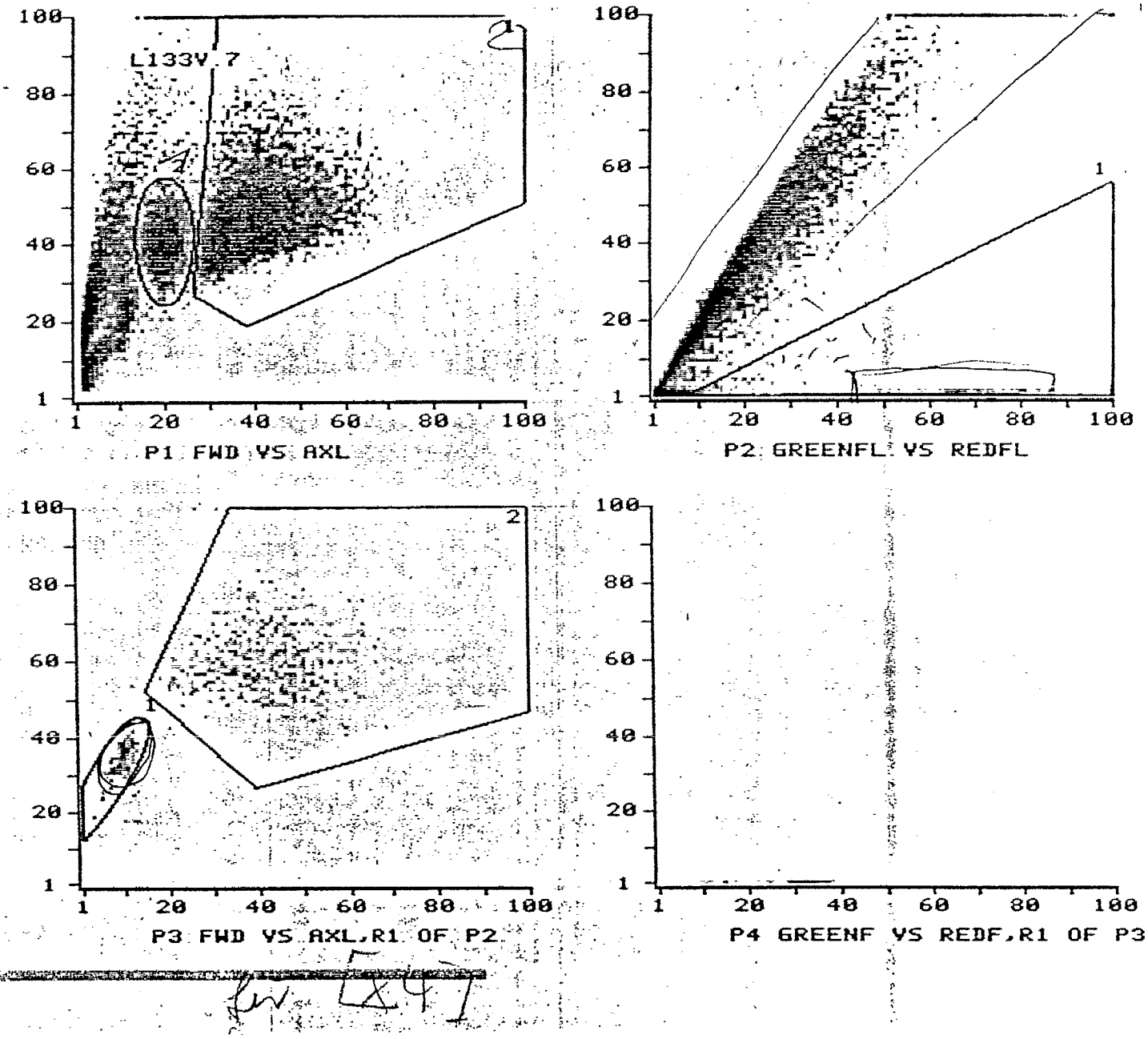

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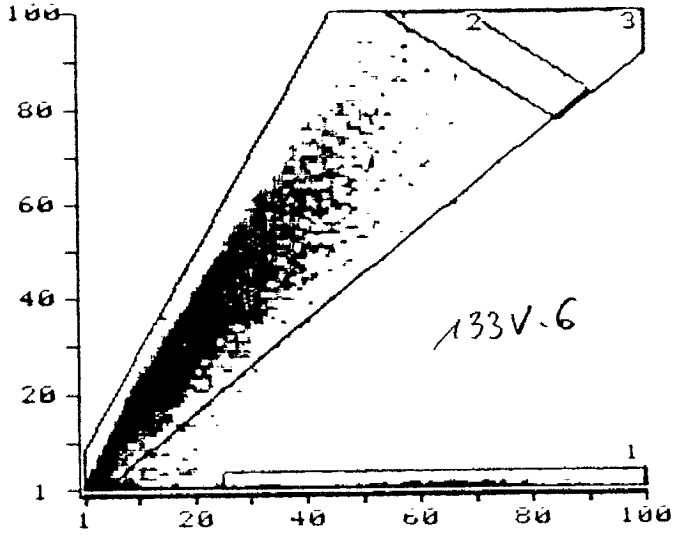

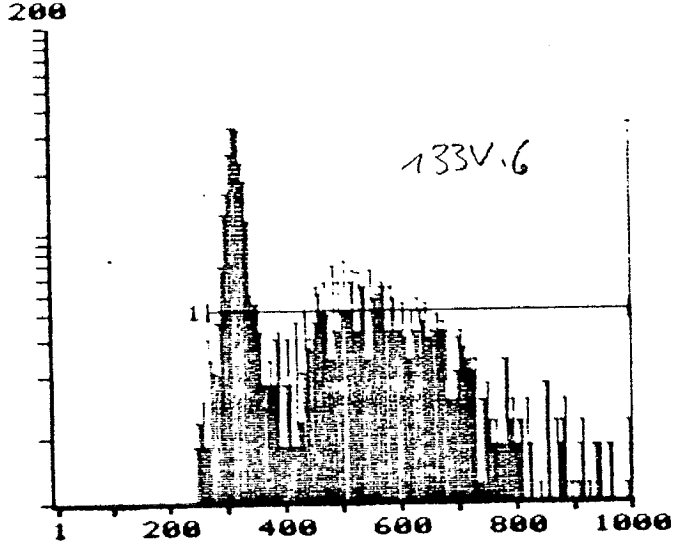

[X5]

20290275Z42

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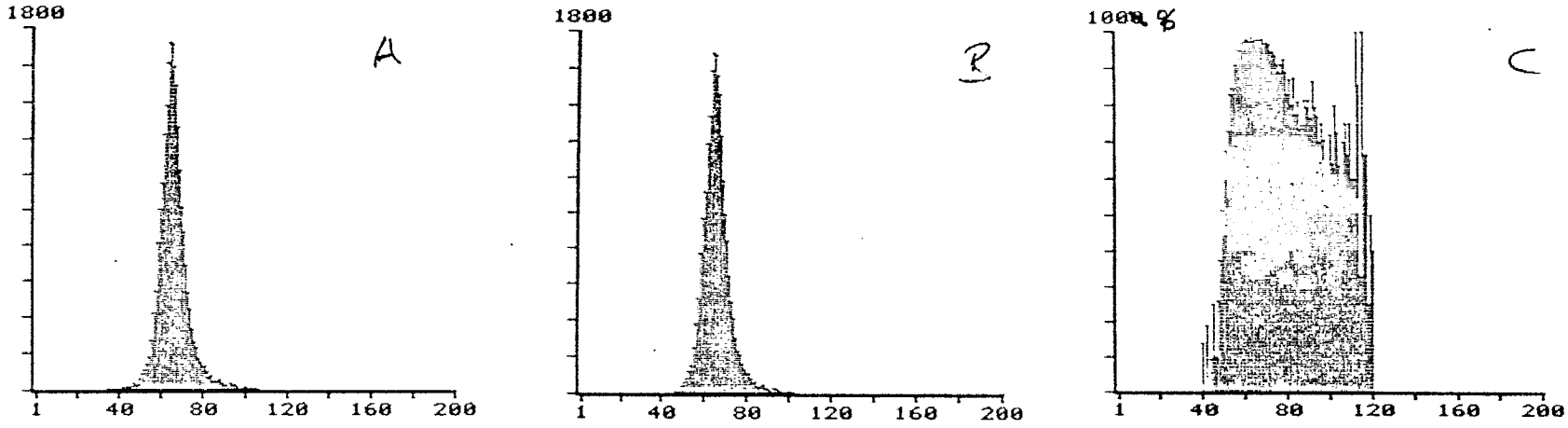
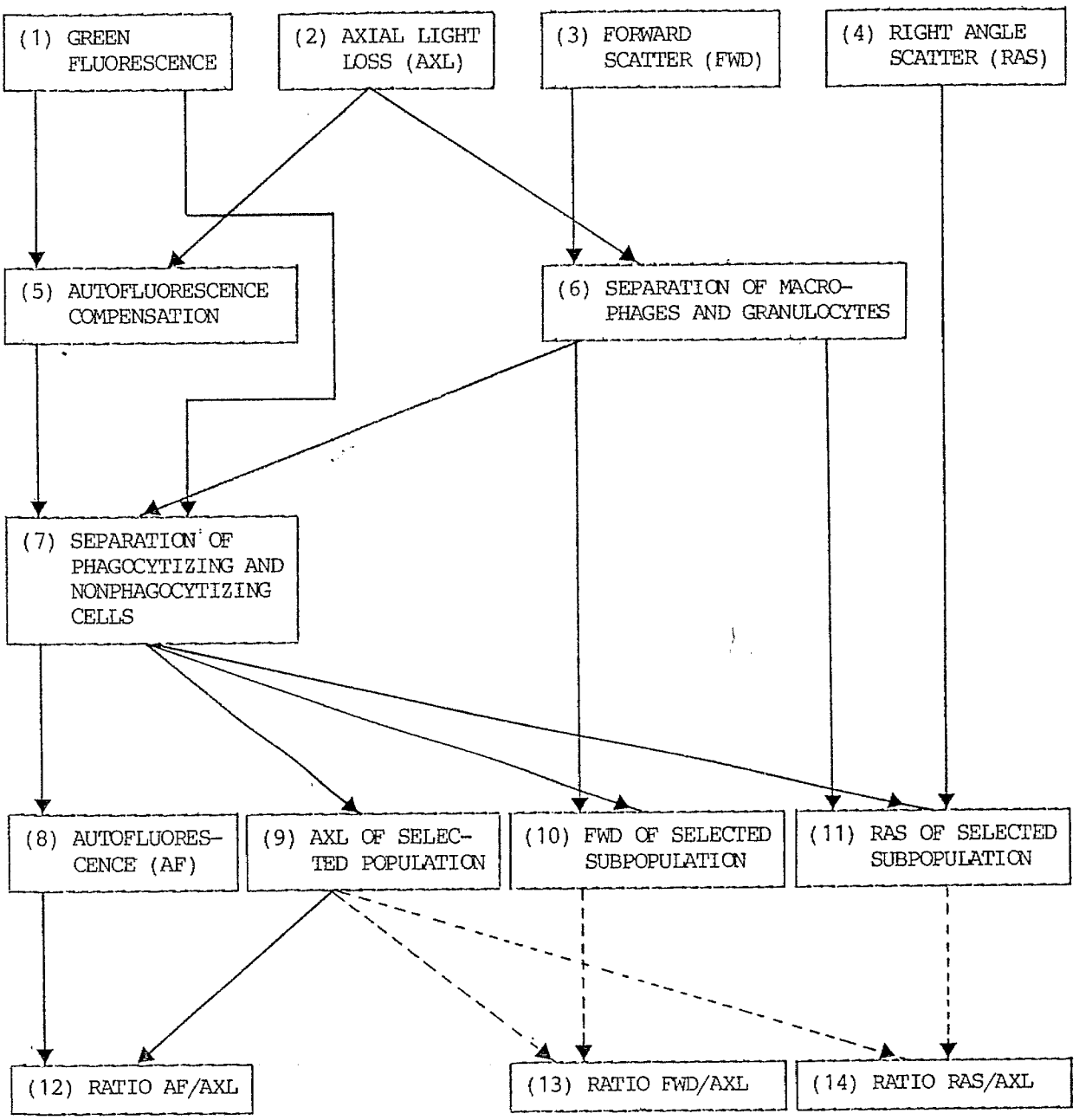
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



X6.1 axl
.2 pkd

0-6R
glands file

20290275 43


(1) GREEN FLUORESCENCE
(2) AXIAL LIGHT LOSS (AXL)
(3) FORWARD SCATTER (FWD)
(4) RIGHT ANGLE SCATTER (RAS)
(5) AUTOFLUORESCENCE COMPENSATION
(6) SEPARATION OF MACRO-PHAGES AND GRANULOCYTES
(7) SEPARATION OF PHAGOCYTIZING AND NONPHAGOCYTIZING CELLS
(8) AUTOFLUORES-CENCE (AF)
(9) AXL OF SELEC-TED POPULATION
(10) FWD OF SELECTED SUBPOPULATION
(11) RAS OF SELECTED SUBPOPULATION
(12) RATIO AF/AXL
(13) RATIO FWD/AXL
(14) RATIO RAS/AXL

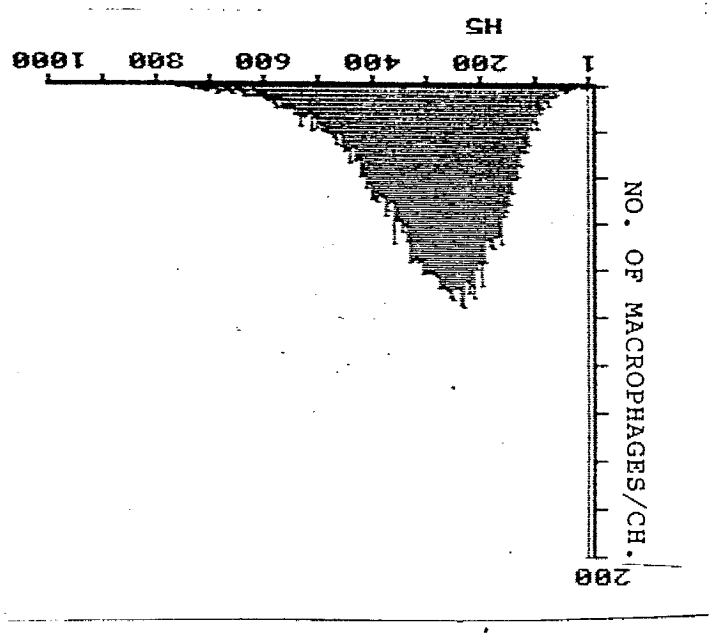
FIGURE

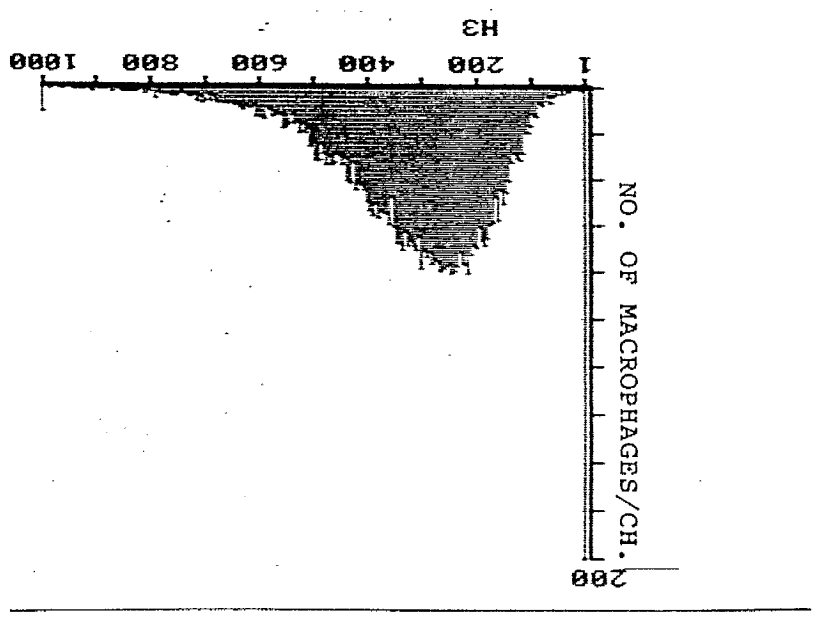
EVALUATION SCHEME FOR PARAMETERS FROM PHAGOCYTOSIS ASSAY

2.KW83

2029027544

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000





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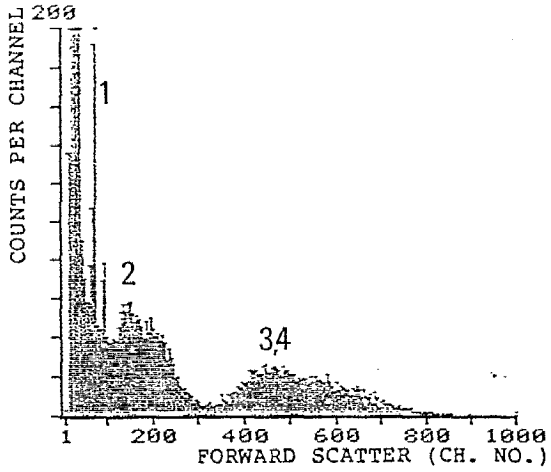

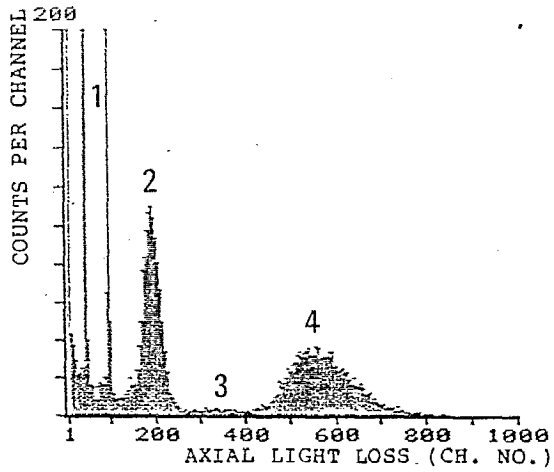

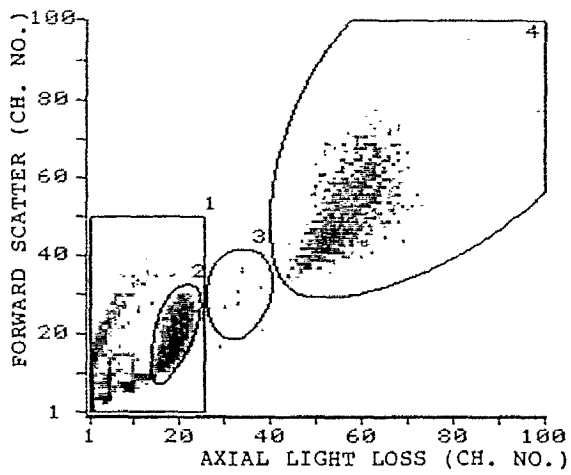

3C FIGURE

X8 CL012 P.0

20290275 46

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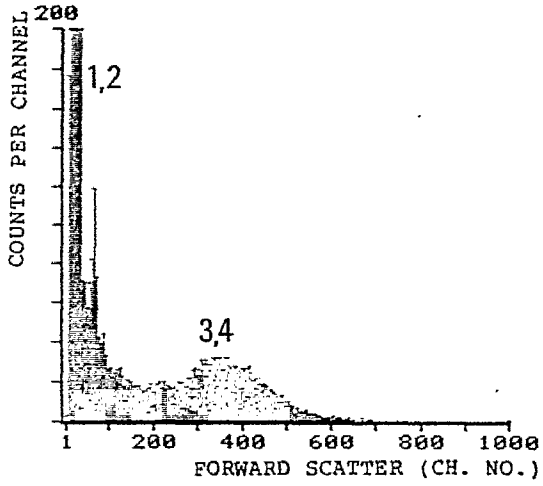

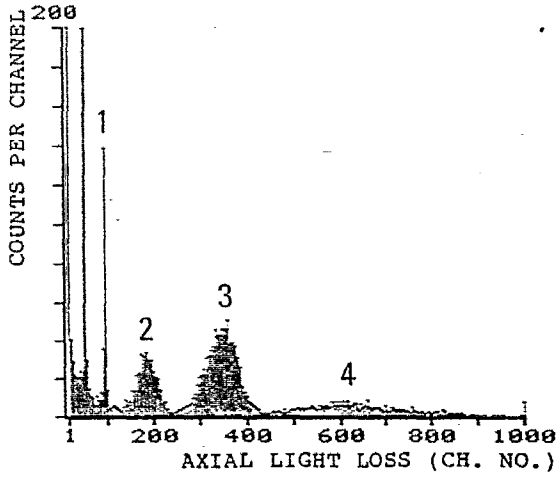

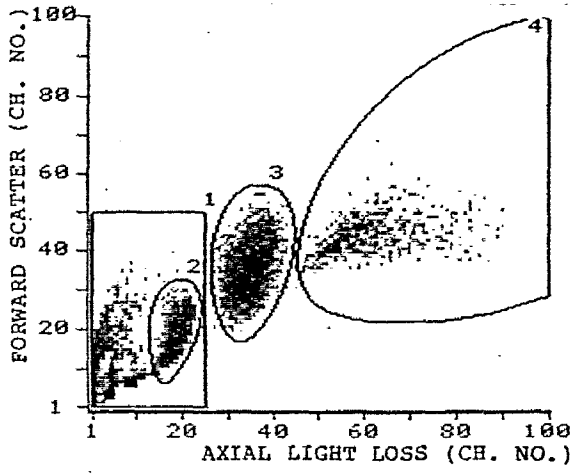

3C FIGURE

X 8 (L122P.0)

20290275A7

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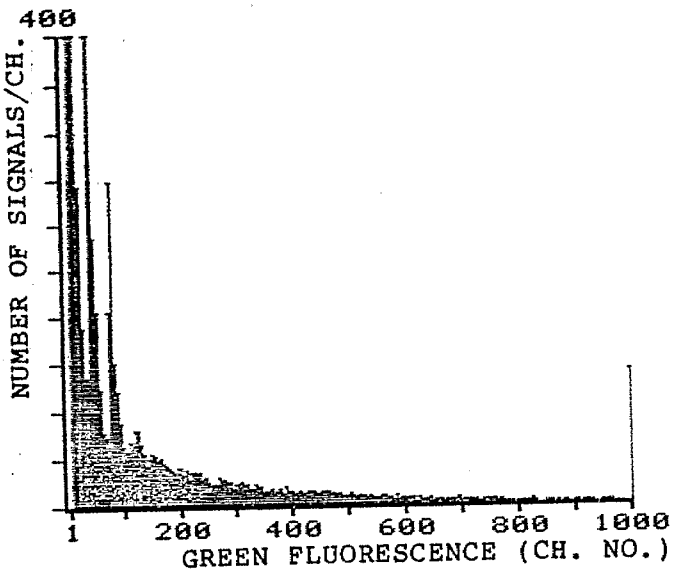

2029027548

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

2029027549

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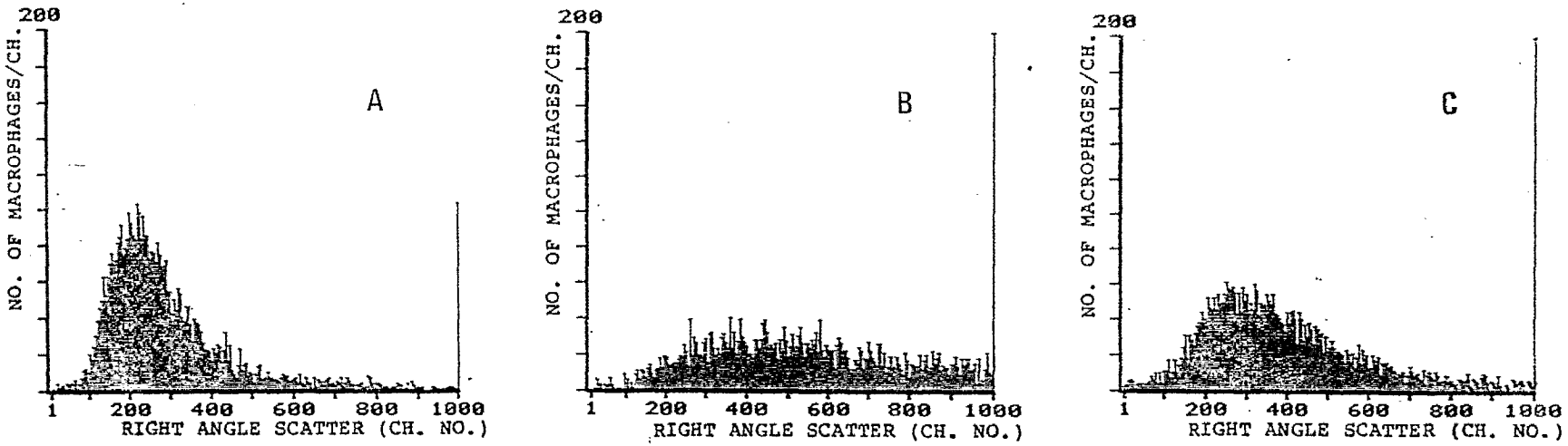
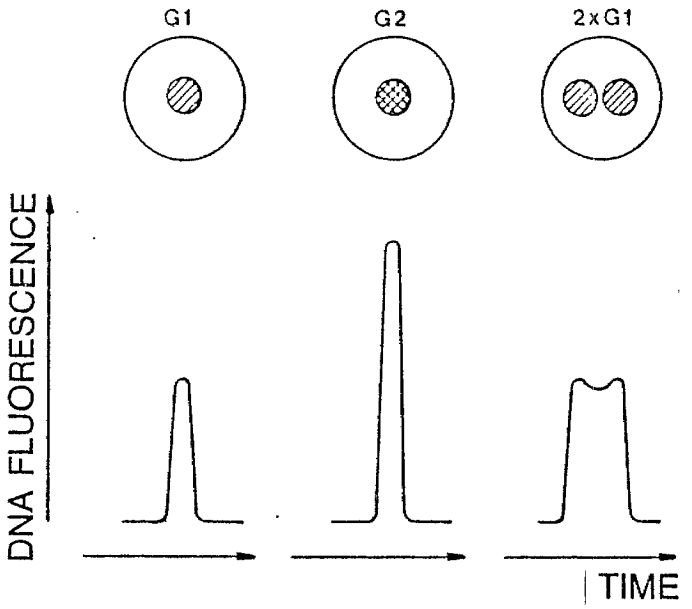


2029027550

X/13

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



DNA Signals from Mono- and Binuclear Cells

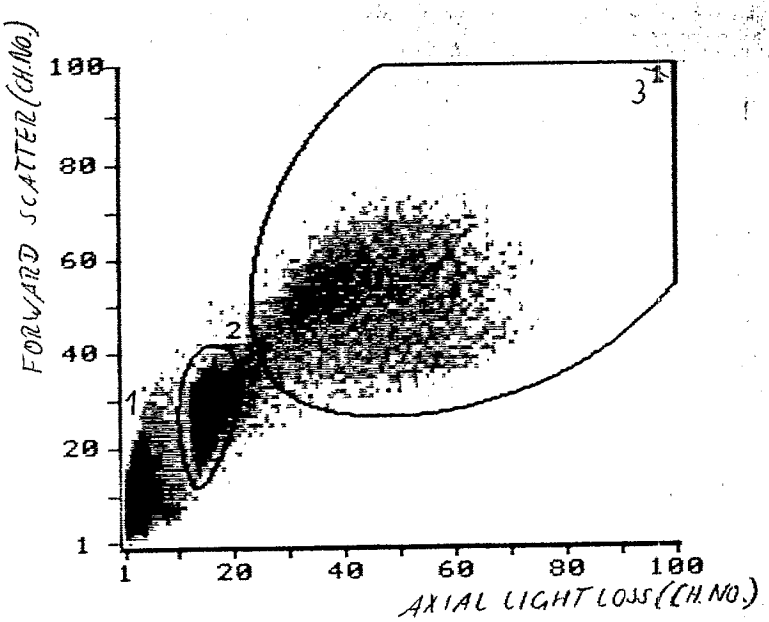
[X 14]

1x verkleinern, Original bei FHO

20290275S1

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

L122DR620



3C FIGURE [X15?]

CYTOGRAM OF TOTAL FORWARD SCATTER VERSUS AXIAL LIGHT LOSS SIGNALS FROM ETHANOL FIXED FLC, DNA ASSAY, EXAMPLE

Remarks: see 5 in 3C FIGURE

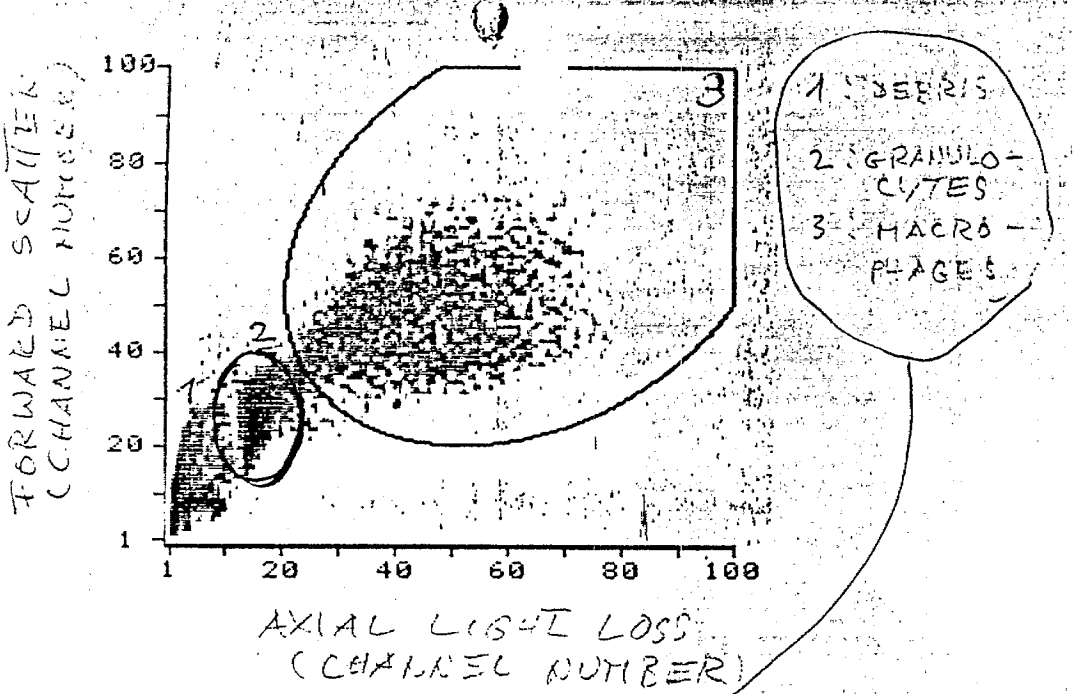
1: Debris
2: Granulocytes
3: Macrophages

20290275S2

Source: https://www.industrydocuments.ucsf.edu/docs/...

L-122 DR620

~~L-132 DR620~~



[X15?]

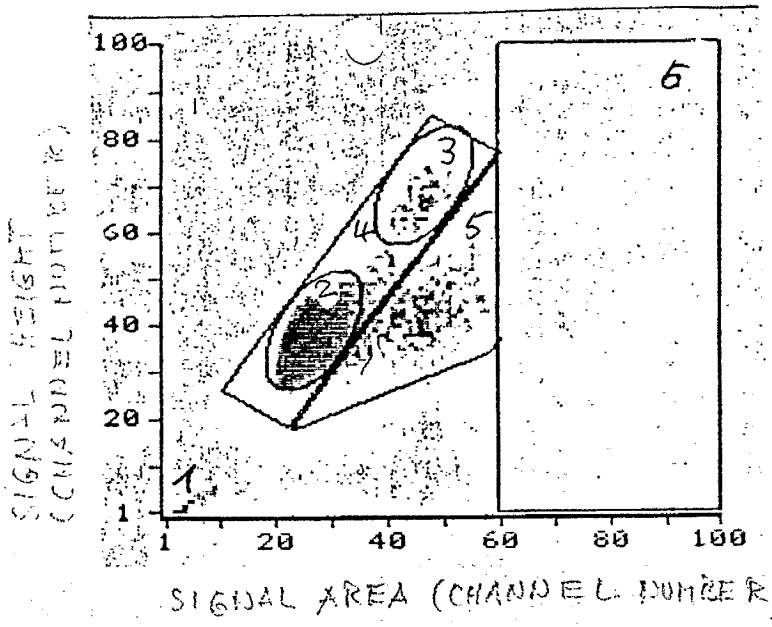
FIG 1

CYTOGRAM OF TOTAL FORWARD SCATTER VS AXIAL LIGHT LOSS SIGNALS FROM ETHANOL FIXED FLC, DNA ASSAY, EXAMPLE

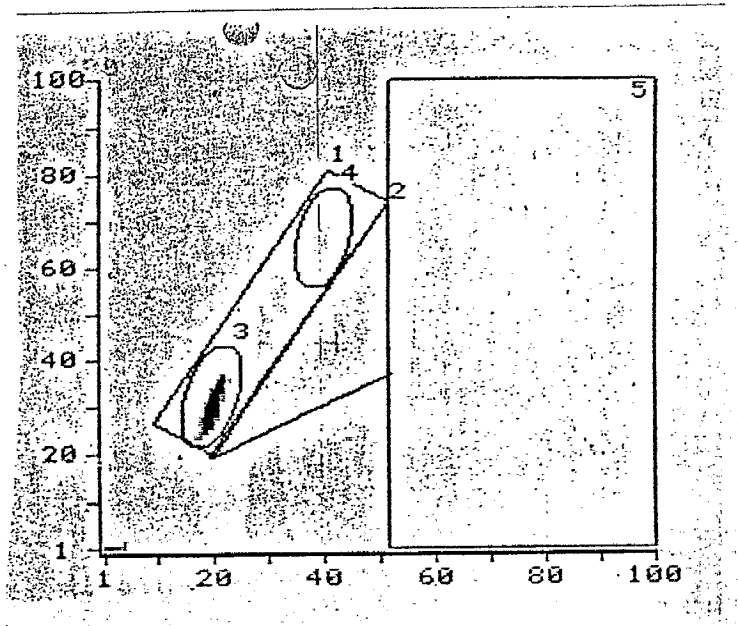
Remarks: See ~~box two~~ 5. in FIGURE

2029027553

Source: https://www.industrydocuments.ucsf.edu/docs/[illegible]



MACROPHAGES



GRANULOCYTES

3C FIGURE [X 18?]

CYTOGRAMS OF RED FLUORESCENCE (DNA) SIGNAL HEIGHT VS AREA FROM FLC, EXAMPLES

Remarks: see ~~box~~ no. 6 in FIGURE

1 = Debris
2 = G0/1
3 = G2
4 = ~~Cells in~~ S when not in windows 2 and 3
5 = 2xG0/1, binuclear cells and doublets
6 = GT. 2xG0/1, multinuclear cells and ~~cell clump~~ agg

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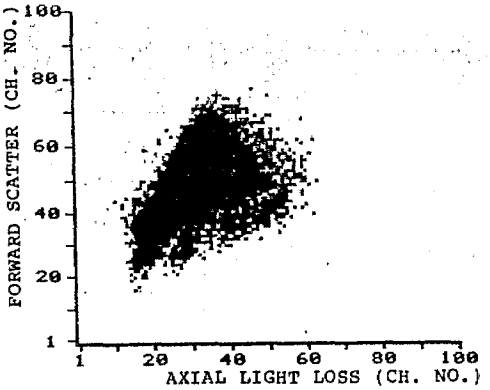
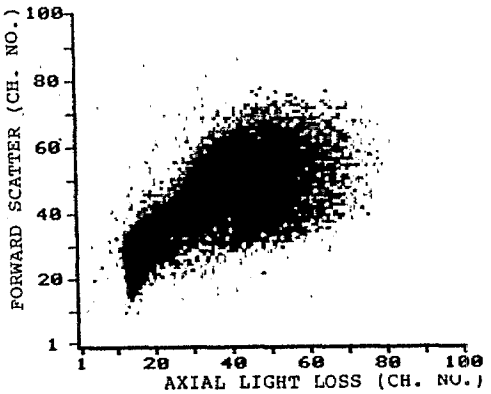
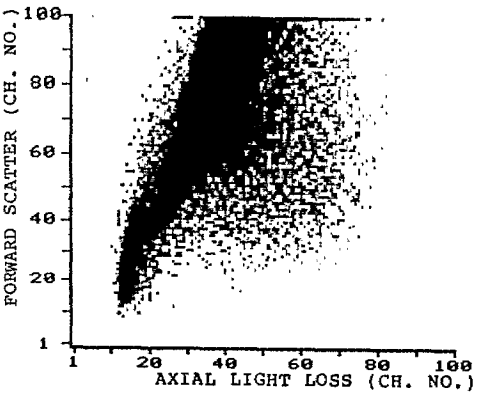
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000







C032DR020-C233JN520

G01A

[X 19?]

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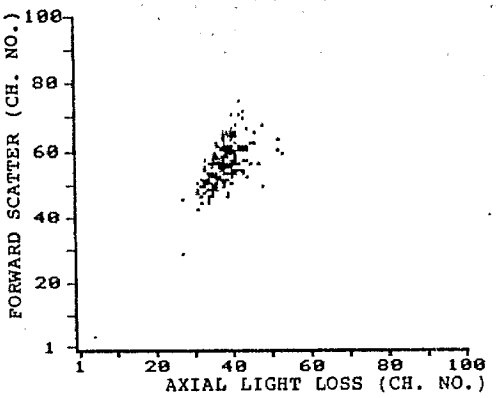
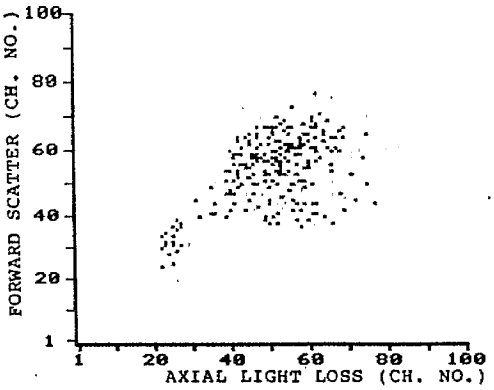
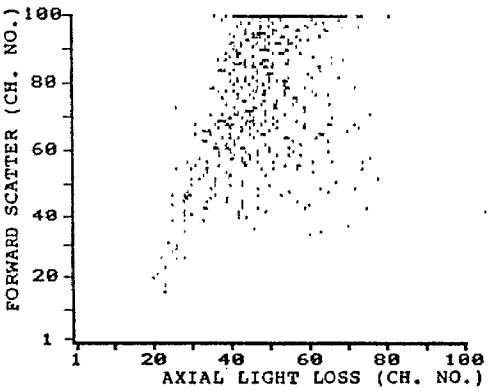


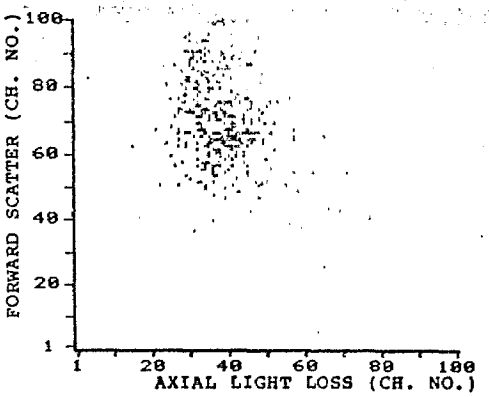


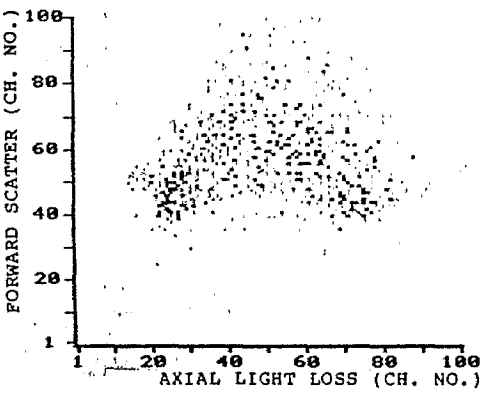


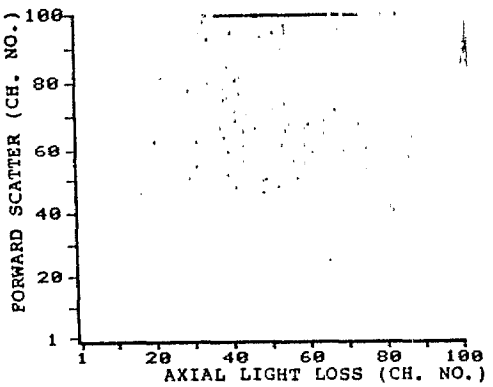
Lo32 DRG20 - L263 DR620 ← 62

20290275556

Source: https://www.industrydocuments.ucsf.edu/docs/[illegible]

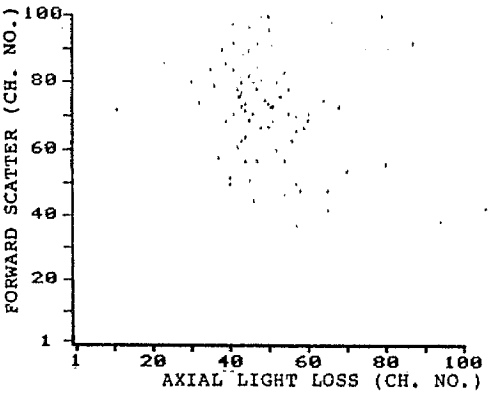


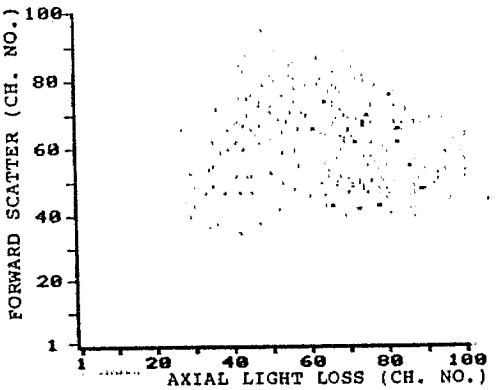


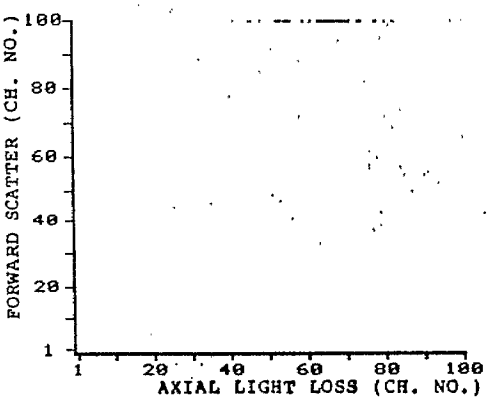


2029027557

Source: https://www.industrydocuments.ucsf.edu/doc/

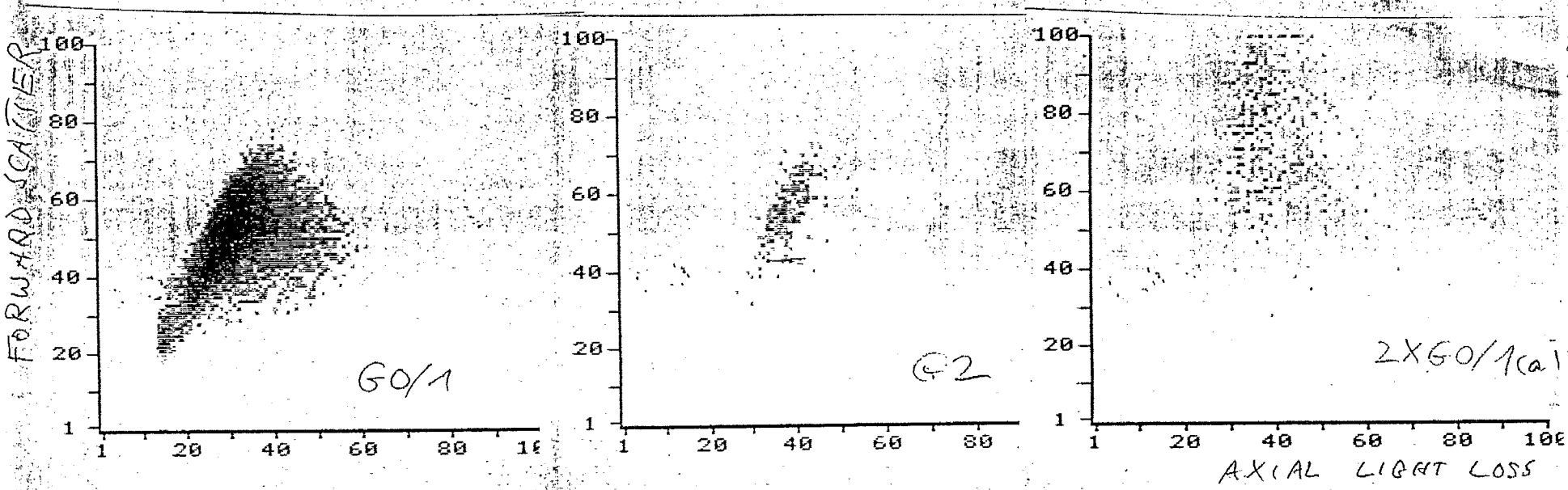






2029027558

Source: https://www.industrydocuments.ucsf.edu/docs/[illegible]0000



BC FIGURE [X 192]

CYTOGRAMS OF FORWARD SCATTER VS AXIAL LIGHT LOSS SIGNALS FROM MACROPHAGES, EXAMPLES (MONO- AND BINUCLEAR

Regions: see box 7. in FIGURE (a) signals enhanced by

20290277559

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

X 79

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

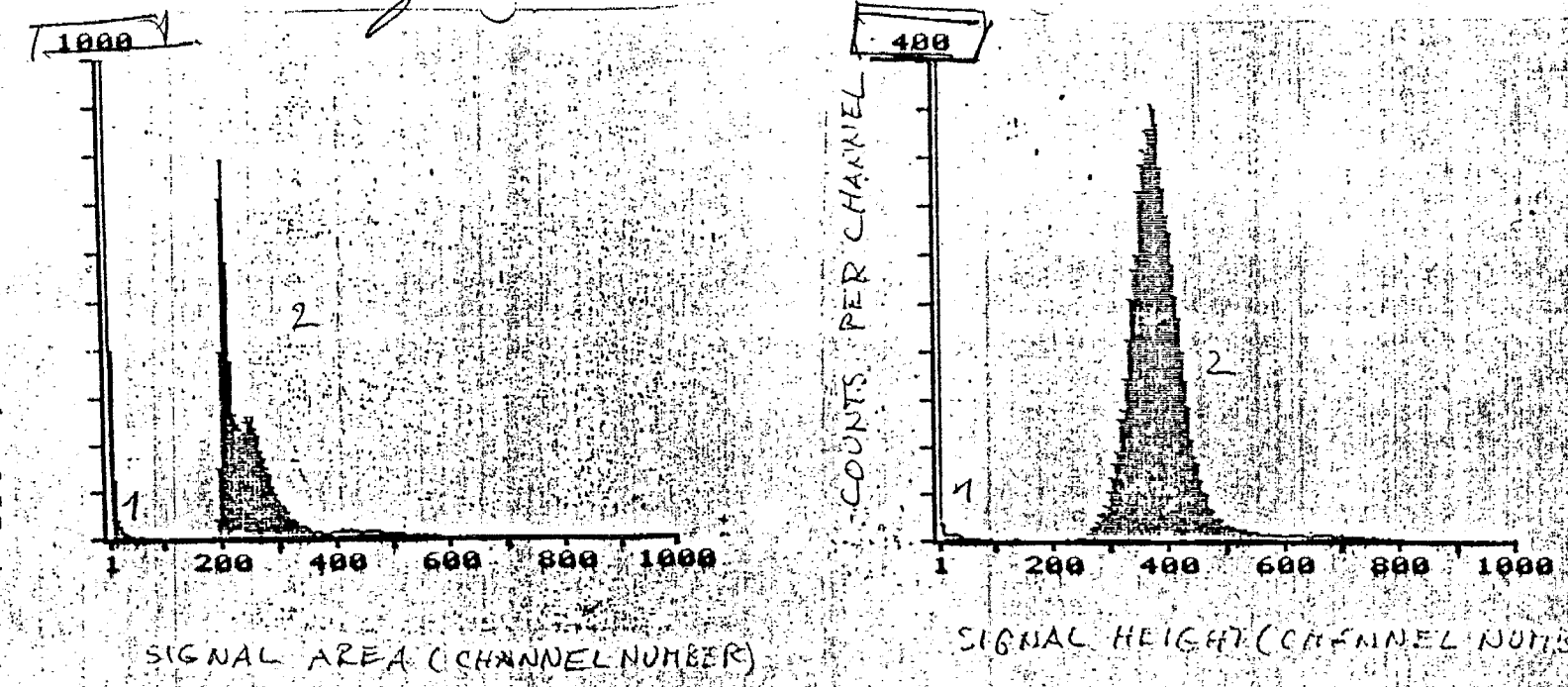


FIGURE 1

HISTOGRAMS OF TOTAL REDFLUORESCENCE (DNA) HEIGHT AND AREA SIGNALS FROM FLC EXAMPLES

Remarks: see ~~box no~~ Band 4 ✓ in FIGURE

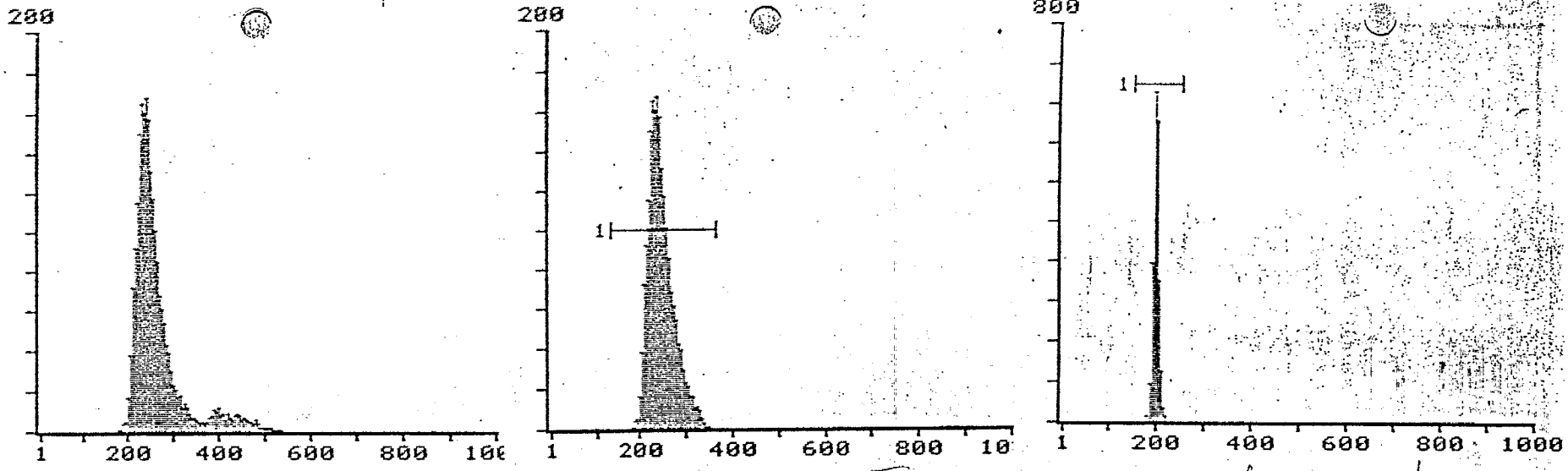
1 : Debris

2 : DNA SIGNALS FROM FLC

19

Source: https://www.industrydocuments.ucsf.edu/docs/2029027561

2029027561



[desgl. signal height, ms, area, PMS height, PMS]

BC FIGURE

HISTOGRAMS OF RED FLUORESCENCE (DNA) SIGNAL AREA DISTRIBUTION OF ~~MACROPHAGES~~, EXAMPLES

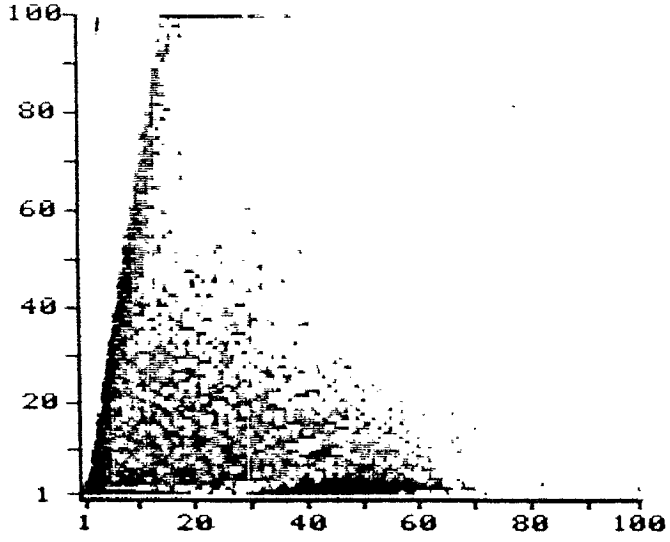
Remarks: see box nb. 8 in FIGURE
A : all signals, not gated for DNA (~~box~~ no. 6 in FIGURE and FIGURE)

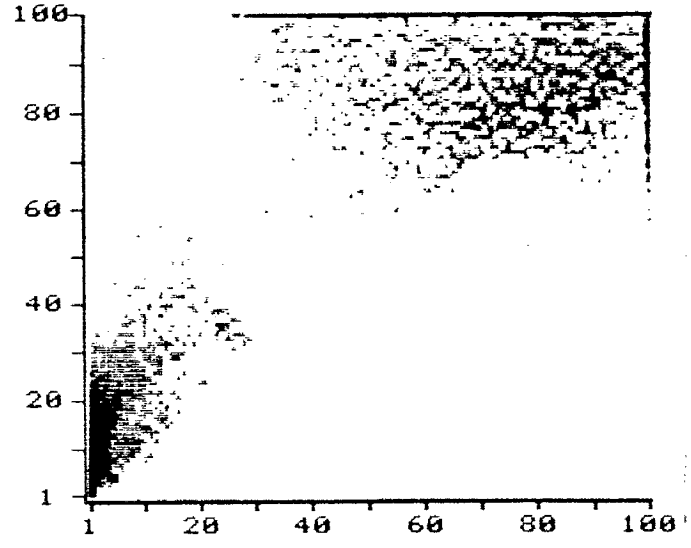
B = G0/1 macrophages, gated by window no. 2 (see FIGURE)

[1 = window used for evaluation of peak position and CV]

2029027562

Source: https://www.industrydocuments.ucsf.edu/docs/ymdl0000





[21]

2029027563

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

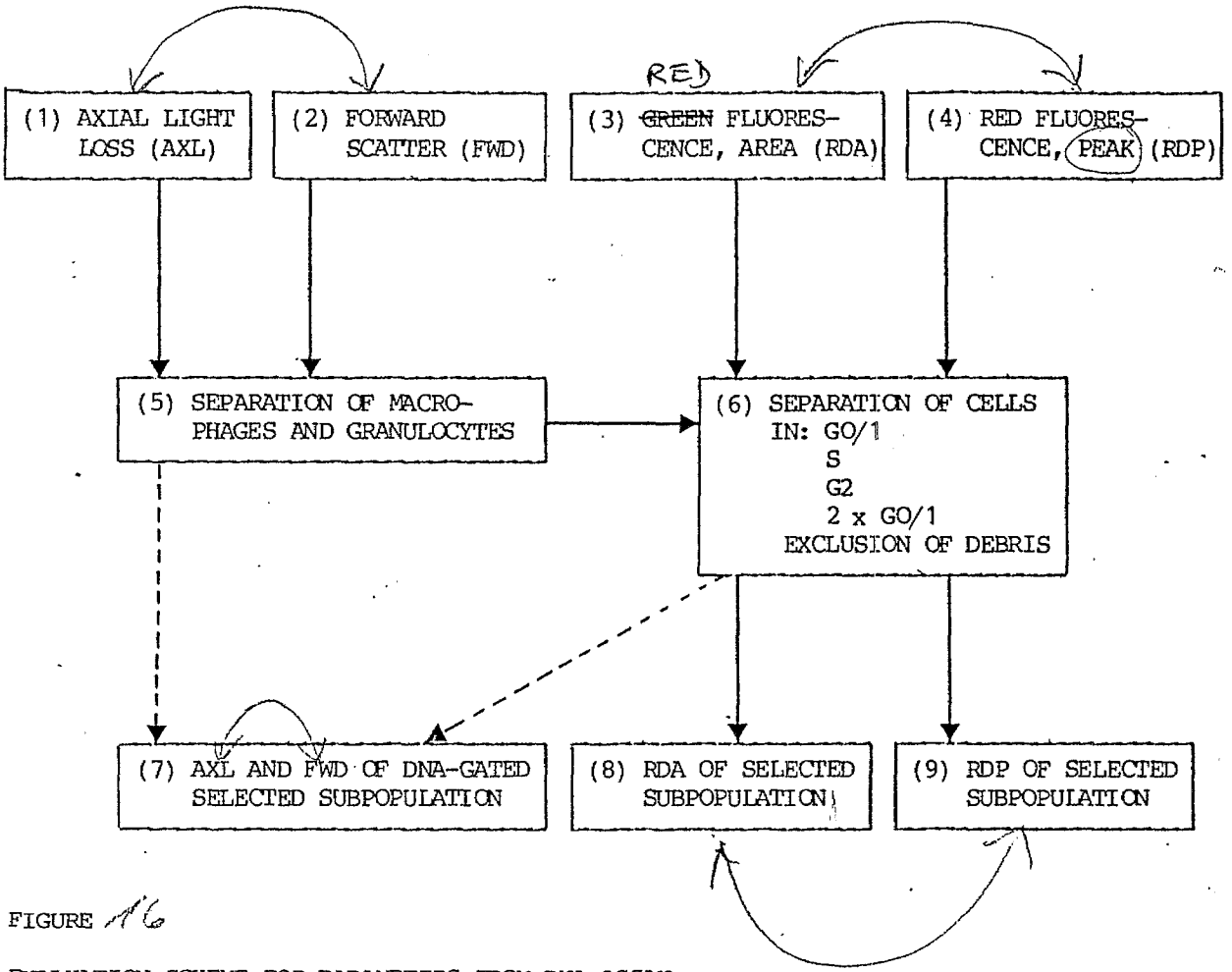

(1) AXIAL LIGHT LOSS (AXL)
(2) FORWARD SCATTER (FWD)
RED
(3) ~~GREEN~~ FLUORESCENCE, AREA (RDA)
(4) RED FLUORESCENCE, PEAK (RDP)
(5) SEPARATION OF MACROPHAGES AND GRANULOCYTES
(6) SEPARATION OF CELLS IN: GO/1
S
G2
2 x GO/1
EXCLUSION OF DEBRIS
(7) AXL AND FWD OF DNA-GATED SELECTED SUBPOPULATION
(8) RDA OF SELECTED SUBPOPULATION
(9) RDP OF SELECTED SUBPOPULATION

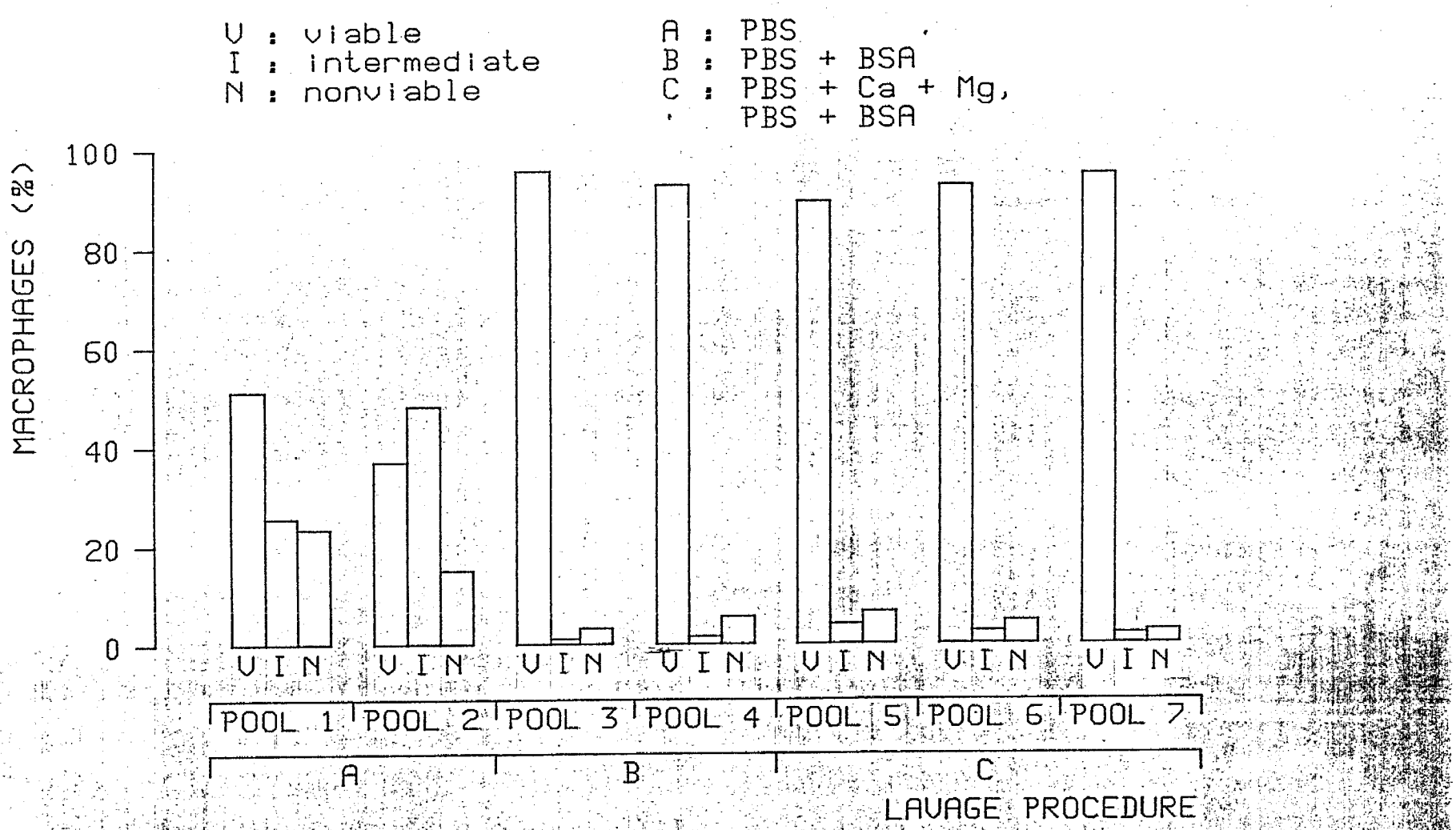
FIGURE 16

EVALUATION SCHEME FOR PARAMETERS FROM DNA ASSAY

2029027564

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03356, TH, U3 F162 U2 U01 O-GR, d 1



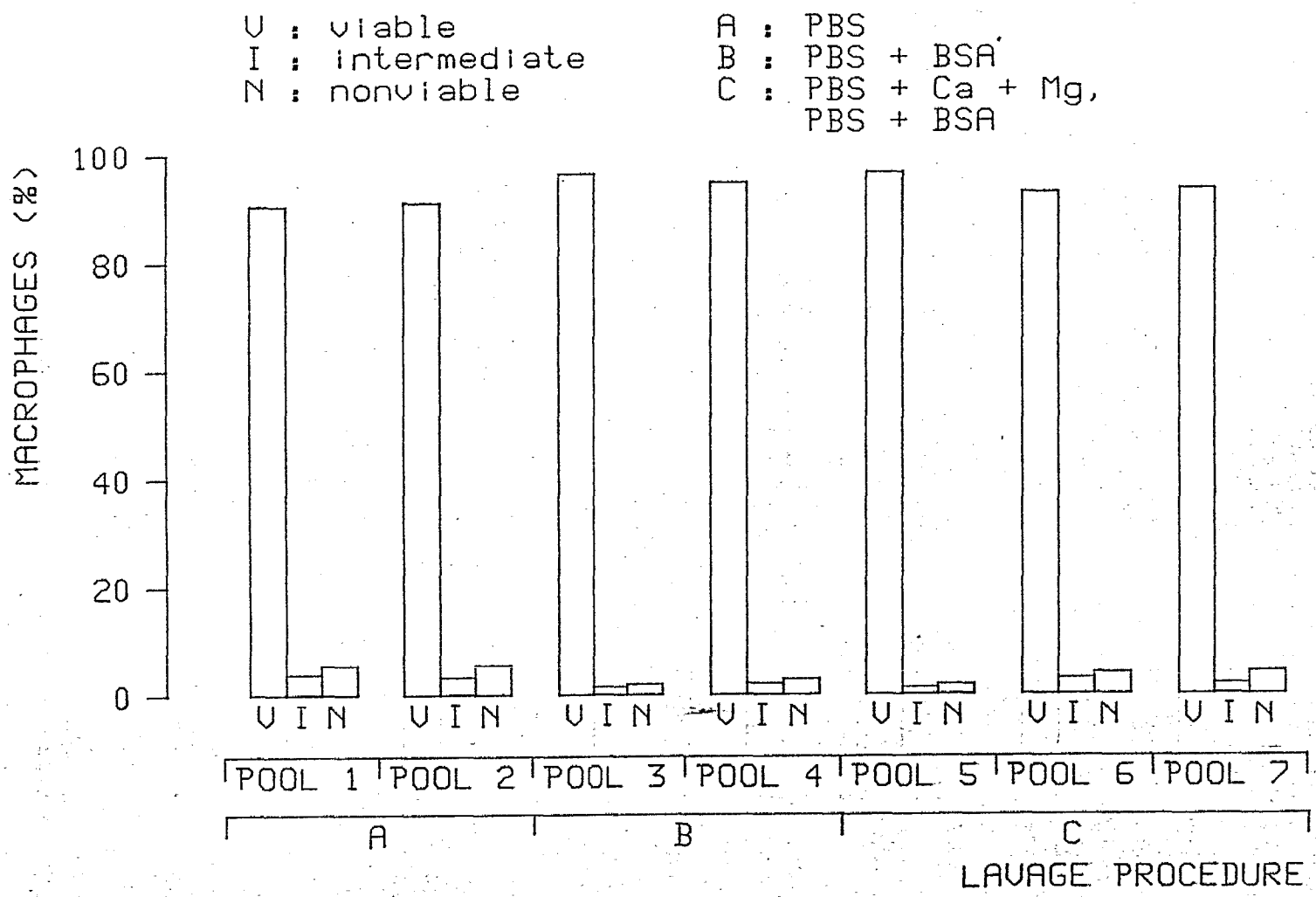
BC FIGURE
VIABILITY OF MACROPHAGES DETERMINED BY FCM, O-GR, EXPERIMENT 1

20290275&5

Remarks: for details see BC TABLE 54

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03524, V24 F162 V4 V03



BC FIGURE

VIABILITY OF MACROPHAGES DETERMINED BY FCM, O-GR, EXPERIMENT 3

20290225 66

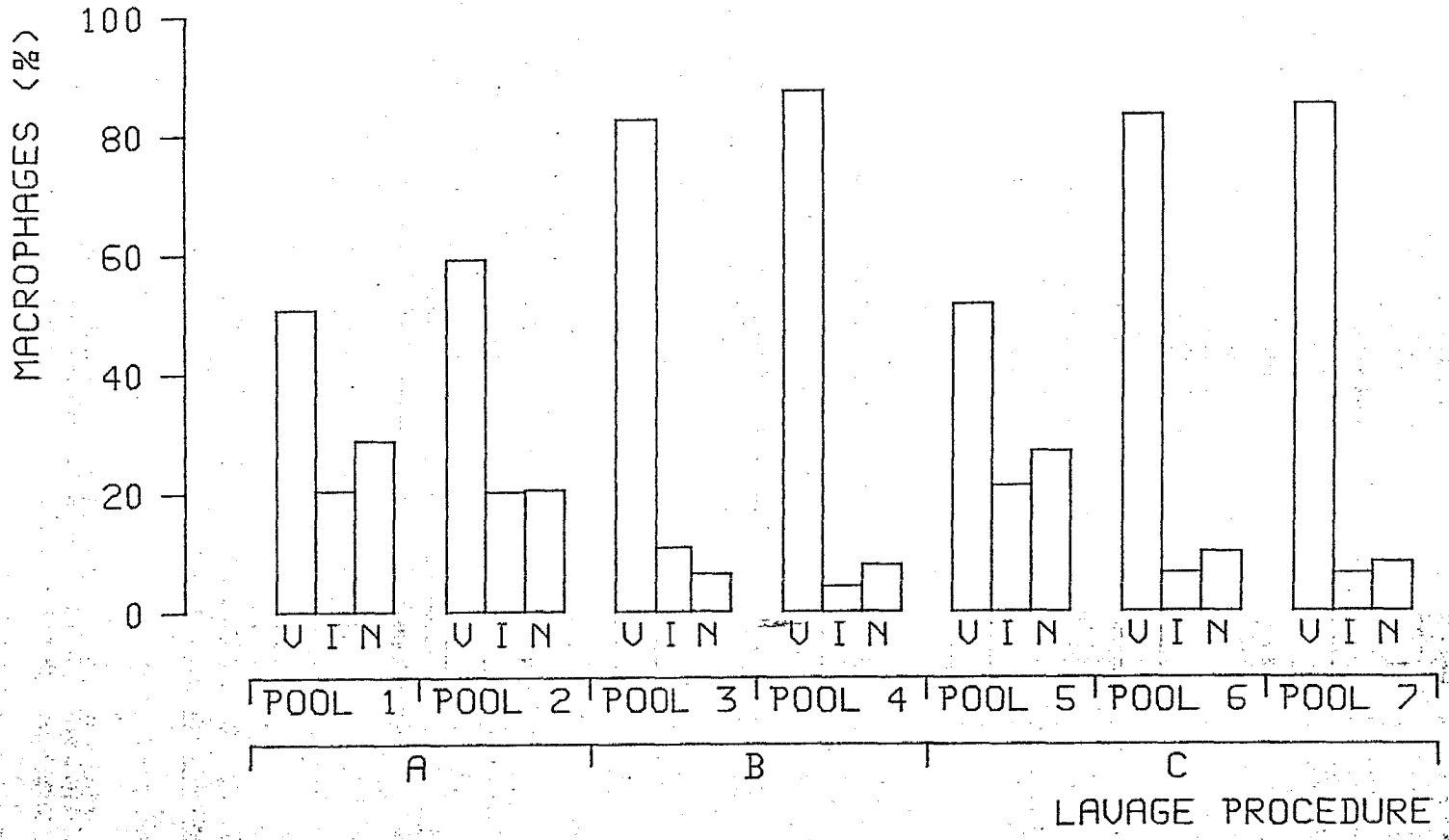
Remarks: for details see BC TABLE 54

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03525, U5 F162 U3, U11 1-GR d-1

V : viable
I : intermediate
N : nonviable

A : PBS
B : PBS + BSA
C : PBS + Ca + Mg, PBS + BSA



3C FIGURE

VIABILITY OF MACROPHAGES DETERMINED BY FCM, 1-GR, EXPERIMENT 1

20290275?

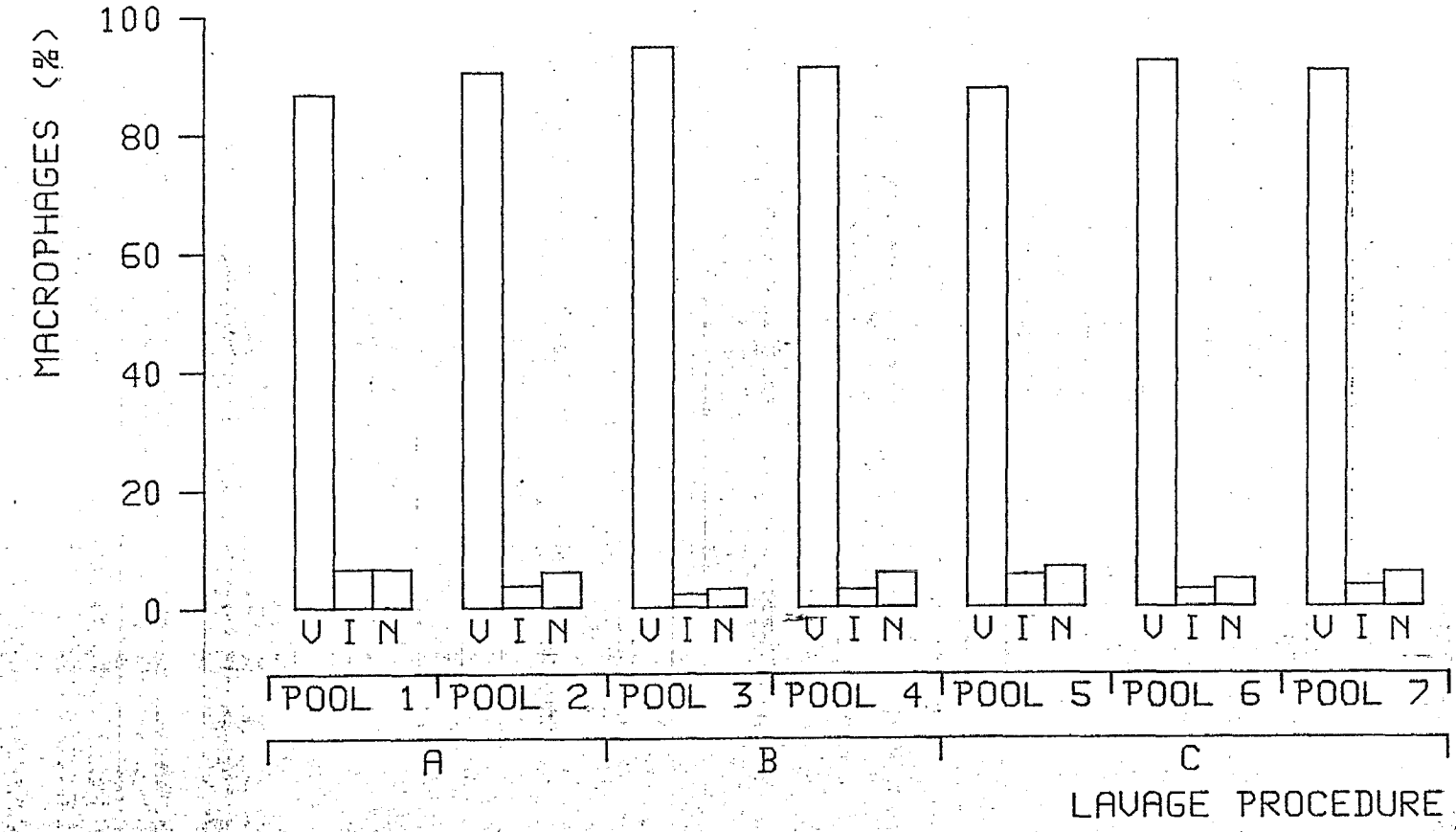
Remarks: for details see 3C TABLE 54

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03526, V6 F162 V5, V13 1GR d3

V : viable
I : intermediate
N : nonviable

A : PBS
B : PBS + BSA
C : PBS + Ca + Mg, PBS + BSA



BC FIGURE

Viability of macrophages determined by FCM, 1-GR, Experiment 3

20290275 68

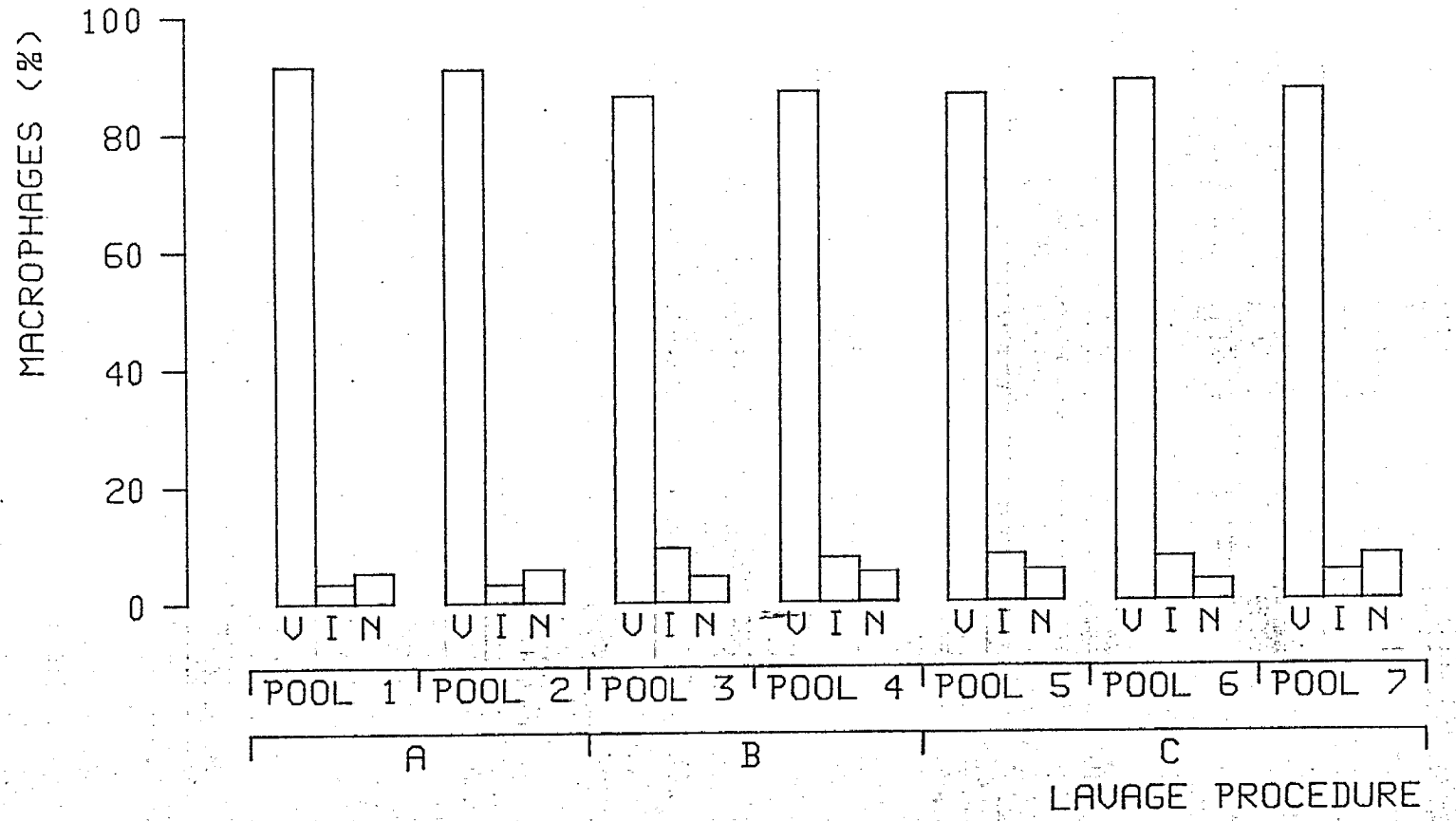
Remarks: for details see BC TABLE 54

Source: https://www.industrydocuments.ucsf.edu/docs/yhdl0000

H 0500/3057, H03527, U7 F162 U6, U23 2-GR d3

V : viable
I : intermediate
N : nonviable

A : PBS
B : PBS + BSA
C : PBS + Ca + Mg, PBS + BSA



BC FIGURE

Viability of macrophages determined by FCM, 2-GR, Experiment 3

Remarks: for details see BC TABLE 54

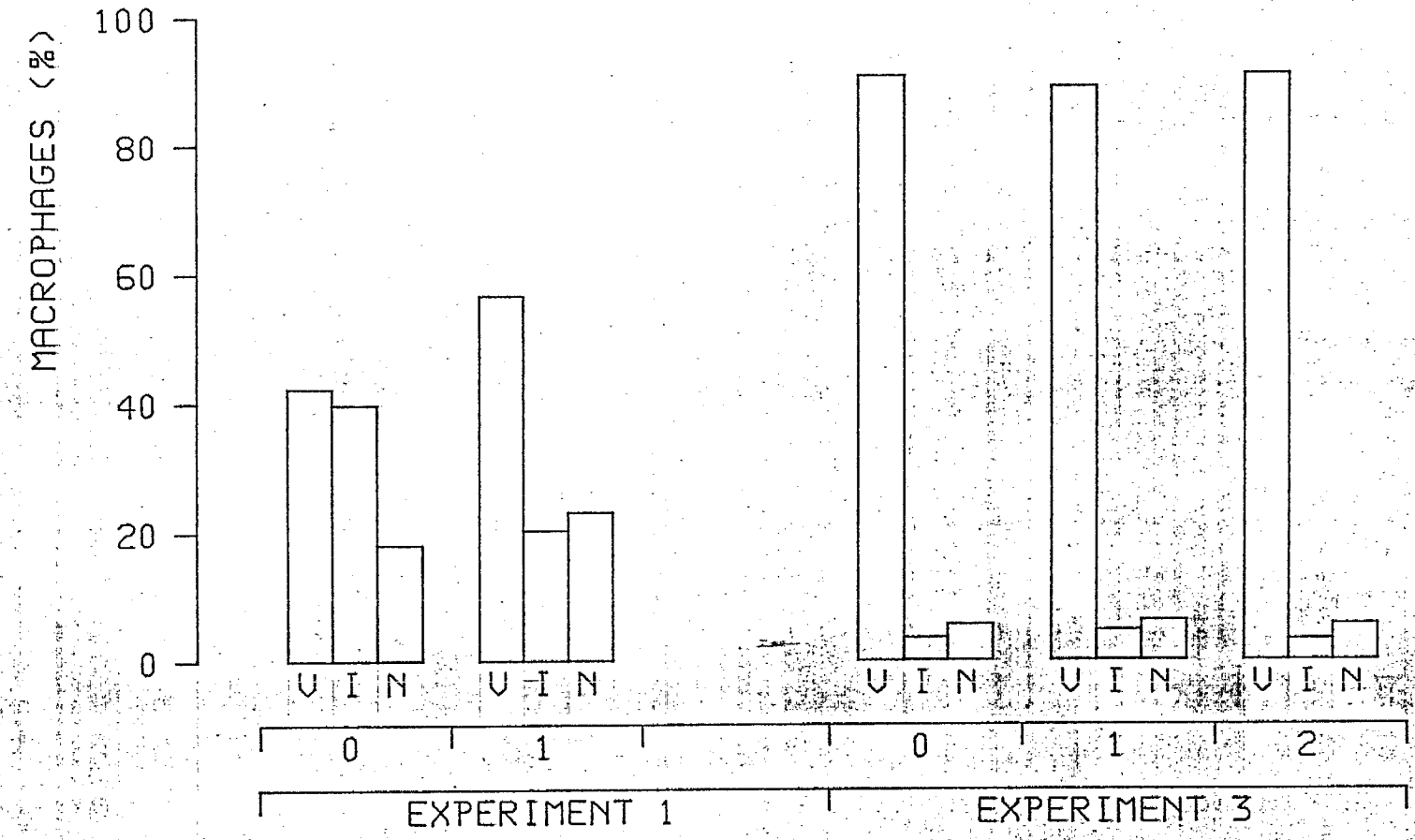
20290275 69

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03355, TH, V18 F162 V17 VWA P 1+2

V : viable
I : intermediate
N : nonviable

0 : 0-GR
1 : 1-GR
2 : 2-GR



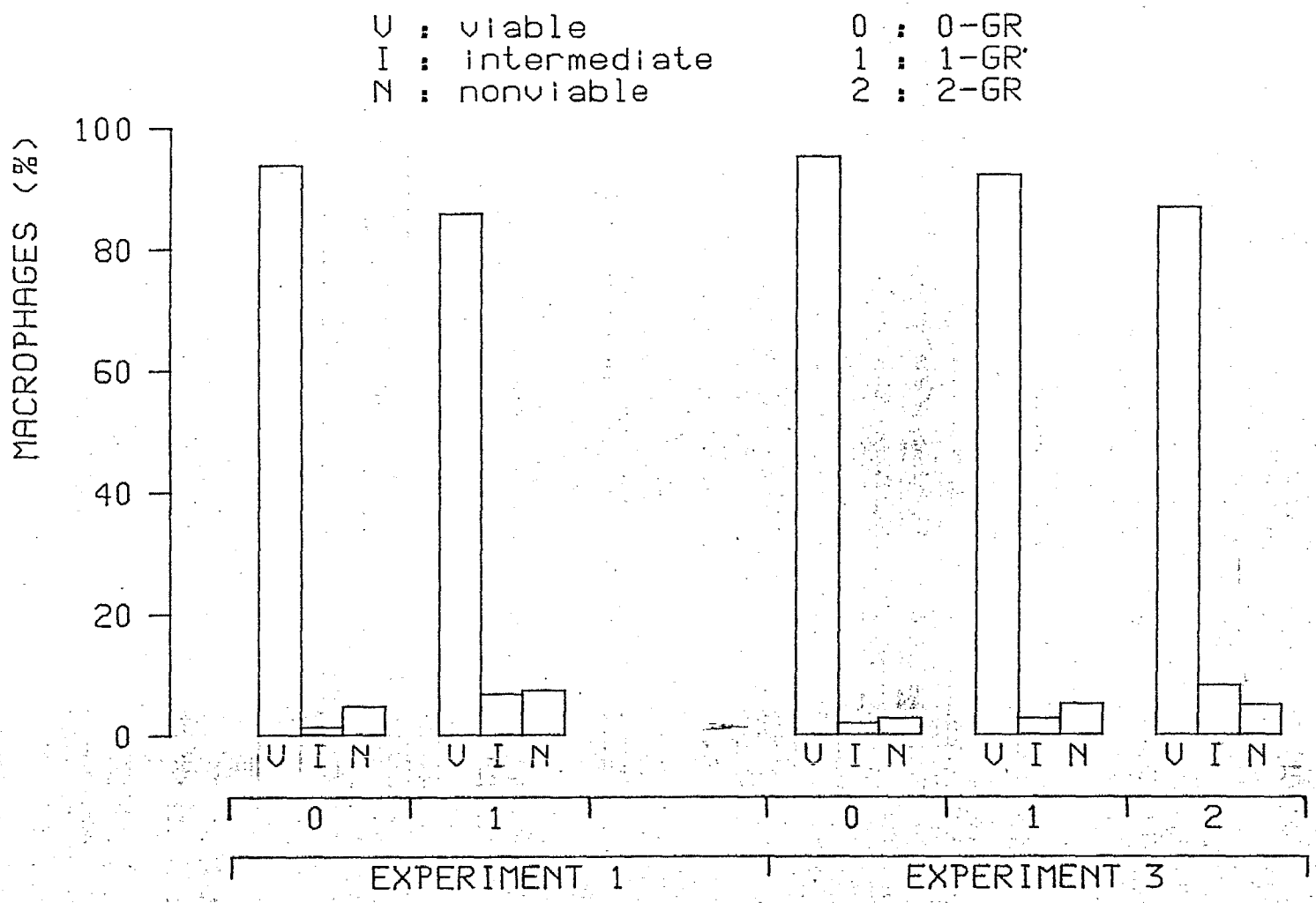
3C FIGURE

Relative number of viable, intermediate and nonviable macrophages, weighted mean of pool 1 and 2

Remarks: for details see 3C TABLE 55

20290227570

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



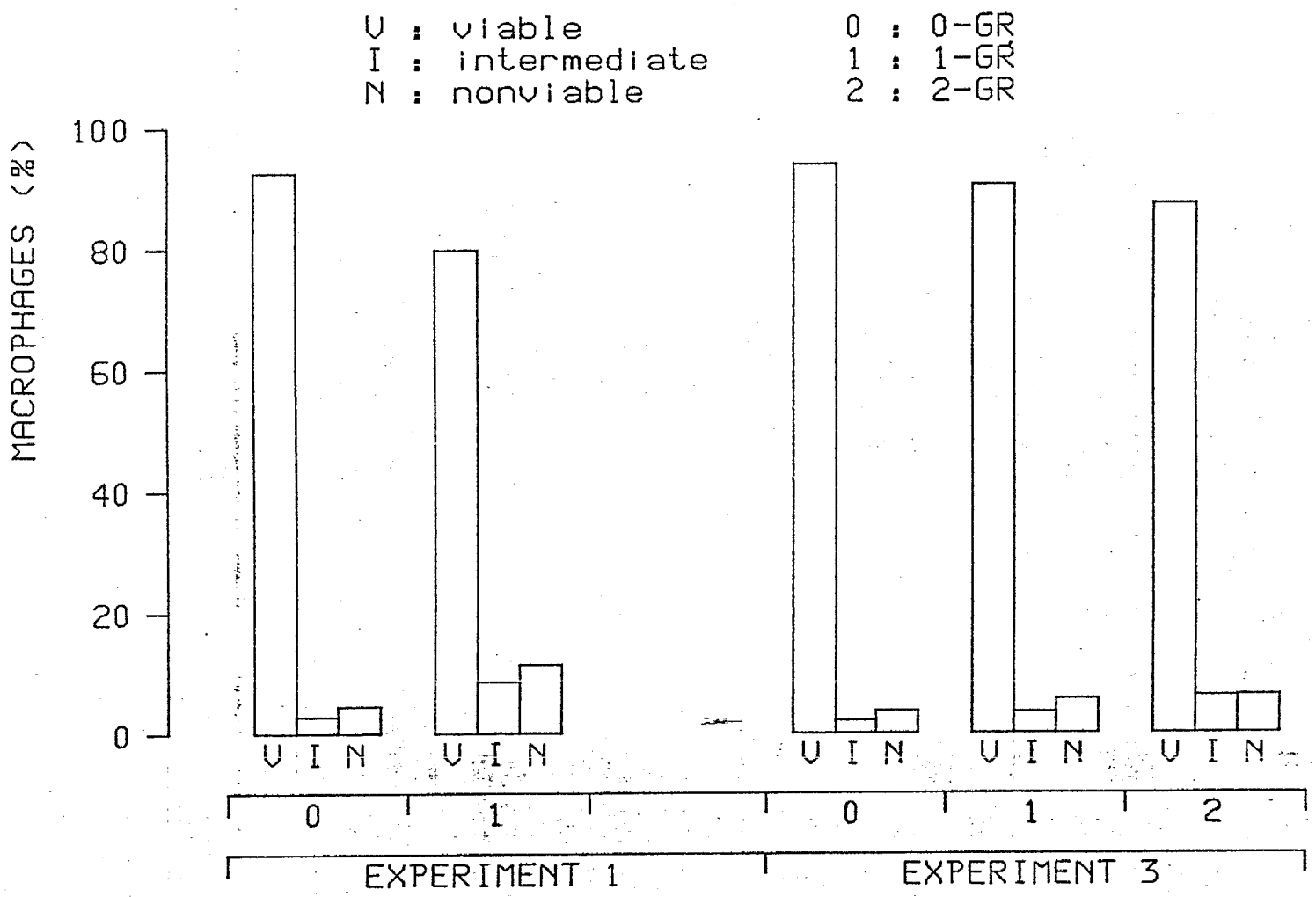
BC FIGURE

Relative number of viable, intermediate and nonviable macrophages, weighted mean of pool 3 and 4

2029027571 for details see BC TABLE 55

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03617, V21 F162 V20, VWC P5,6+7



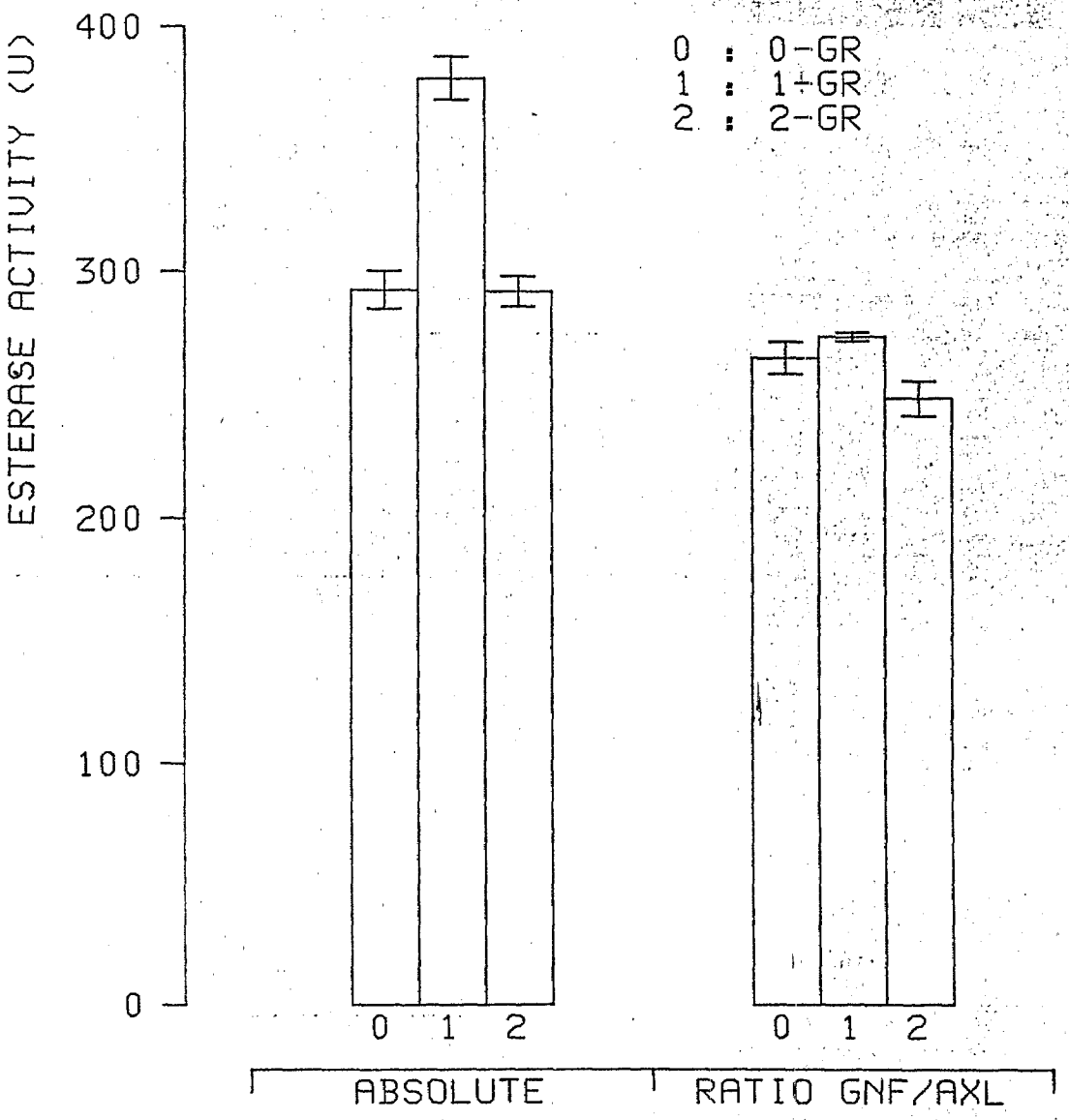
BC FIGURE

Relative number of viable, intermediate and nonviable macrophages, weighted mean of pool 5, 6 and 7

Remarks: for details see BC TABLE 55

20290275?2

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE

Esterase activity of macrophages, weighted mean of pools, Experiment 3, statistical parameters

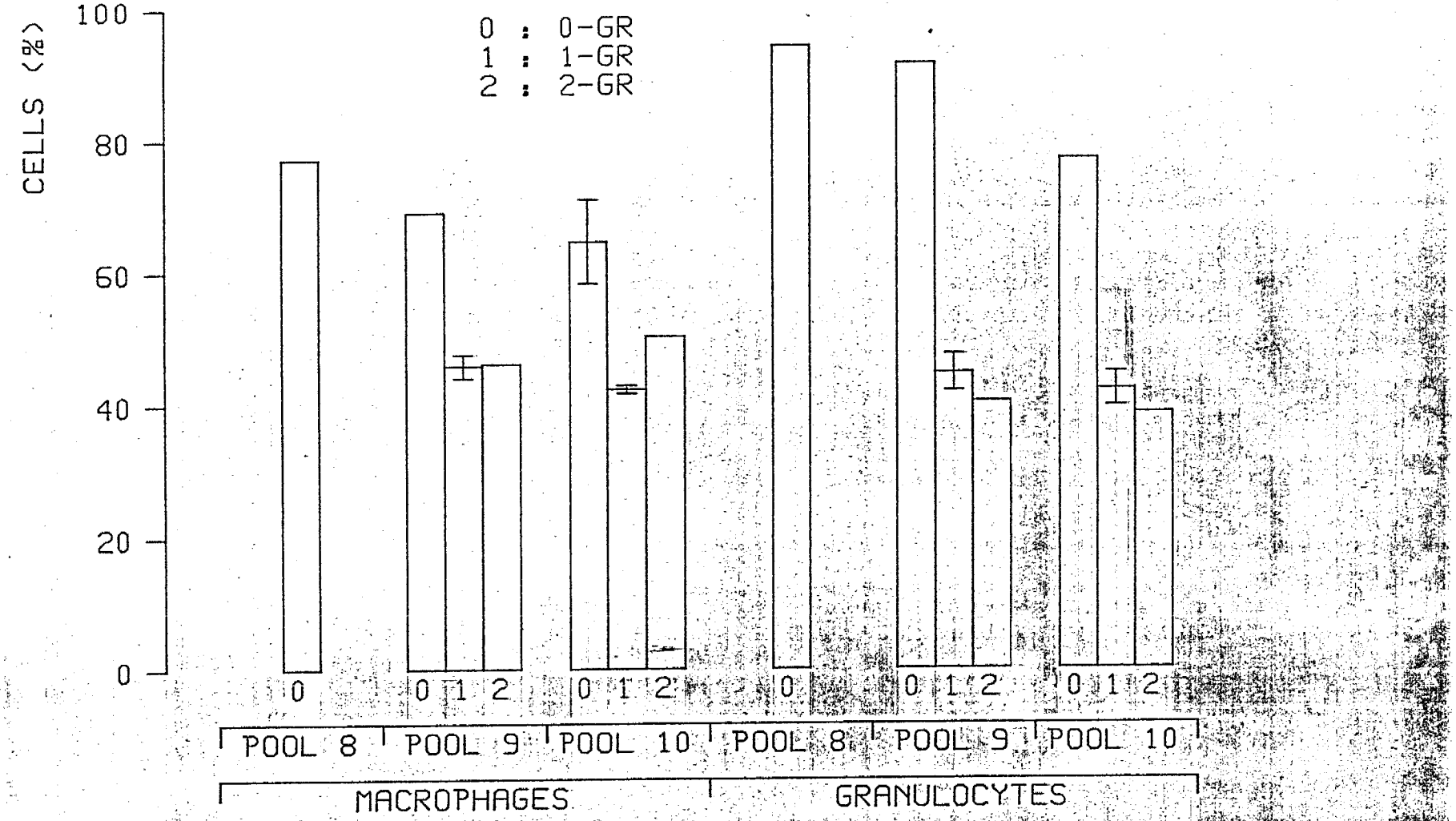
Remarks: for details see BC TABLE 59

A 0500/3057, H03357, TH, V145 F155 YFS EAW mean

20290275?3

Source: https://www.industrydocuments.ucsf.edu/docs/[illegible]

A 0500/3057, H03353, TH, V142 F155 V60 CNPhag



BC FIGURE

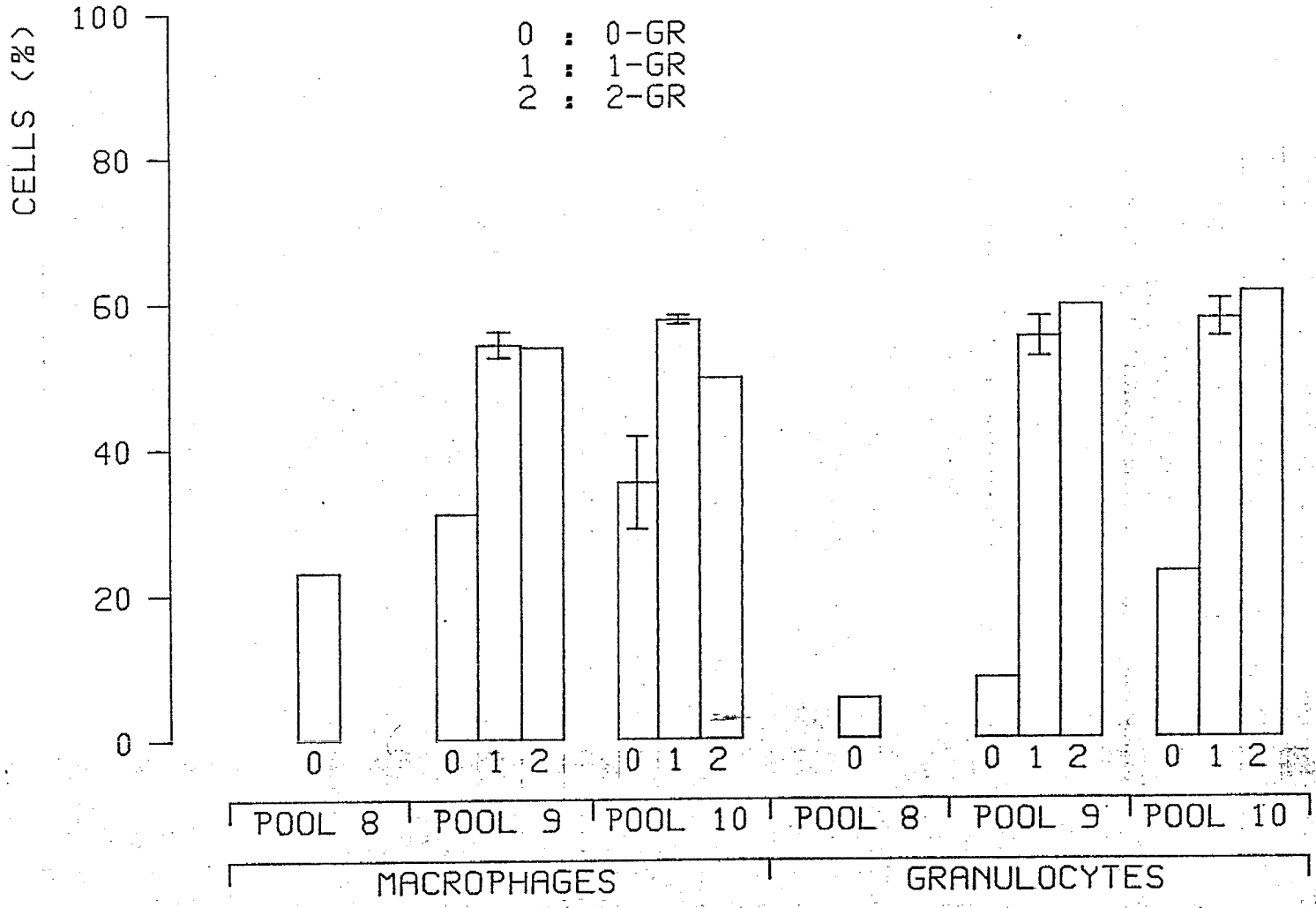
Relative number of nonphagocytising macrophages and granulocytes, statistical parameters

Remarks: for details see BC TABLE 63 and 69

2029027574

Source: https://www.industrydocuments.ucsf.edu/docs/ynnl0000

A 0500/3057, H03513, V144 F155 V143 CGE 1



BC FIGURE

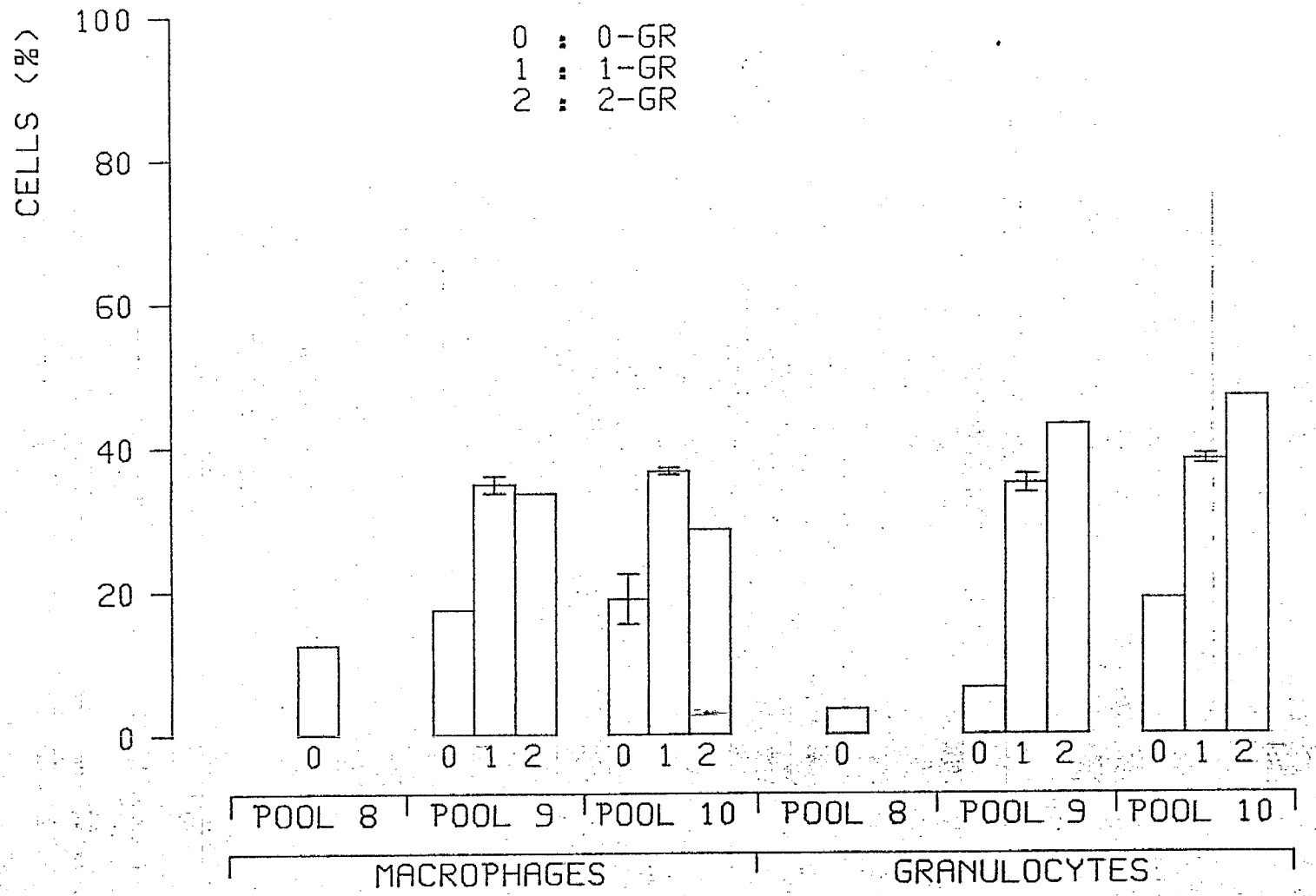
Relative number of macrophages and granulocytes phagocytising .GE. 1 bead, statistical parameters

20290275?5

Remarks: for details see BC TABLE 64 and 70

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03512, V25 F162 VXX CGE 3



BC FIGURE

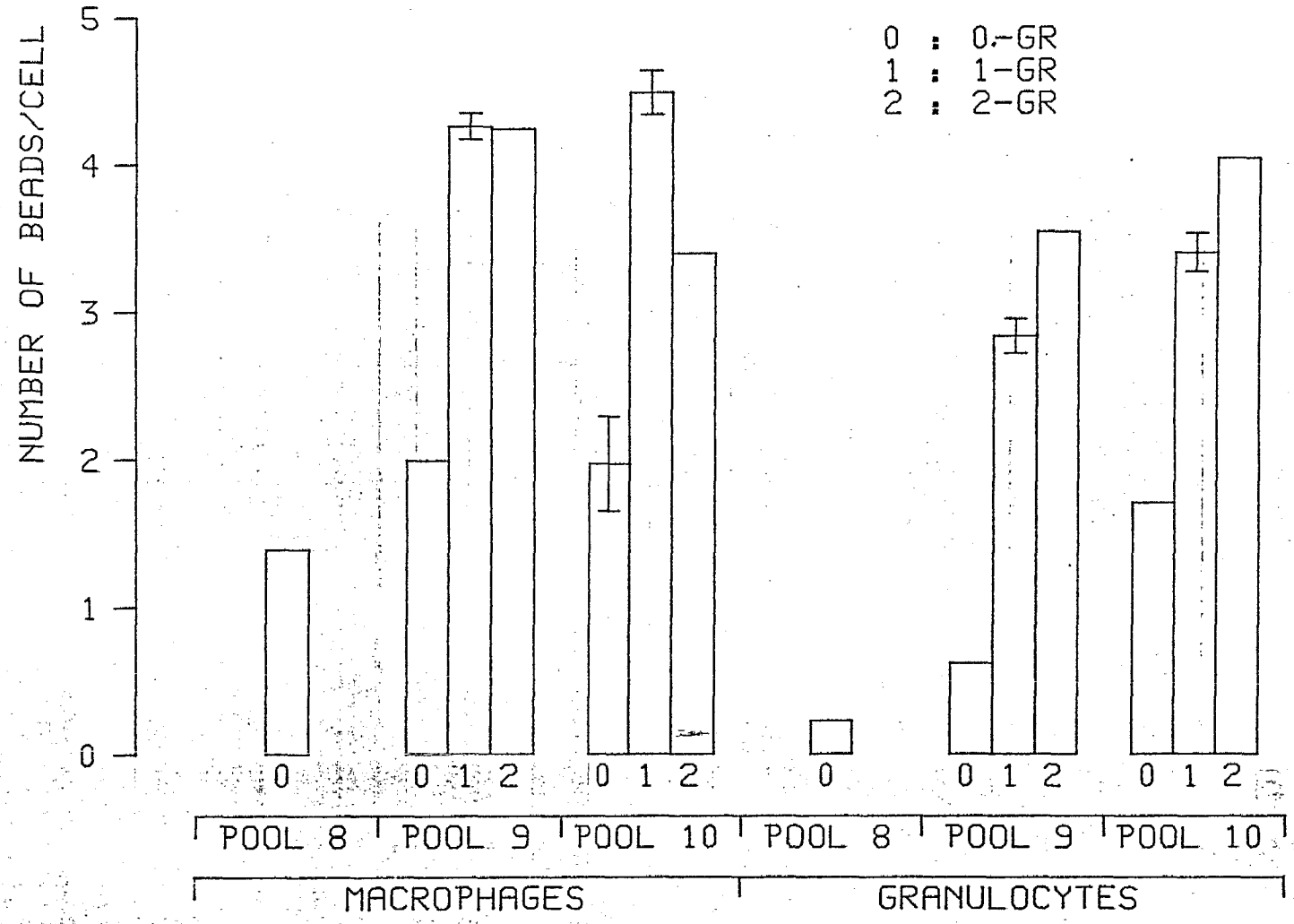
Relative number of macrophages and granulocyte phagocytising. GE. 3 beads, statistical parameters

20290275?6

Remarks: for details see BC TABLE 65 and 71

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03352, TH, V1 F162 VXX BPC



BC FIGURE

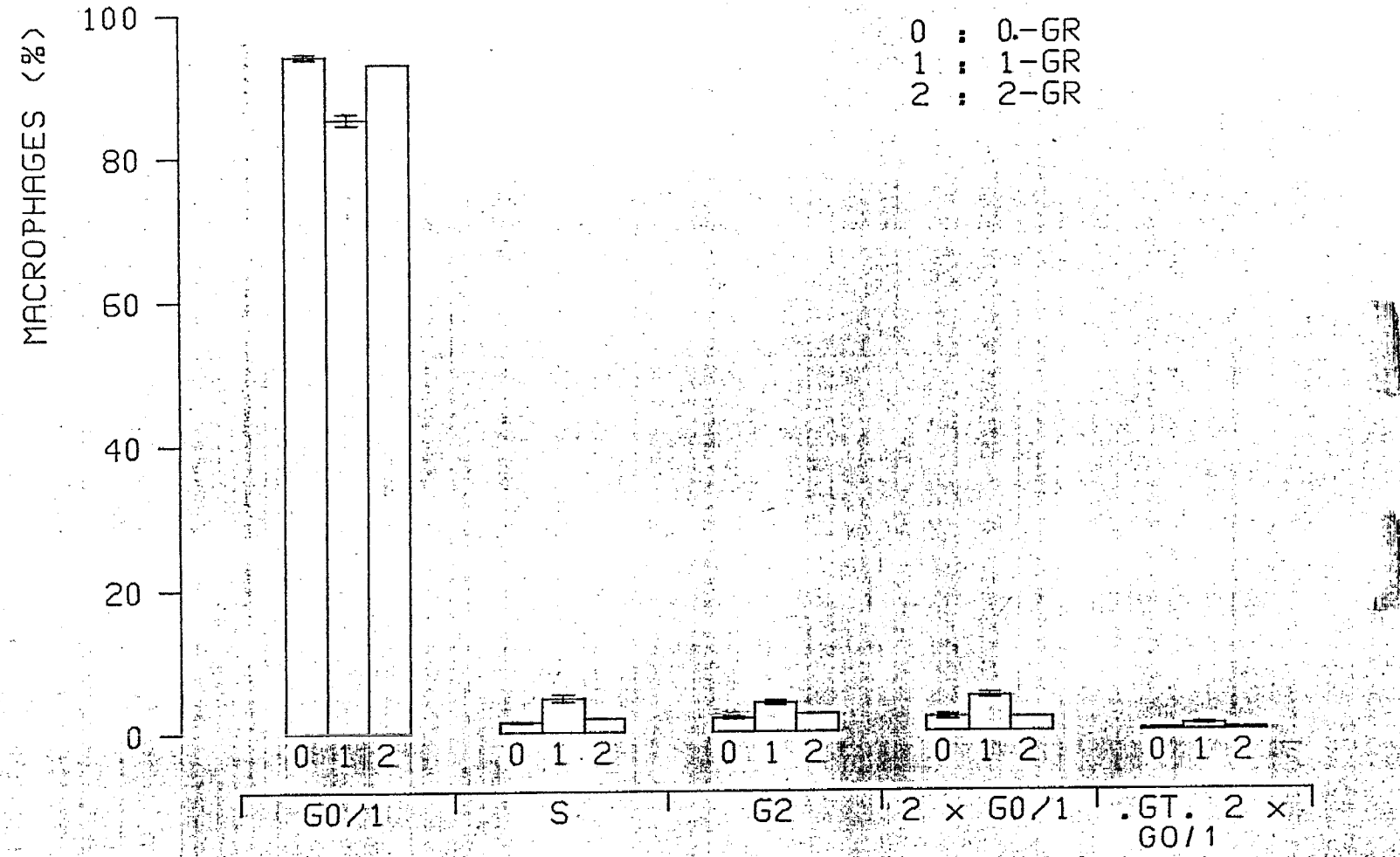
Mean number of phagocytized beads per macrophage and granulocyte, statistical parameters

20290275??

Remarks: for details see BC TABLE 67 and 73

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, HO3354, TH, U23 F162 UXX M, DCM



BC TABLE

Relative number of macrophages in G0/1, S, G2, 2x G0/1 and GT. 2x G0/1, - statistical parameters

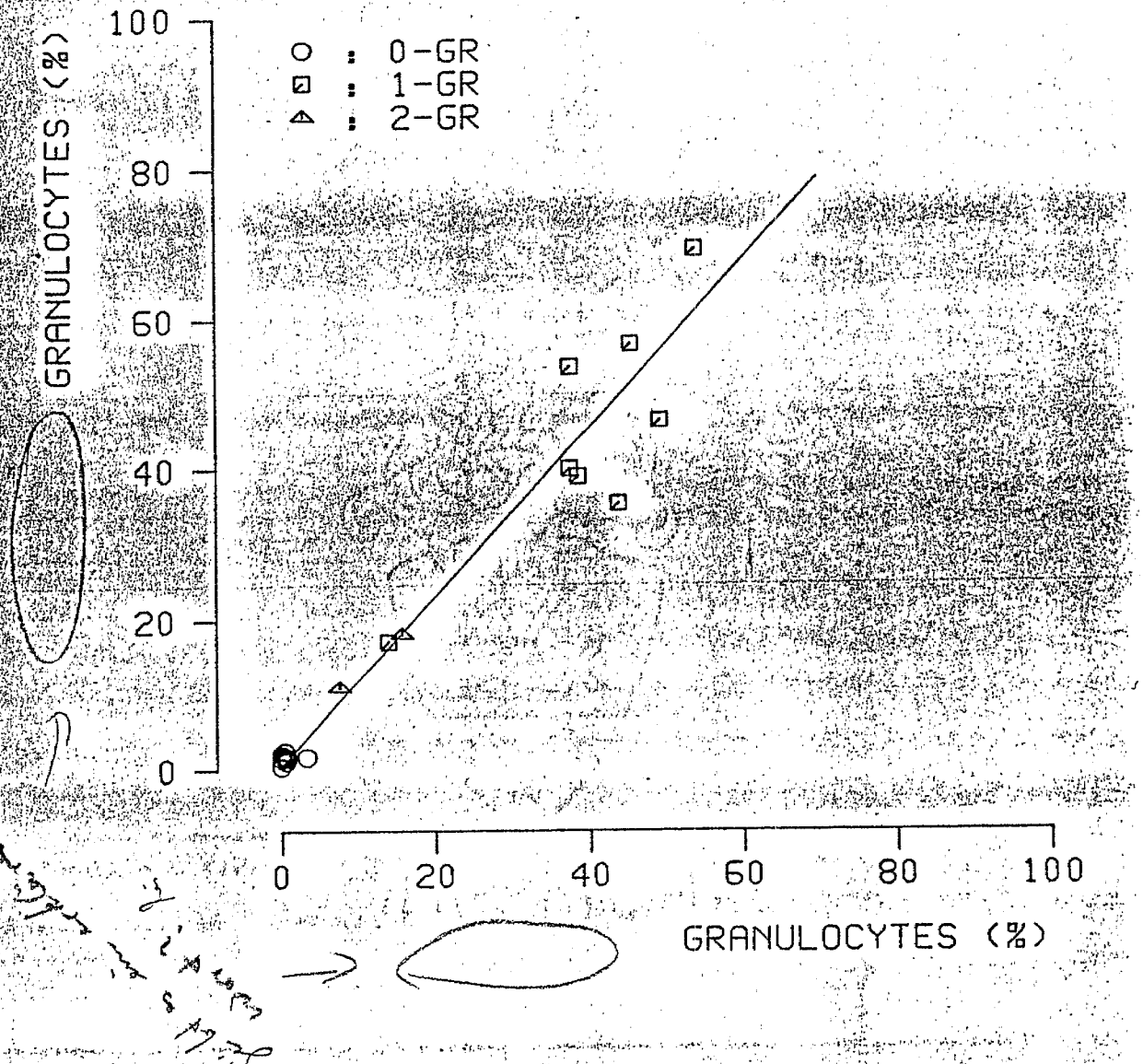
20290275?8

Remarks: for details see BC TABLE 95, 96, 97, 98 and 99

Source: https://www.industrydocuments.ucsf.edu/docs/ynnl0000

8. OKT. 1984

FHO → HGK
BC



A 0500/3057, H03609, V22 F162 V10 DGM

2029027579

2284

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03614, U15 F162 U14, 4C11 P1-7, Ep. 1+3 C Viab SCT

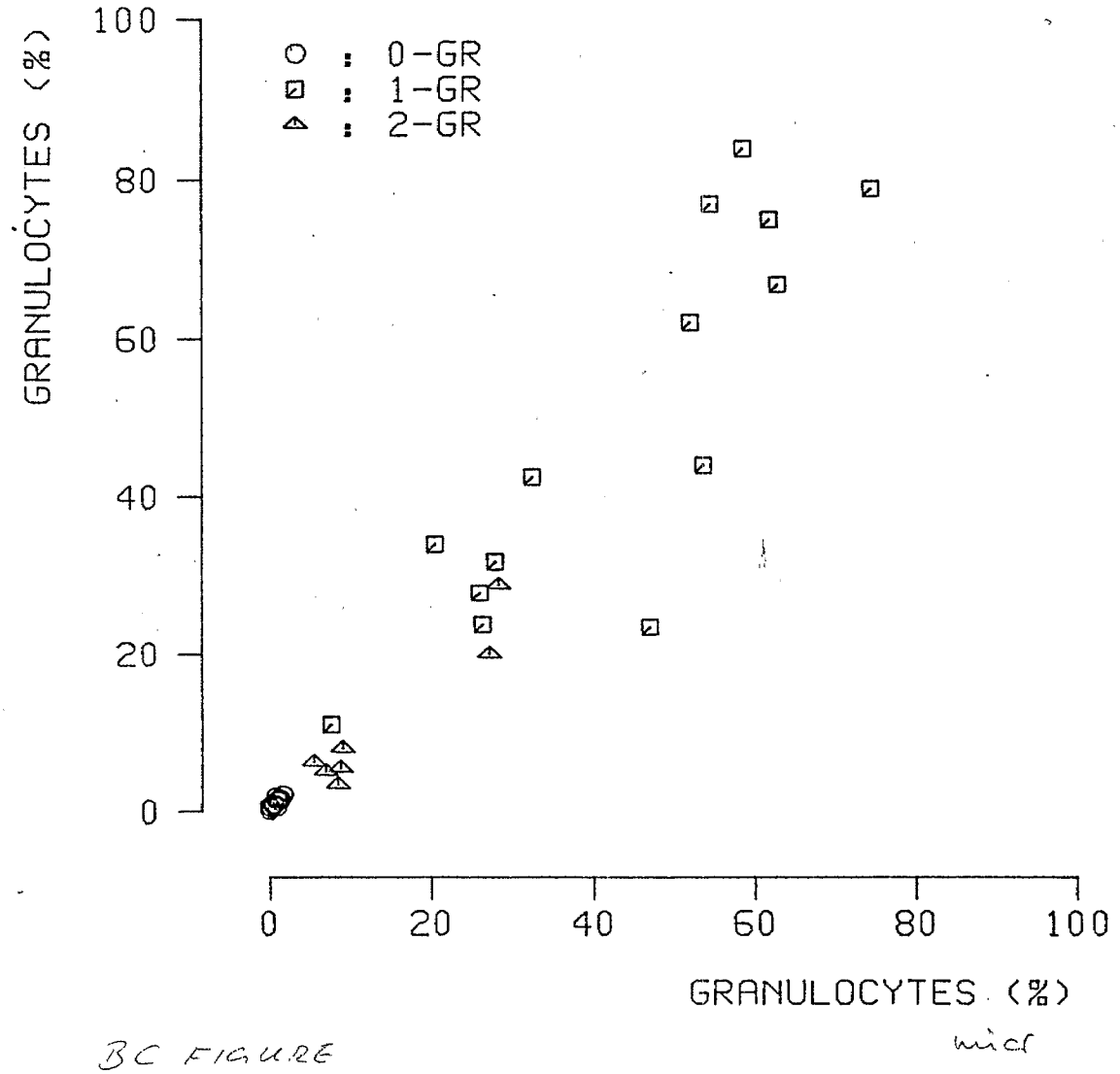
3C FIGURE

Relative number of granulocytes, resuspension medium, cytocentrifuge preparation, versus relative number of granulocytes determined from FWD versus SSC, viability and esterase assay; Pool 1 to 7, Experiment 1 and 3

Remarks: for details see 3C TABLE 2a and 11a

2029027580

Source: https://www.industrydocuments.ucsf.edu/docs/...



A 0500/3057, H03613, U14 F162 U13, V 11 Pool 1-7 day 1 + 3

BC FIGURE

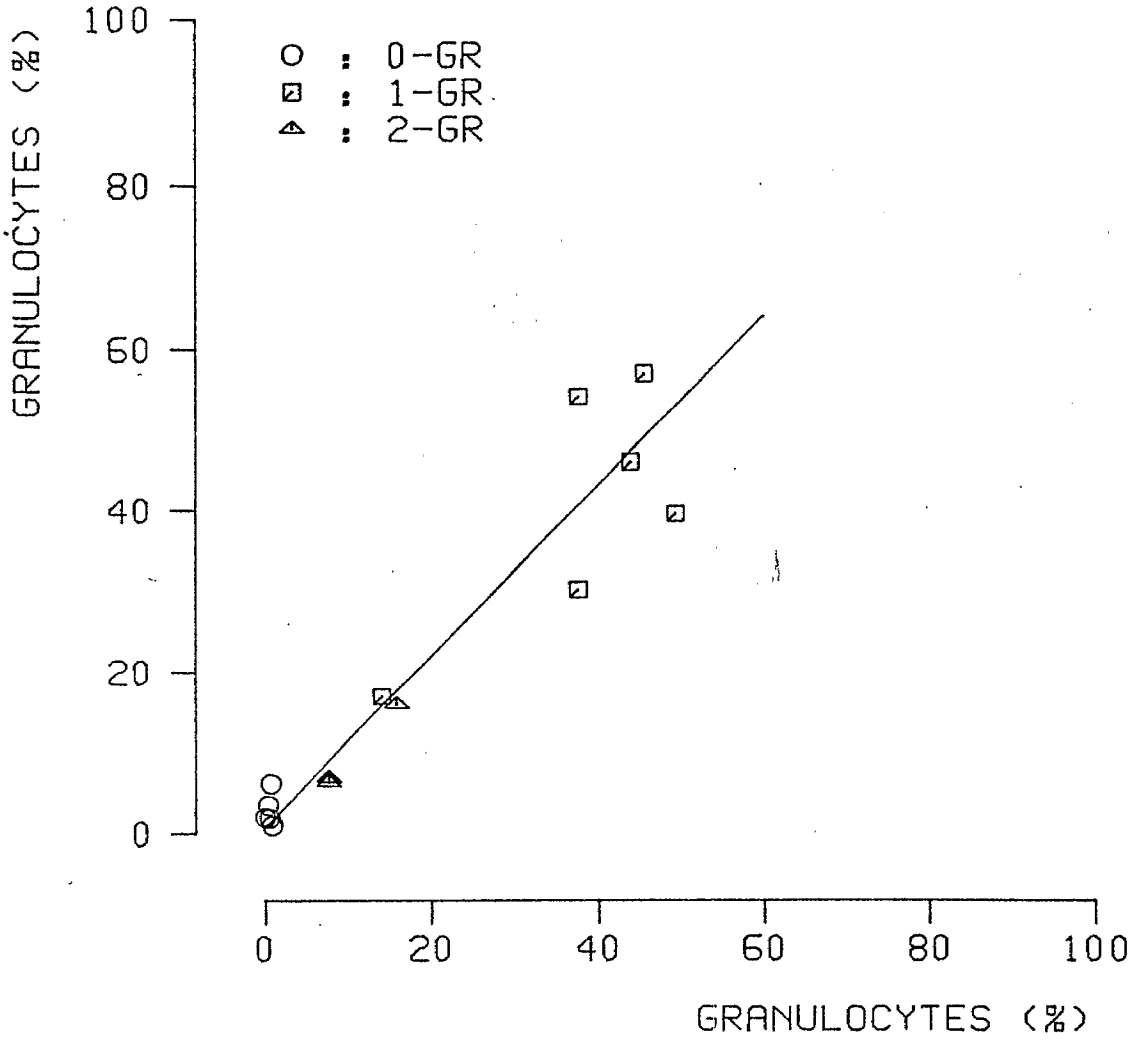
Relative number of granulocytes, resuspension medium, cytocentrifuge preparation, versus relative number of granulocytes determined from esterase activity, viability and esterase assay, Pool 1 to 7, Experiment 1 and 3.

Remarks: for details see BC TABLE 29 and 124

2029027581

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

0500/3057, H03358, TH, V8 F162 UXY USM8C



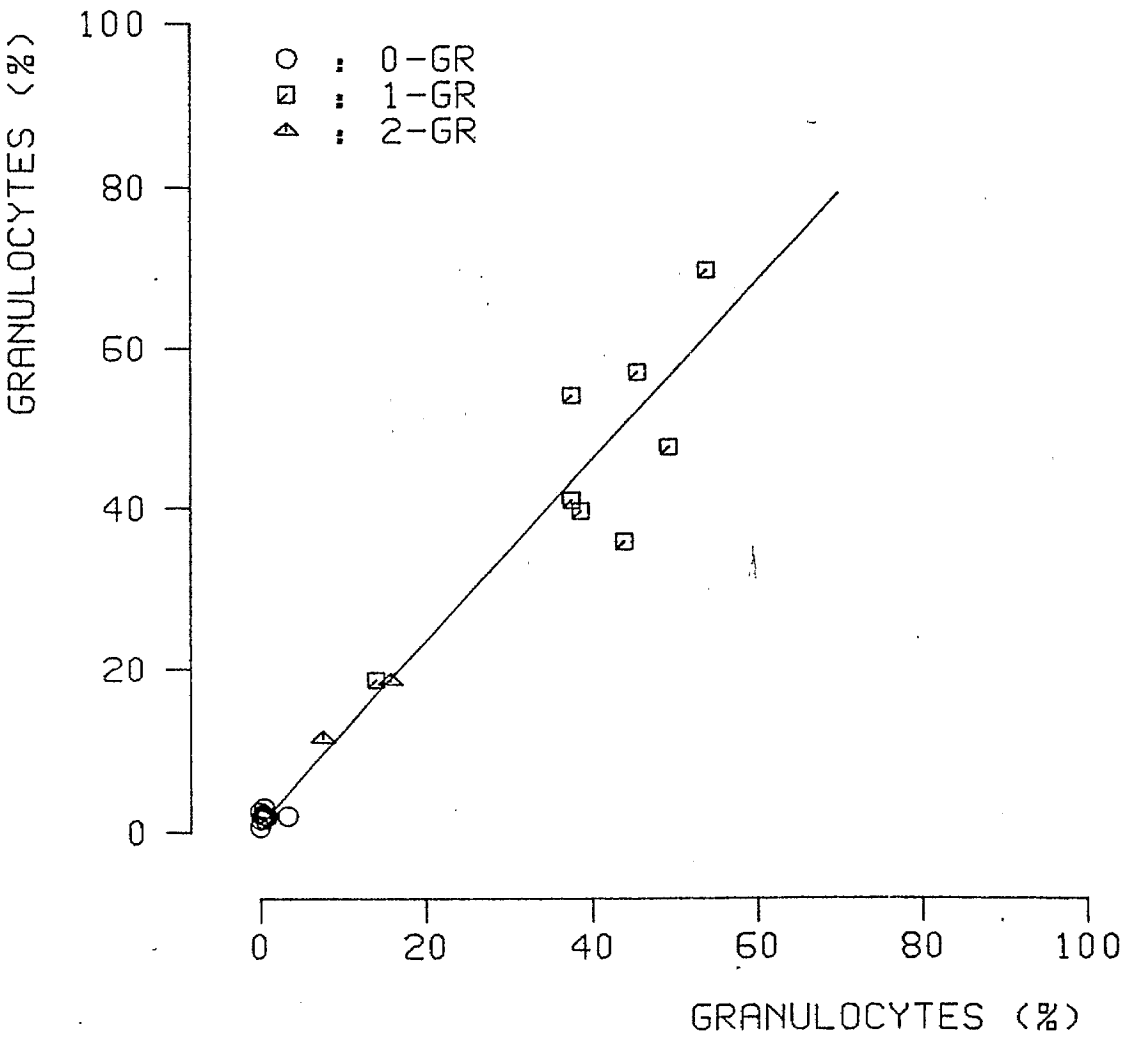
BC FIGURE

Relative number of granulocytes in resuspension medium versus relative number of granulocytes determined from FWD versus AXL, viability and esterase assay, weighted mean of pool 8 to 10, Experiment 1 and 3

Remarks: for disclosure BC TABLE 112 and 115

2029027582

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE

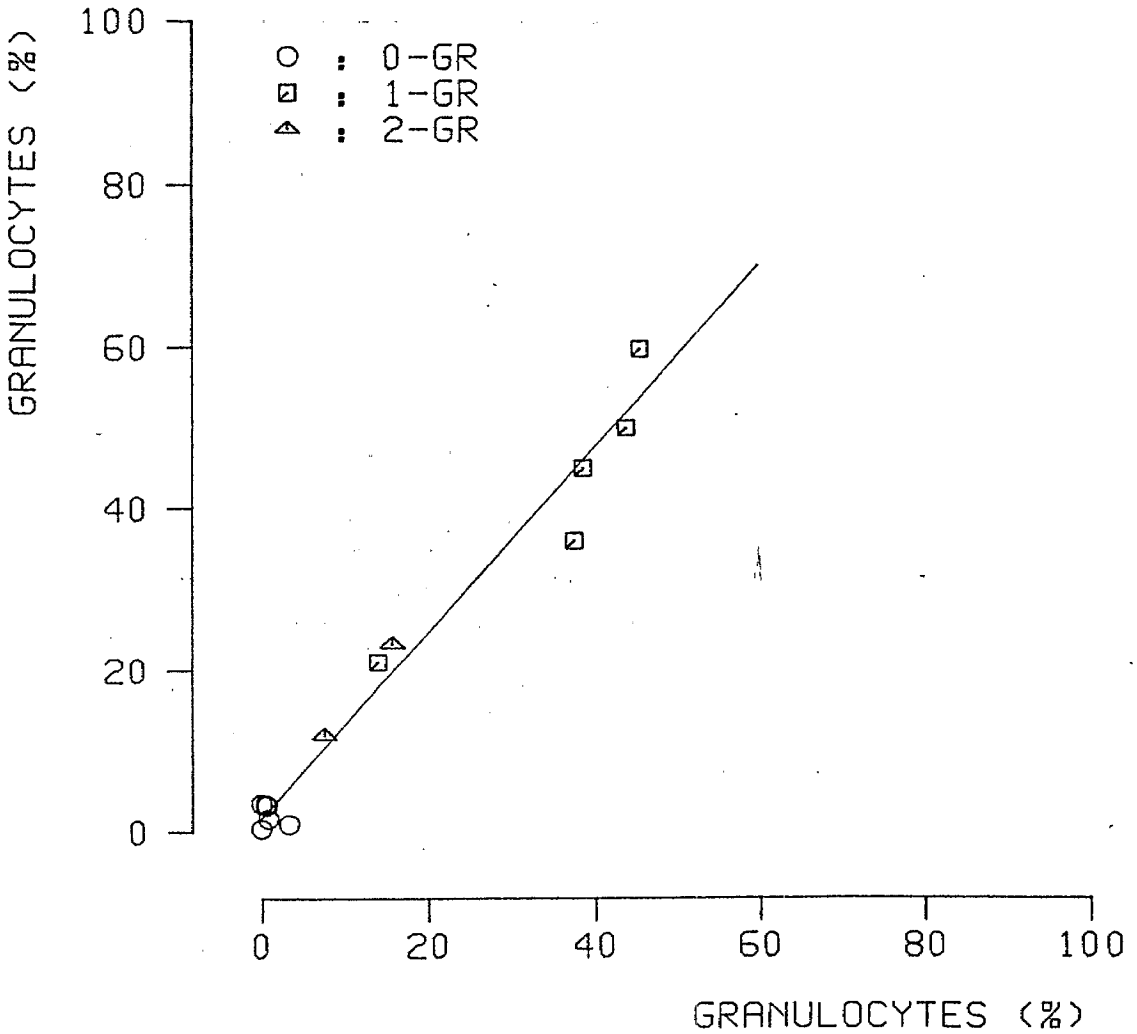
Relative number of granulocytes in suspension medium versus relative number of granulocytes determined from FMD versus AXL, DNA assay, Post 8 to 40, Experiment 1 to 3

Remarks: for details see BC TABLE 112 and 117

A 0500/3057, H03610, U11 F162 U10, II

2029027583

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE

Relative number of granulocytes in resuspension medium versus relative number of granulocytes, Phagocytosis assay, Pool 8 to 10, Experiment 1 to 3

Remarks: for details see BC TABLE 112 and 121

A 0500/30057, H03608, U9 F162 U8, PSN

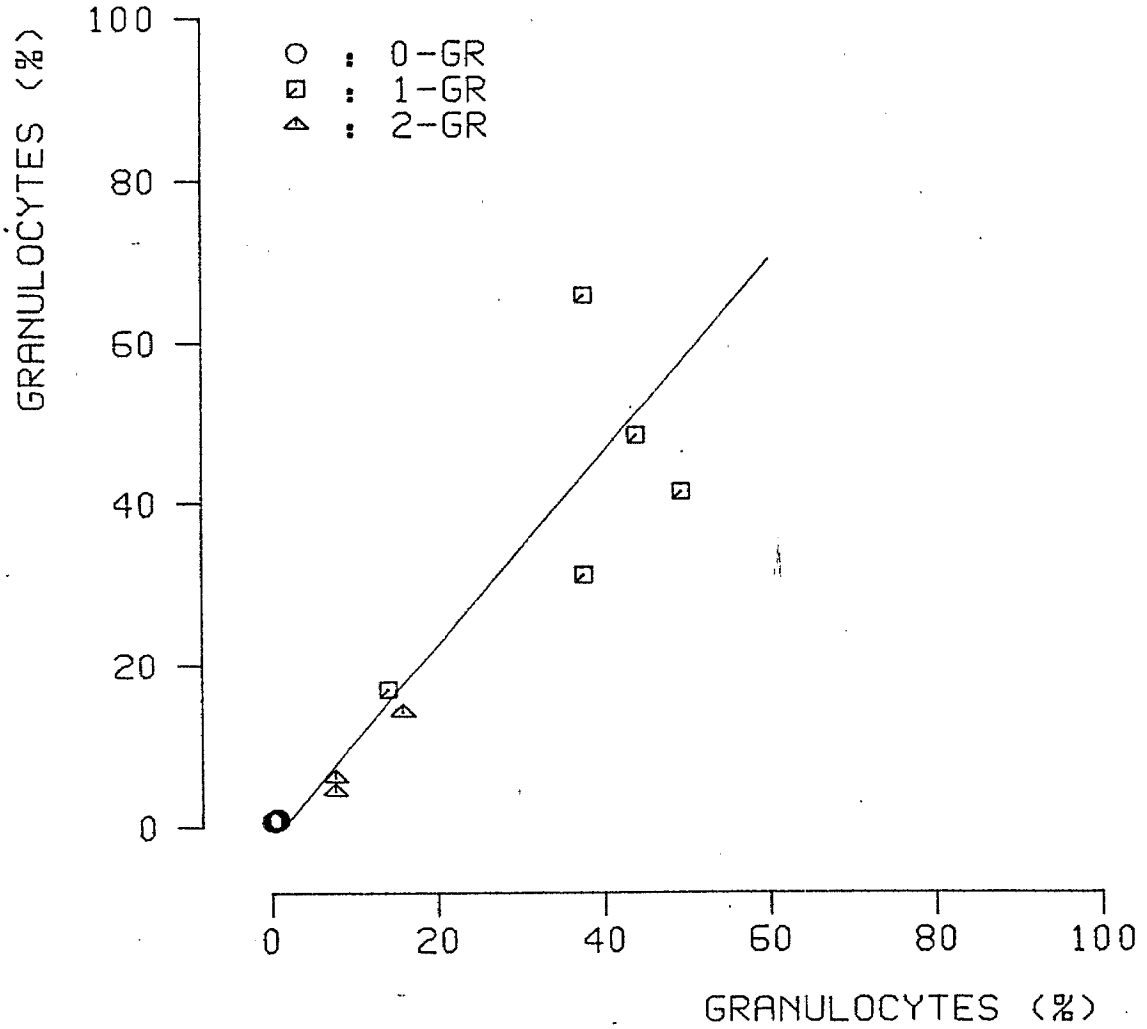
2029027584

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000

A 0500/3057, H03611, U12 F162 U11, K 18

Probe 8, 9, 10

Viab. est.



BC FIGURE

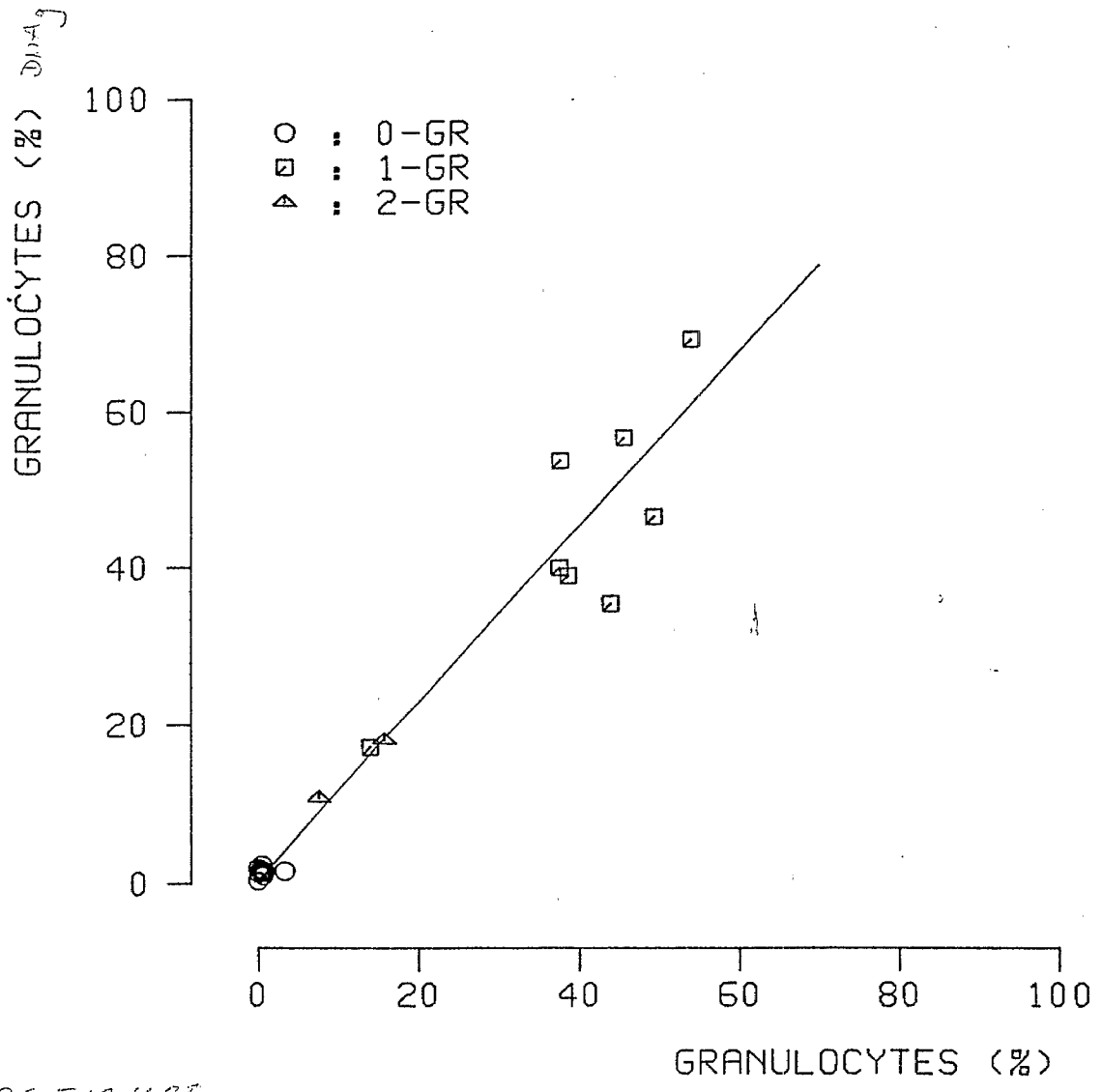
wic F.

Relative number of granulocytes in suspension mediums versus relative number of granulocytes determined from esterase activity, viability and colony assay, Pool 8 to 10, Experiment 1 and 3

Remarks: for details see BC TABLE 112 and 115

2029027585

Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE

Miller

Relative number of granulocytes in suspension culture versus relative number of granulocytes, DNA-gated signals, DNA assay, Pool 8 to 10, Experiment 1 to 3

Remarks: for details see BC TABLE 112 and 127

A 0500/3057, H03609, U22 F162 U10 DE

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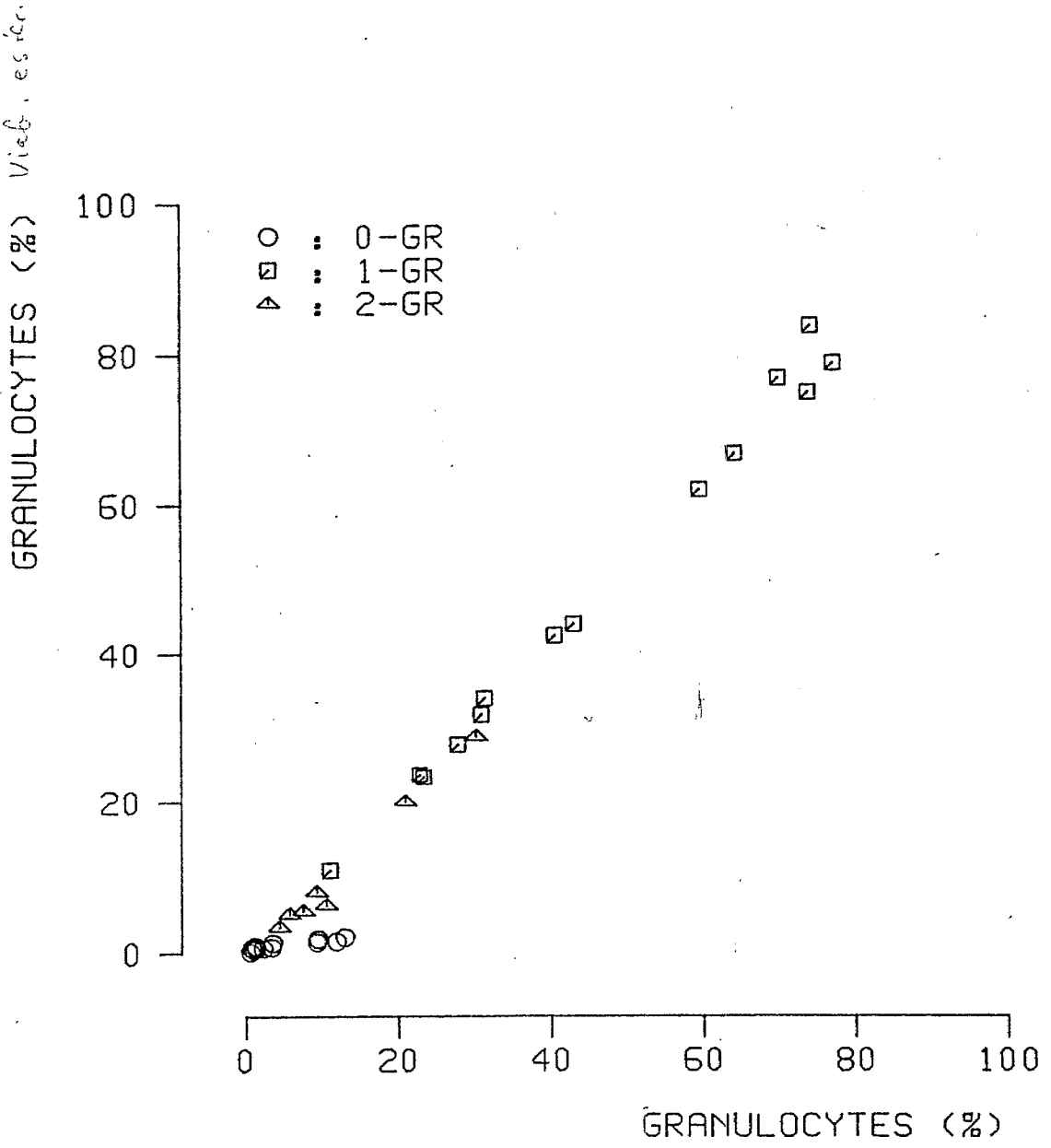
Source: https://www.industrydocuments.ucsf.edu/docs/yndl0000



BC FIGURE

Relative number of granulocytes determined from FluD versus AXL versus relative number of granulocytes determined from esterase activity, viability and esterase assay, Pool 1 to 7, Experiment 1 and 3

Remarks: for details see BC TABLE 114 and 124

A 0500/3057, H03612, V13 F162 V12, V IS

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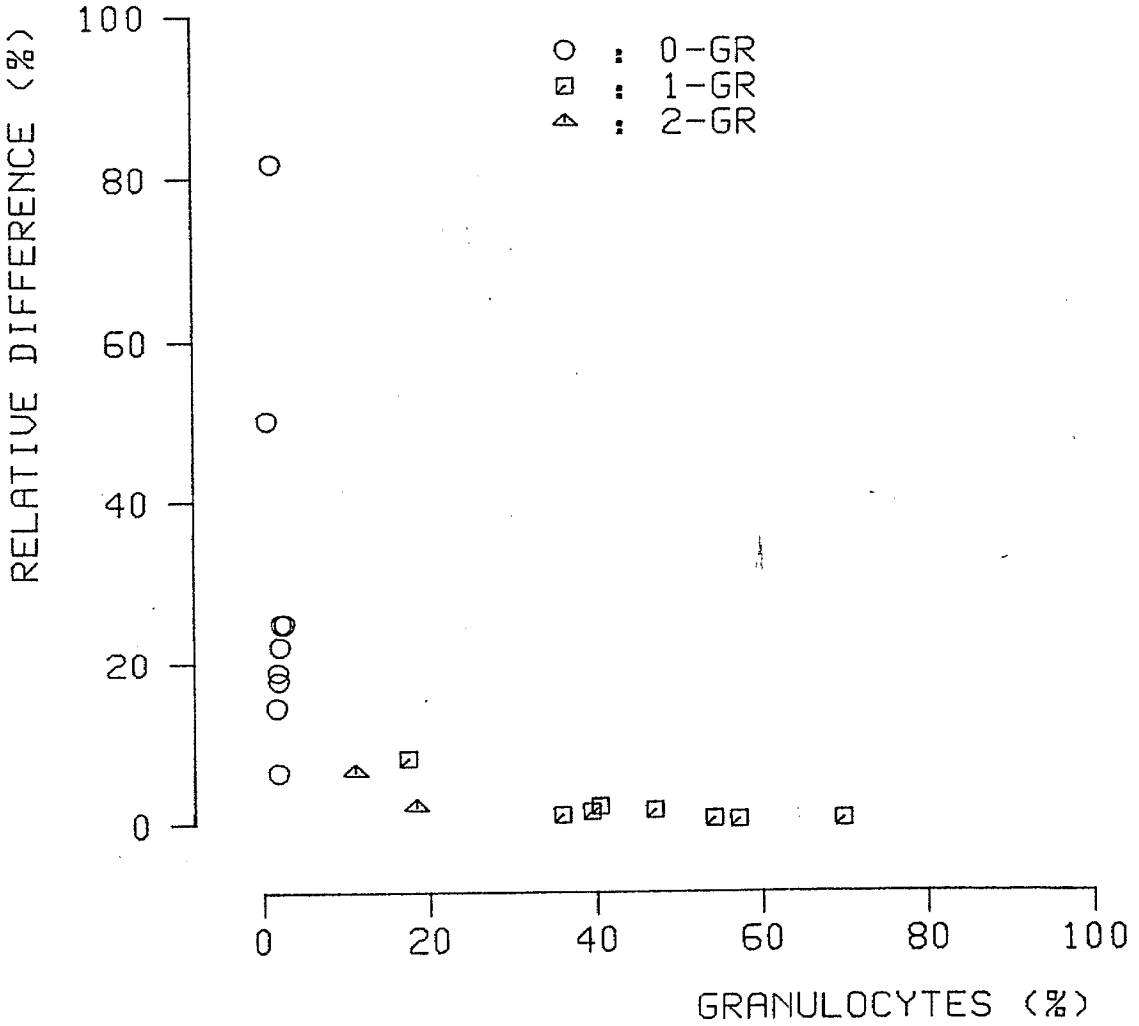

BC FIGURE

Relative number of granulocytes, DNA-gated signals, versus relative difference between the relative number of granulocytes obtained without and with DNA-gating, DNA assay, Pool 8 to 10, Experiment 1 to 3

Remarks: for details see BC TABLE 127 and 130

A 0500/3057, H03615, V16 F162 V15, R 'G

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Epidermal ornithine decarboxylase activity after single dermal application of nicotine to NMRI mice,
Study Director : Dr.rer.nat. R. Mull
Study Codirector: -
issued: 11.Jul.83

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INBIFO study A 0500/3013,
24-day smoke inhalation study on male rats with 2R1 and MW cigarettes and nicotine vapor added to non-tobacco smoke, biochemical and microbiological analyses,
Study Director : Dr.rer.nat. W. Reininghaus
Study Codirector: J. Kühl
issued: report in preparation

INBIFO study A 0500/3018,
91-day smoke inhalation study on male rats with 2R1 cigarette, response of bioassays to subacute inhalation recovery,
Study Director : Dr.rer.nat. W. Reininghaus
Study Codirector: J. Kühl
issued: 17.Mar.80

INBIFO study A 0500/3025,
21-day inhalation study with 2R1 standard reference cigarettes and BVW cigarettes on rats (PT),
Study Director : Dr.rer.nat. W. Reininghaus
Study Codirector: J. Kühl
issued: proposal in preparation

INBIFO study A 0500/3047,
21-day smoke inhalation study with mainstream and sidestream cigarette smoke of standard reference cigarette 2R1 on rats,
Study Director : D. Becker
Study Codirector: Dr.rer.nat. R.-A. Walk
issued: report in preparation

INBIFO study A 0500/3056,
21-day smoke inhalation study with cigarette smoke of standard reference cigarette type 2R1 on Sprague Dawley rats, sister chromatid exchange and chromosome aberrations in peripheral blood lymphocytes,
Study Director : D. Becker
Study Codirector: Dr.rer.nat. R.-A. Walk
issued: 3.Jan.83

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